

Mecanismos fisiológicos en la modulación afectiva de procesos de plasticidad sináptica

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Jorge A. Bergado Rosado

Doctor en Ciencias Biológicas

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SÍNTESIS

Los procesos neuroplásticos son la base funcional que sustenta las capacidades adaptativas del Sistema Nervioso, tanto para las actividades cotidianas como el aprendizaje en cualquiera de sus formas, como para enfrentar situaciones de agresión o daño al sistema, resistir y recuperar funciones. Una extensa base empírica sugiere que estos mecanismos pueden ser afectados por factores afectivos (emoción, motivación). Sin embargo, no se conocen los mecanismos por los cuales se lleva a cabo esa modulación. Utilizando el modelo de reforzamiento conductual de la potenciación sináptica duradera (LTP, del inglés: Long-Term Potentiation) hemos establecido que la modulación afectiva de procesos neuroplásticos se lleva a cabo mediante un sistema neural en el que intervienen estructuras límbicas como la amígdala y el septo medial, así como el locus coeruleus del tronco encefálico. Las proyecciones noradrenérgicas del locus y colinérgicas del septum parecen activar procesos celulares metabólicos (síntesis de proteínas) que conducen al reforzamiento en el tiempo de procesos de potenciación sináptica. Estos mecanismos se afectan por la edad asociada al deterioro cognitivo y parecen tener un carácter universal considerando los resultados obtenidos con el uso de otros modelos en otras regiones cerebrales. El conocimiento ganado sustenta, sobre firmes bases fisiológicas, la relación entre afectividad y cognición, así como entre estos y la restauración neurológica.

INTRODUCCIÓN

Esta tesis esta conformada por un conjunto de artículos encaminados a determinar los mecanismos fisiológicos implicados en la modulación de procesos neuroplásticos por factores afectivos, como el estado emocional y la motivación.

Desde hace mucho tiempo existen evidencias de que esta relación existe. De hecho, a partir de evidencias empíricas, todos los sistemas pedagógicos reconocen y enfatizan la importancia de una buena motivación para lograr un aprendizaje más efectivo. De la misma forma, todos los sistemas de condicionamiento animal con fines científicos, de servicio, detectivescos o de espectáculo, reconocen que todo intento de enseñar algo a un animal será infructuoso si no existe una motivación adecuada según la etología de la especie.

El surgimiento de la Neurología Restaurativa en años recientes, ha puesto en evidencia que también el éxito de los progresos en la recuperación funcional, espontáneos o inducidos mediante un programa de rehabilitación, son influenciados fuertemente por el estado afectivo de los pacientes.

A pesar de la importancia que tiene comprender cuales son las bases fisiológicas de estas interacciones entre afectividad y neuroplasticidad, poco se ha avanzado hasta hoy en el tema. Nosotros hemos empleado un modelo de neuroplasticidad: la potenciación sináptica duradera y su reforzamiento por factores motivacionales (reforzamiento conductual) para estudiar estos procesos y establecer sus bases funcionales, tanto al nivel sistémico con la identificación de qué estructuras están implicadas en esos mecanismos, como al nivel celular, identificando neurotransmisores, receptores y procesos metabólicos involucrados.

Los resultados obtenidos son de absoluta novedad científica y contribuyen a la comprensión de las bases fisiológicas de la relación entre los procesos neuroplásticos que sustentan funciones de memoria, adaptación y recuperación del Sistema Nervioso; y el estado afectivo, determinado por el Sistema Límbico.

Estos resultados ayudan a interpretar efectos conocidos, como la relación entre el aprendizaje y la motivación; o la relación entre el estado afectivo y el resultado de la rehabilitación. El conocimiento de las bases neuroquímicas puede también sustentar el desarrollo de una farmacoterapia de apoyo a estos procesos en el marco de tratamientos neurorestaurativos.

Los resultados son la culminación de un esfuerzo conjunto entre el Departamento de Neurofisiología Experimental –que dirijo en el CIREN- y el Departamento de Neurofisiología del Instituto Leibniz de Neurobiología de Magdeburgo, Alemania – dirigido por la Prof. J.U. Frey. La mayor parte de los trabajos fueron realizados en Cuba. Los que se hicieron en Alemania fueron realizados personalmente por mí durante estancias de trabajo en ese país. Todos los trabajos fueron planificados, organizados y dirigidos por mí, con la colaboración de la Prof. Frey. De la misma forma, todos los artículos que conforman la tesis fueron escritos por mí.

MARCO TEORICO

Los antecedentes científicos necesarios para facilitar la comprensión de esta tesis y su ubicación dentro del contexto del problema que se aborda, aparecen recogidos en dos artículos de revisión publicados por el optante.

El primero de ellos presenta una revisión amplia del tema de la neuroplasticidad en todas sus dimensiones y facetas, con particular énfasis en la plasticidad sináptica, que es el modelo sobre el cual se trabaja en los artículos que conforman los resultados de la tesis.

Aunque fue publicado en el año 2000 considero que conserva, en lo esencial, actualidad y vigencia.

El segundo aborda el tema de la plasticidad sináptica y su relación con el deterioro funcional que acompaña al envejecimiento cerebral y ofrece un marco referencial adecuado para los tres artículos donde se abordan problemas relacionados con el envejecimiento.

Mecanismos celulares de la neuroplasticidad

J.A. Bergado-Rosado, W. Almaguer-Melian

CELLULAR MECHANISMS OF NEUROPLASTICITY

Summary. Objective. To present a unified vision of the principal known mechanisms of neuroplasticity, emphasizing their universality. Development. The concept of the central nervous system as an immutable entity has been considerably modified during the second half of the 20th century. Neuroplasticity, that is the ability of the brain regarding change and repair is expressed in different ways, from functional modifications of existing structures to the formation, by growth and proliferation, of new structures and neurons. This study considers the molecular and cellular mechanisms of neuroplastic phenomena and classifies them into two main groups: plasticity due to growth, including the mechanisms of axonal regeneration, collateralization and reactive synaptogenesis; and functional plasticity, which includes changes in the efficacy of synaptic transmission such as long-term potentiation and the activation of silent synapses. We also describe some of the relations of neuroplastic phenomena with disease of the central nervous system, together with examples of physiological, physical and pharmacological factors which may be used in future as therapeutic tools to stimulate and modulate neuroplasticity. Conclusion. Neuroplastic mechanisms show a high degree of phylogenetic and ontogenetic conservation. They are important both in the genesis of disorders and disease of the nervous system and for its repair after different types of damage and trauma. Modulation of neuroplastic mechanisms by physical and chemical agents would appear to be one of the most powerful therapeutic tools of restorative neurology. [REV NEUROL 2000; 31: 1074-95] [<http://www.revneurolog.com/3111/j111074.pdf>]

Key words. Collateralization. Complex environment. Cortical plasticity. Gangliosides. Long term potentiation. Neurogenesis. Neuroplasticity. Neurotrophic factors. Orotic acid. Pathology. Physical exercise. Regeneration. Steroids. Synaptogenesis.

LA NEUROPLASTICIDAD. CONCEPTO

Durante muchos años se consideró al sistema nervioso central (SNC) como una estructura funcionalmente inmutable y anatómicamente estática. El dogma 'no nuevas neuronas', insidiosamente extendido, significó también en todo ese tiempo: no nuevas conexiones. El sistema, una vez concluido su desarrollo embrionario, era una entidad terminada y definitiva, mutable sólo por lesión o degeneración e irreparable por su propia naturaleza. Ramón y Cajal escribió en su obra *Degeneración y regeneración en el sistema nervioso*: '...la especialización funcional del cerebro impone a las neuronas dos grandes lagunas: incapacidad de proliferación e irreversibilidad de la diferenciación intraprotoplasmática. Es por esta razón que, una vez terminado el desarrollo, las fuentes de crecimiento y regeneración de los axones y dendritas se secan irrevocablemente. En los cerebros adultos las vías nerviosas son algo fijo, terminado, inmutable. Todo puede morir, nada puede regenerarse'. Pero Don Santiago no sería el Maestro si no hubiera escrito al final del párrafo: 'Corresponde a la ciencia del futuro cambiar, si es posible, este cruel decreto' [1].

En los últimos 40 años, el dictamen ha cambiado radicalmente. El rígido esquema de circuitos invariables, tanto en el número de sus unidades como en las conexiones entre ellas, ha sido sustituido progresivamente por un sistema en que la modificación dinámica de sus propiedades, en respuesta a cambios en su ambiente y sus ingresos, constituyen la noción fundamental para comprender sus extraordinarias propiedades. Esta nueva visión se sustenta en el concepto de la neuroplasticidad y es hoy un elemento unificador esencial para comprender procesos tan aparentemente diferentes como el aprendizaje y la recuperación de funciones tras una lesión.

De acuerdo con esta concepción, el SNC es un producto nunca terminado, es el resultado, siempre cambiante y cambiable, de la interacción de factores genéticos y epigenéticos.

Tal vez la importancia de la concepción neuroplástica del SNC radique en la nueva mentalidad que impregna actualmente el amplio espectro de las Neurociencias, tanto experimentales como aplicadas. Del fatalismo del 'nada puede hacerse' se transita hoy aceleradamente hacia la búsqueda y ensayo constante de nuevas formas de estimular los cambios plásticos que permitan la restauración de funciones alteradas por traumas, accidentes vasculares o enfermedades degenerativas (Tabla I), no sólo por la sustitución, sino buscando también la recuperación de las áreas dañadas [2]. Comienza a constituirse una Neurología Restaurativa que ha de ser, sin duda, la Neurología del nuevo siglo.

MECANISMOS DE LA NEUROPLASTICIDAD

Los mecanismos de la neuroplasticidad son muy diversos y pueden abarcar desde modificaciones morfológicas extensas, como las que se observan en la regeneración de axones y formación de nuevas sinapsis, hasta sutiles cambios moleculares que alteran la respuesta celular a los neurotransmisores [3]. En esta revisión enfatizaremos los mecanismos neuronales de la plasticidad, aunque no debe perderse la perspectiva de que cada neurona del SNC es sustentada por una unidad trófica formada por otras neuronas, células de la glía, vasos sanguíneos y moléculas de la matriz celular [4], y a ellas nos referiremos en aquellos casos en que su papel está mejor establecido. Hemos omitido algunos mecanismos considerados clásicamente como neuroplásticos, por ejemplo la diasquisis, porque no lo son. La diasquisis resulta del enmascaramiento de funciones por un desequilibrio transitorio entre excitación e inhibición [2] y nada tiene que ver con la neuroplasticidad.

La regeneración, formación de colaterales axónicas y de nuevas sinapsis, constituye la base de la reorganización y recuperación de funciones perdidas por daño a las neuronas. De sus características esenciales nos ocuparemos brevemente en la primera parte.

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Centro Internacional de Restauración Neurológica (CIREN). La Habana, Cuba.

Correspondencia: Dr. Jorge A. Bergado Rosado. Centro Internacional de Restauración Neurológica (CIREN). Ave. 25, #15.805. Playa. CP 12100 Ciudad de La Habana, Cuba. Fax: (537) 332420. E-mail: bergado@neubas.sld.cu

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Tabla 1. La neuroplasticidad está vinculada a las enfermedades más importantes que afectan al sistema nervioso. Los procesos neuroplásticos son responsables, en buena medida, de la recuperación de las funciones en los pacientes que sufren las consecuencias de trastornos cerebrovasculares. Procesos de neuroplasticidad aberrantes están implicados en la progresión y, tal vez, en la propia génesis de muchas formas de epilepsia. En las enfermedades neurodegenerativas, como la demencia tipo Alzheimer y el morbus Parkinson, el agotamiento de las capacidades neuroplásticas podría ser el responsable de algunas de las consecuencias más invalidantes de estos trastornos. (Datos tomados de Price DL. Nature 1999; 399 (Suppl)).

Enfermedad	N.º de casos en Estados Unidos
Enfermedad cerebrovascular	1,5 millones de casos nuevos por año
Epilepsia	2,5 millones de casos
Enfermedad de Alzheimer	5 millones de casos
Enfermedad de Parkinson	500.000 casos
Esclerosis múltiple	300.000 casos

La modificación de las capacidades funcionales de sinapsis existentes puede contribuir a la compensación funcional a expensas de sinapsis poco activas o silentes que están en la base de lo que se ha dado en llamar plasticidad conductual. Estos cambios de conectividad sináptica también se consideran hoy en día fundamentos fisiológicos de los procesos de aprendizaje y memoria, como fuera anticipado por Donald Hebb y Hansjürgen Matthies [5-7]. Revisaremos sus mecanismos fundamentales en el hipocampo y su expresión en otras áreas, en particular en los procesos de maduración funcional de la corteza cerebral.

Plasticidad por crecimiento

Regeneración axonal

Desde el siglo pasado se conoce que los axones del sistema nervioso periférico pueden regenerarse por crecimiento a partir del cabo proximal. Ello no ocurre en el SNC de los mamíferos, aunque sí en vertebrados más primitivos [8]. Al parecer, la ausencia de regeneración no se debe a una incapacidad esencial de las neuronas centrales, por cuanto cerca de las neuronas dañadas se encuentran signos de regeneración abortiva, llamada gemación (*sprouting*) regenerativa [9]. Existen evidencias de que la mielina central y los oligodendrocitos que la producen contienen sustancias que inhiben la regeneración axonal [10,11].

La regeneración axonal sería útil sobre todo para la reparación de tractos de fibras largas, como los del nervio óptico –que no es un nervio periférico– o los que actúan en la médula espinal. Actualmente se experimentan nuevas estrategias para promover su regeneración: puentes de nervio periférico, factores tróficos o anticuerpos monoclonales diseñados para bloquear los factores inhibidores gliales (Ver resumen en [8]).

Colateralización o gemación colateral

La ausencia de regeneración axonal no significa que no ocurran cambios regenerativos ante la pérdida de inervación. Estos cambios, además, pueden tener profundas influencias en la recuperación de funciones perdidas.

Una forma bien estudiada es la llamada colateralización o gemación (*sprouting*) colateral. La colateralización se diferencia de la regeneración en que el crecimiento ocurre a expensas de axones sanos, que pueden provenir de neuronas no afectadas por la lesión o de ramas colaterales de los mismos axones dañados que

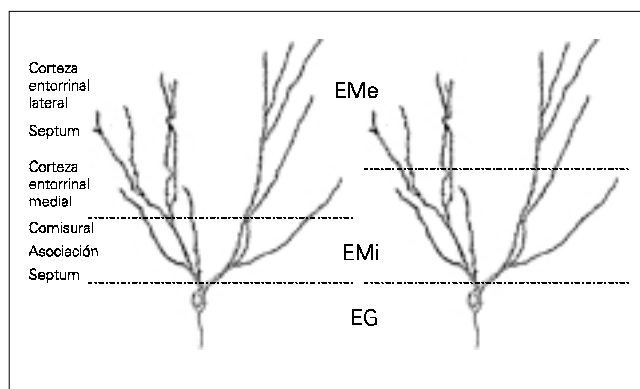


Figura 1. Modelo corteza entorrinal-giro dentado para el estudio de mecanismos neuroplásticos. A la izquierda se menciona el origen de las aferencias y se indica la zona en que se distribuyen en el estrato molecular del giro dentado sobre una neurona granular. a) Patrón normal. Los ingresos al giro dentado siguen un patrón de distribución ordenado. El estrato molecular externo recibe a las fibras de la vía perforante lateral en su región más distal y de la corteza entorrinal medial en su zona proximal. Al estrato molecular interno llegan fibras comisurales (del hipocampo contralateral) y asociativas (del mismo hipocampo). Las fibras de origen septal se distribuyen en ambos estratos. b) Después de lesionar la corteza entorrinal. La lesión de la corteza entorrinal destruye el ingreso al estrato molecular externo. La colateralización y sinaptogénesis reactiva conducen a la expansión del estrato molecular interno. EG: estrato granuloso; EMI: estrato molecular interno; EMe: estrato molecular externo.

la lesión no llegó a afectar. Aunque suele distinguirse esta segunda variante con el nombre de efecto de poda (*pruning*), los mecanismos de ambas formas de crecimiento axonal colateral parecen ser muy similares [9] a pesar de agentes diferentes los inician.

La colateralización puede ocurrir a partir de axones del mismo tipo de los dañados (colateralización homotípica) o de otro tipo (colateralización heterotípica). El proceso de colateralización normalmente concluye con la formación de nuevas sinapsis que reemplazan a las que se han perdido por la degeneración retrógrada de los axones destruidos. Este proceso se ha llamado sinaptogénesis reactiva, para distinguirlo de la sinaptogénesis que normalmente sucede en las etapas intermedias del desarrollo embrionario; no obstante, no parece existir diferencia alguna entre los mecanismos de una y otra.

La mayor parte de los estudios experimentales sobre los mecanismos de colateralización se ha realizado utilizando el modelo de lesión de la vía perforante que proyecta de la corteza entorrinal al giro dentado del hipocampo. Esta proyección es glutamatérgica y las fibras que la integran se distribuyen en el campo dendrítico de las neuronas granulares del giro dentado siguiendo un patrón regular y ordenado (Fig. 1). Las regiones más distales del campo dendrítico reciben las fibras que se originan en la parte lateral de la corteza entorrinal, mientras que aquellas que proceden de la porción medial de la corteza entorrinal terminan sobre el tercio medio del campo dendrítico. Estas dos subregiones se corresponden anatómicamente con el llamado estrato molecular externo. El tercio interno del campo dendrítico, correspondiente al estrato molecular interno, recibe fibras colinérgicas y gabérgicas del septum, noradrenérgicas y serotoninérgicas desde núcleos de la formación reticular, así como fibras asociativas y comisurales de otras regiones del hipocampo ipsi y contralateral, respectivamente. La lesión de la corteza entorrinal provoca una reducción significativa del estrato molecular externo que se acompaña de la expansión del estrato molecular interno.

En este modelo, los primeros signos de crecimiento de cola-

terales axónicas aparecen seis días después de la lesión y son muy intensos en la segunda y tercera semana [12]. Los agentes que inician este crecimiento no se conocen con precisión y se han formulado varias hipótesis, no alternativas, que podrían desencadenar procesos de colateralización:

- Especializaciones post-sinápticas vacantes. Los axones sobrevivientes tras la degeneración de los cabos distales de las fibras transectadas detectan la presencia de ‘plazas vacantes’ y ello estimula su crecimiento.
- Ausencia de inhibición competitiva. La densidad de innervación de una neurona podría estar controlada por señales inhibitorias que limitan el crecimiento axonal. La pérdida de una cantidad sustancial de terminales eliminaría este freno al crecimiento axonal.
- Cambios en la actividad sináptica. La pérdida de aferentes altera la actividad de las neuronas. Ello, a su vez, podría conducir a la liberación de factores tróficos del crecimiento axonal.
- Presencia de terminales en degeneración. Las terminales que degeneran liberan sustancias que estimulan la colateralización.
- Las células gliales que fagocitan los axones degenerados liberan factores tróficos que estimulan el crecimiento colateral [9].

La acción cooperativa de varios de los factores antes enunciados contribuye a crear lo que se ha dado en llamar un ambiente promotor de crecimiento que pone en marcha la gemación y extensión de los axones o ramas intactas.

La importancia de algunos factores para la colateralización se ha puesto de manifiesto en hallazgos recientes. Así, por ejemplo, la activación de receptores de tipo N-metil-D-aspartato (NMDA) en las neuronas post-sinápticas parece necesaria para promover el crecimiento axonal. El bloqueo de estos receptores impide la inducción de la proteína GAP-43 y el crecimiento colateral [13,14].

La fosfoproteína GAP-43 (*Growth Associated Protein*), también llamada F-1 o B-50, se relaciona con las terminales axónicas y podría tener alguna función en la transmisión sináptica normal, pero su expresión se incrementa dramáticamente en axones que se elongan [15,16]. Los niveles más altos de GAP-43 se encuentran siempre en neuronas que colateralizan [17] y se considera, por lo tanto, como un marcador específico de axones en crecimiento. Estudios con microscopía electrónica indican que GAP-43 se transporta predominantemente hacia los axones que sufren remodelación. Se desconoce la función específica de esta proteína en el proceso de crecimiento axonal y cómo se modifica su función al ser fosforilada por la proteinocinasa C. Por otra parte, GAP-43 no parece ser la única proteína sináptica vinculada al crecimiento axonal. Existen evidencias de que la proteína SNAP-25 también participa en el crecimiento axonal colateral [21] y otras como la sinapsina I, cuyo gen se activa sólo en el momento de establecer el contacto sináptico [22,23]. A pesar de esto, GAP-43 es un marcador esencial para el estudio experimental de la colateralización.

Otro aspecto que parece importante para la iniciación y desarrollo de la colateralización son las interacciones gliales. Como mencionamos anteriormente, las células gliales son necesarias para eliminar las terminales axónicas degeneradas. Existe una secuencia de activación glial que involucra primero a la microglía y luego incluye a los astrocitos. La respuesta microglial es evidente ya a las 24 horas de la lesión. La activación astrocítica únicamente es evidente dos días después, mientras que los primeros signos de colateralización se observan a los tres días. La activación microglial se mantiene durante cuatro semanas después de lesión y la astrocítica durante tres semanas [24]. Esta reacción no

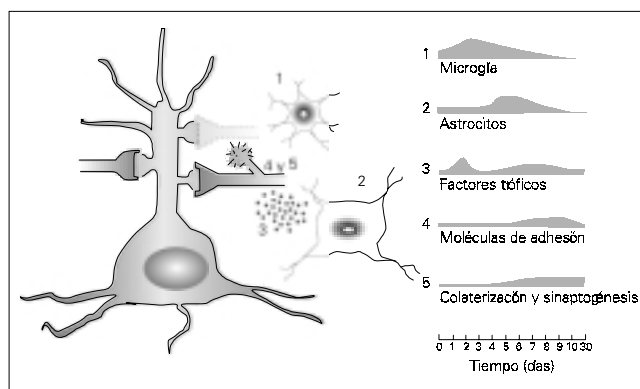


Figura 2. Mecanismos de colateralización. (1) La presencia de células dañadas atrae a la microglía, responsable principal de eliminar el detrito celular. Más tarde (2), se produce una reacción astrocítica que libera factores tróficos (3) estimuladores del crecimiento axonal colateral, el cual se acompaña de expresión de moléculas de adhesión (4) que guían y estabilizan las neuritas en crecimiento. El proceso culmina con la formación de nuevos contactos sinápticos (5).

incluye, sin embargo, a los linfocitos T [25] u otros componentes de la cohorte inmune (Fig. 2).

La función fagocítica corresponde predominantemente a la microglía, mientras que los astrocitos parecen responsables de la producción de factores tróficos que estimulan el crecimiento axonal. La lesión induce en los astrocitos la expresión de formas truncadas del receptor tirosinocinasa B (TrkB) que, se piensa, actúan como ‘presentadoras’ de neurotrofinas a los axones en crecimiento [26], y de moléculas de adhesión celular, como la NCAM (*Neural Cell Adhesion Molecule*) [27-29] y la tenascina C [30], que participan en la guía de los axones en crecimiento hacia sus blancos y forman verdaderas ‘pistas’ de elongación [31-33].

El proceso de colateralización, seguido de la formación de nuevos contactos sinápticos, puede desempeñar un papel muy importante en la recuperación de funciones perdidas como consecuencia de lesión o en el retraso de la aparición de trastornos manifiestos en las enfermedades neurodegenerativas.

Si la colateralización es homotípica, su valor restaurativo resulta evidente, pero aun una colateralización heterotípica puede ser beneficiosa. Primero, porque la presencia de fibras aferentes es necesaria para el mantenimiento dendrítico; y, por otra parte, la colateralización heterotípica puede contribuir al equilibrio excitación-inhibición y con ello a una restauración parcial de la función neural.

En el modelo de lesión entorrinal antes citado, se ha descrito colateralización de fibras asociativas y comisurales (homotípica), así como de fibras colinérgicas septales o noradrenérgicas (heterotípica) [34,35]. Esto se expresa morfológicamente en la expansión del estrato molecular interno [36] y funcionalmente en la recuperación de funciones de memoria afectadas por la lesión [37].

También se han demostrado procesos de colateralización en otros modelos de deservación hipocampal, como la lesión de fimbria-fórnix que interrumpe la aferencia colinérgica procedente del septum. En este modelo se produce un crecimiento heterotípico, principalmente noradrenérgico, y un crecimiento homotípico [38], reacción que, aunque está bien descrita en roedores, no parece tan importante en los primates [39].

Sinaptogénesis reactiva

El brote y extensión de nuevas ramas axónicas serían totalmente inútiles si no culminasen con la formación de nuevos contactos

sinápticos. La sinaptogénesis reactiva es parte insoluble de un solo proceso que comienza con la colateralización y concluye con la formación de nuevos contactos funcionales.

En el modelo de lesión entorrinal, se ha demostrado que la desnervación inicial conduce a la pérdida de más del 85% de las sinapsis en el estrato molecular, que es seguido por un período de sinaptogénesis acelerada [40,41]. El elemento presináptico es aportado por las colaterales axónicas crecidas como consecuencia de la desnervación [42]. Las nuevas sinapsis muestran, en un principio, una talla reducida de los elementos que la integran, similar a las de sinapsis recién formadas durante el período embrionario de sinaptogénesis. Con el paso del tiempo, el tamaño de los elementos sinápticos aumenta y adquiere características 'adultas' [40].

En este proceso de sinaptogénesis no sólo es importante la colateralización de los axones, sino que también las dendritas, que aportan el elemento post-sináptico, sufren modificaciones como consecuencia de la desnervación y participan activamente en el proceso de reconstitución.

En zonas como la corteza cerebelosa o el núcleo geniculado, donde no ocurren procesos de colateralización, las dendritas muestran dos tipos de respuesta a la desaferentización: un proceso de axonización, en el cual aparecen en las dendritas especializaciones presinápticas y formación de sinapsis dendrodendríticas; o, en su defecto, las dendritas sufren un proceso de atrofia gradual que conduce al incremento relativo de la densidad de innervación a expensas de los axones sobrevivientes, no colateralizados. Ambos fenómenos contribuyen a cierta recuperación funcional, a pesar de la ausencia de colateralización.

Los mecanismos dendríticos implicados en la sinaptogénesis reactiva no se conocen con detalle, aunque desde hace tiempo se sabe que ésta requiere una síntesis de proteínas activa [43]. En correspondencia, se ha demostrado la existencia de transporte dendrítico de ARN mensajero (p. ej., de la subunidad R1 del receptor NMDA) y un incremento notable de síntesis dendrítica de proteínas que sólo se manifiesta en las regiones desnervadas [44]. El transporte dendrítico de ARNm involucra también modestos incrementos de proteínas dendríticas de importancia funcional, como el de la proteína relacionada con microtúbulos 2 (MAP-2, del inglés *Microtubule Associated Protein*) y la proteinocinasa II dependiente de calcio-calmodulina (PK-II) [45].

La MAP-2 es una proteína regulada por fosforilación que sólo se localiza en las dendritas (a diferencia de tau que es exclusivamente axónica), forma parte del citoesqueleto dendrítico y podría tener un papel en el proceso de remodelación dendrítica [46,47].

Neurogénesis

La producción de nuevas células nerviosas en el cerebro adulto se ha demostrado en todas las clases de vertebrados [1]. En las aves se piensa que está directamente implicado en los procesos de maduración posnatal y en funciones estacionales como el canto [48,49]. En roedores se conocen dos áreas donde la neurogénesis se mantiene activa hasta edades muy avanzadas de la vida: la zona subventricular (ZSV) de los ventrículos laterales y el giro dentado del hipocampo [50,51].

Las células progenitoras son capaces de generar neuronas, astrocitos y oligodendrocitos, y su diferenciación parece controlada por señales ambientales que incluyen al ácido retinoico, a la adenosina monofosfato cíclico (AMPC) y factores tróficos. La depleción de serotonina reduce la producción de células nerviosas en el giro dentado y la ZSV [52], y lo mismo ocurre por deficiencia en hormonas tiroideas [53]. Por otra parte, las crisis epilépticas

aceleran la neurogénesis y la formación de circuitos aberrantes [54] que son importantes en la progresión del trastorno [55]. El establecimiento de circuitos aberrantes significa que las nuevas neuronas pueden formar sinapsis, a lo cual contribuye la expresión de moléculas de adhesión como la NCAM [56]. Un hallazgo interesante es el hecho de que ratas viejas que habitan en un ambiente complejo muestran un incremento de la neurogénesis [57]. Las células nerviosas recién formadas pueden migrar a regiones distantes [58], lo que añade un posible valor terapéutico a este interesante mecanismo.

Una comunicación reciente va más allá al proponer que neuronas ya diferenciadas pueden recuperar sus capacidades mitóticas si se colocan en un ambiente adecuado [59]. Se trata de un único estudio *in vitro* que debe ser confirmado, pero extraordinariamente provocativo.

Aunque no está resuelta la controversia sobre si existe neurogénesis en el cerebro adulto de los primates [60], es indudable que poder modular la formación de nuevas células nerviosas es una promesa de enormes potencialidades para la Neurología Restaurativa, tanto para la recuperación *in situ* de neuronas perdidas, como para el trasplante de células precursoras en zonas dañadas.

Plasticidad funcional

Plasticidad sináptica

Las sinapsis son especializaciones anatómicas y funcionales mediante las cuales la información, que circula en forma de pulsos eléctricos, es transferida de una neurona a otra. Las características funcionales de estas estructuras y los mecanismos de suma espacial y temporal que realizan las neuronas post-sinápticas son la base de las propiedades integradoras del sistema nervioso.

La importancia de las sinapsis en los procesos de almacenamiento de información se ha postulado desde la época de Ramón y Cajal y más recientemente en los trabajos de Hebb y Matthies [5,61-65]. Estos modelos 'conectivistas' de la memoria predicen cambios en la eficacia de la transmisión sináptica, en los circuitos neuronales implicados en la adquisición de nuevos contenidos de memoria. Atribuyen, por lo tanto, propiedades plásticas a las sinapsis y rompen con los conceptos primitivos que consideraban a las sinapsis inmutables en sus propiedades funcionales, como puntos de soldadura entre los componentes de un circuito eléctrico.

Formas de plasticidad

Las capacidades plásticas de las conexiones sinápticas pueden expresarse de formas diversas por su duración y por los mecanismos implicados [66].

Existen mecanismos que conducen a cambios transitorios, del orden de milisegundos a minutos, de la eficacia sináptica. La facilitación o inhibición por pulsos pareados y la llamada potenciación posttetánica son ejemplos de estas formas efímeras de plasticidad [67], que parecen depender de la acumulación de Ca^{2+} residual en la terminal presináptica [68]; asimismo, su duración es limitada por los mecanismos de tampón que reducen la concentración de este ion [69].

Sin embargo, existen formas mucho más duraderas de plasticidad sináptica. En 1973, se publicaron dos artículos simultáneos en el *Journal of Physiology* (Londres), en los que se describía un fenómeno de modificación a largo plazo de la eficacia de la transmisión sináptica [70,71]. Este fenómeno se ha llamado potenciación a largo plazo (LTP, del inglés *Long-Term Potentiation*) y se considera, hasta hoy, como el mejor modelo de cambio funcional en la conectividad sináptica dependiente de la

actividad. Desde su descubrimiento se le vinculó a los procesos de memoria, aunque en la actualidad se propone también como un mecanismo importante en la maduración funcional de las sinapsis y en los procesos de remodelación que conducen a la recuperación de funciones perdidas como consecuencia de lesiones o trastornos degenerativos.

La plasticidad sináptica a largo plazo puede también expresarse en una disminución de la eficacia en la transmisión. Si el cambio se produce en una población previamente potenciada, suele llamársele despotenciación, en otro caso se denomina depresión a largo plazo (LTD, del inglés *Long-Term Depression*). La LTP y la LTD pueden ocurrir en las mismas sinapsis dependiendo de la frecuencia de estimulación utilizada [72]. Frecuencias bajas, entre 1 y 5 Hz, conducen a LTD, mientras que frecuencias mayores de 25 Hz producen LTP [73]. En ambos casos se ha probado la participación de receptores de tipo NMDA [74] y corrientes de Ca^{2+} a la terminal post-sináptica. En el caso de la LTP, ello conduce a la activación de proteinocinasas, mientras que en la LTD, donde el incremento de Ca^{2+} es menor, se activan fosfatasa que tienen una función antagonista [75-77]. Aunque ambos fenómenos, en interacción dinámica, parecen implicados en los procesos de memoria [78], nos ocuparemos en lo sucesivo de la LTP cuya importancia funcional y mecanismos están mejor estudiados.

Mecanismos de la LTP. La LTP fue descrita inicialmente en el hipocampo. Si bien hoy sabemos que no es un fenómeno exclusivo de las sinapsis de esta estructura cerebral, la mayoría de los estudios acerca de los mecanismos de la LTP se han realizado en las sinapsis del giro dentado y de la región 1 del cuerno de Ammon (CA1). Por esa razón, describiremos a continuación los mecanismos de la LTP en estas poblaciones y después comentaremos la LTP en otras regiones del SNC.

Receptores glutamatérgicos. Existen dos tipos fundamentales de receptores glutamatérgicos. Los receptores ionotrópicos forman canales iónicos y son responsables de la despolarización de la membrana post-sináptica. En ellos se distinguen los de tipo AMPA, kainato y NMDA, según el nombre del agonista más afín. El otro tipo son los llamados receptores metabotrópicos, una familia de ocho miembros conocidos que —como su nombre indica— se relacionan con cambios metabólicos más que con conductancias iónicas [79,80]. Los receptores AMPA/kainato son los responsables de la transmisión sináptica normal, pues median corrientes de sodio que despolarizan la membrana post-sináptica. Sin embargo, cuando el nivel de despolarización alcanza un valor umbral en presencia de glutamato, se produce la activación de los receptores de tipo NMDA. Este ionóforo abre canales de Ca^{2+} y su activación es imprescindible para la inducción de la LTP [81,82] (Fig. 3). Los receptores glutamatérgicos metabotrópicos también parecen implicados en la inducción de la LTP [83-85], aunque los resultados han sido contradictorios [86]. La entrada de Ca^{2+} parece ser el hecho decisivo. Los receptores AMPA son importantes porque posibilitan la activación NMDA, como indican estudios recientes en mutantes carentes de estos receptores [87], y también porque el resultado final de todos los procesos celulares implicados en la LTP podría ser el aumento en la densidad de estos receptores [88].

Papel del calcio. La entrada de calcio en la terminal post-sináptica es condición necesaria para iniciar los procesos de potenciación sináptica [89]. El ingreso de Ca^{2+} se realiza normalmente por los

ionóforos NMDA, aunque también puede lograrse por la vía de canales de activación lenta dependientes de voltaje (VDCC) [90]. Este ion, como segundo mensajero, media todas las formas de plasticidad sináptica a largo plazo, LTP y LTD. La diferencia parece depender de la cantidad de calcio que penetra y, por consiguiente, del incremento en su concentración intracelular [91,92]. Si el incremento sobrepasa un nivel umbral, lo que normalmente ocurre con estimulación de alta frecuencia, el Ca^{2+} conduce a la activación de proteinocinasas que son responsables, por una parte, de mantener transitoriamente el estado de respuesta incrementada y, por otra, de activar procesos de transcripción/traducción que conducen a la estabilización del cambio sináptico. Algunas de estas proteinocinasas pueden actuar sobre los reservorios de Ca^{2+} mitocondriales y del retículo endoplasmático y liberar calcio al citoplasma, lo cual contribuye a la elevación de la concentración intracelular de este ion.

Proteinocinasas. La adición de grupos fosfato a las proteínas es un importante mecanismo de regulación de su función. Esta función está a cargo de enzimas especiales conocidas como proteinocinasas. Existen dos grandes grupos de proteinocinasas según el residuo aminoácido blanco de la fosforilación: las tirosinocinasas (como es el caso de los receptores de neurotrofinas) y las serina-treonina proteinocinasas. Aunque existen evidencias recientes que involucran a algunas tirosinocinasas en la LTP, su papel no está bien establecido. Nos referiremos, por lo tanto, a las tres serina-treonina proteinocinasas con funciones comprobadas en los procesos de plasticidad sináptica.

La PK-II es activada por iones de Ca^{2+} y calmodulina por lo que también es llamada Ca-CamK-II. La activación se produce por fosforilación del residuo de treonina en posición 286 y ello conduce a un mecanismo multiplicador por autofosforilación [93-96].

Inhibidores de la PK-II, como el calmidazolol, bloquean la LTP desde etapas tempranas (<30 minutos) [97]. Se han encontrado resultados coincidentes en ratones mutantes carentes de la subunidad alfa de la PK-II [98,99], lo que demuestra la importancia de una activación temprana de la PK-II para el mantenimiento de la LTP.

La PK-II es muy abundante en las densidades post-sinápticas y se relaciona con las subunidades NR-1 y NR-2B del receptor NMDA, una posición estratégica para su activación inmediata por el calcio que penetra a través de este ionóforo [100]. Este hecho coloca a esta enzima en la vecindad de su principal sustrato: el receptor AMPA, responsable de la despolarización post-sináptica y, por lo tanto, de la eficacia de la transmisión. De este modo, la PK-II puede mediar la potenciación de sinapsis glutamatérgicas, incrementando la conductancia de canales AMPA existentes en la membrana post-sináptica [101].

Otra serina-treonina cinasa implicada en la LTP es la proteinocinasa C (PKC). Esta proteinocinasa se activa por Ca^{2+} y diacilglicerol (DAG). El calcio penetra por los canales NMDA. La fuente de DAG implica la activación previa de fosfolipasas, que actúan sobre los lípidos de membrana liberando DAG e inositol-3-P (IP3). El IP3 puede incrementar la concentración citoplásmica de Ca^{2+} y activar su liberación desde almacenes intracelulares.

La importancia de la PKC para la LTP está bien establecida. La administración de bloqueadores de la PKC no afecta la inducción de la LTP; sin embargo, la eficacia sináptica retorna a su valor basal en aproximadamente dos horas [102-104]. Por otra parte, activadores de la PKC como los ésteres de forbol prolongan una LTP débil [105,106]. La PKC es fosforilada y activada de forma duradera

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como la anisomicina, bloqueadores de la síntesis de estas macromoléculas, no afectan la inducción inicial del fenómeno plástico pero limitan su duración a unas 4-6 horas [130-132]. Dendritas separadas del soma desarrollan, pero no mantienen, la LTP [133]. Estos estudios dieron origen al concepto de que la LTP se desarrolla en fases dependientes de mecanismos moleculares diversos, como la memoria [134,135].

Ha resultado muy difícil identificar qué proteínas particulares son sintetizadas tras la inducción de la LTP [136,137], dada la multiplicidad de proteínas que son activadas como resultado de la activación nuclear. En una primera oleada, se activan proto-oncogenes (también conocidos como genes de respuesta inmediata) [138]; entre ellos se han identificado varios como el *zif-268*, la subunidad alfa de la PK-II, *c-fos*, *c-jun*, *jun B*, *jun D* y *krox-20* [139-142]. Los productos de muchos de estos proto-oncogenes, a su vez, constituyen factores de transcripción para otros genes estructurales. Tal es el caso de las proteínas Fos y Jun que se dimerizan para constituir el factor de transcripción AP-1. Ello conduce a una segunda oleada, más tardía, de síntesis de proteínas que también participan en los procesos de plasticidad. Se ha calculado que existen entre 500 y 1.000 genes relacionados con la plasticidad [143], lo que sin dudas dificulta la identificación de esos agentes. No obstante, trabajos recientes apuntan a un incremento en el número de receptores glutamatérgicos post-sinápticos [144], particularmente de tipo AMPA [145].

Un problema derivado de la relación LTP-síntesis de proteínas es su distribución. La LTP es específica y sólo las sinapsis activadas se potencian. ¿Cómo reconocen las proteínas responsables del cambio plástico aquellas sinapsis que fueron previamente activadas? Frey y Morris han demostrado recientemente que la inducción de la LTP en una sinapsis no sólo aumenta temporalmente su eficacia de transmisión por cambios locales (p. ej., fosforilación de receptores AMPA), sino que establece en ella una marca temporal que sirve de señal de identificación para las proteínas responsables del cambio duradero de conectividad (p. ej., inserción de nuevos receptores AMPA) [146,147]. La naturaleza molecular de la marca sináptica se investiga actualmente.

Cambios morfológicos. Los mecanismos antes citados explican la LTP en términos de modificaciones moleculares que conducen a cambios funcionales. Existen evidencias de que además, especialmente en fases más tardías (>8 horas), pueden aparecer cambios detectables en la morfología de las sinapsis que también podrían estar implicadas en la LTP.

Así, por ejemplo, se ha observado un aumento del número de sinapsis perforadas, con zonas de transmisión divididas que más tarde se convierten en espinas dendríticas dobles [148,149], las cuales, al parecer, representan un proceso de proliferación sináptica local. El incremento de espinas dendríticas cortas y gruesas después de la potenciación [150] podría ser expresión de este fenómeno.

La participación en la LTP de moléculas de adhesión, como la NCAM [27,151,152], que sirven de guía para el crecimiento axonal, o de proteínas presinápticas como la SNAP-25 [153,154] como parte de la intensa síntesis proteica relacionada con la LTP, son compatibles con la idea de que respuestas de diferenciación y proliferación puedan participar en la LTP [155].

Estos hallazgos permiten suponer que la sinaptogénesis podría ser la base de las fases más tardías de la LTP (días, semanas) [156]. La sucesión de mecanismos implicados en el sustento temporal de la LTP demuestra la estrecha imbricación de los mecanismos neuroplásticos. Comienza por cambios en el área funcional y culmina con procesos de crecimiento. En su aparente diversidad

y complejidad, este universo fenomenológico es uno, simple y parsimonioso.

Cambios presinápticos. Puede lograrse mayor eficacia sináptica mediante:

1. El aumento de la cantidad de neurotransmisor liberado por la terminal presináptica.
2. El aumento de la afinidad de los receptores post-sinápticos por el neurotransmisor.
3. El aumento de la densidad de los receptores post-sinápticos.

Los mecanismos descritos hasta ahora tienen lugar, y afectan, principalmente los componentes post-sinápticos, lo que no excluye la participación de elementos presinápticos.

Aunque existen evidencias de incrementos en la liberación de glutamato en la LTP [157-159], ésta no ha podido demostrarse en etapas tardías de la LTP. Evidencias indirectas, como el aumento de proteínas relacionadas con la fusión vesicular y la liberación del neurotransmisor, 5 horas después de la inducción de la LTP hablan en favor de esta hipótesis [160]. Relacionado con ello se encuentra el hecho de que la LTP aumenta la confiabilidad de la transmisión sináptica en el hipocampo [161], que es normalmente poco fiable porque algunos potenciales de acción no provocan liberación de neurotransmisor.

El componente presináptico de la LTP requiere, sin embargo, la activación de las neuronas post-sinápticas para producirse. Se ha planteado que la neurona post-sináptica libera algún mensajero que difunde retrógradamente hasta la terminal presináptica y allí provoca los cambios mencionados. Este mensajero hipotético no ha sido identificado y se han propuesto diversos candidatos, entre ellos la adenosina [162], el factor neurotrófico derivado del cerebro (BDNF, del inglés, *Brain Derived Neurotrophic Factor*) [163] el óxido nítrico y el monóxido de carbono [164-166], el factor activador de plaquetas [167] y el ácido araquidónico [168].

Interacciones heterosinápticas. Para inducir una LTP duradera se requieren estímulos intensos y de alta frecuencia que provoquen la activación simultánea de un gran número de aferentes glutamatérgicas. Este ha sido uno de los argumentos más fuertes en contra de la LTP como mecanismo fisiológico de plasticidad sináptica. Ello, unido al hecho antes citado de que algunas cinasas implicadas en la LTP no parecen ser activadas eficazmente por la vía de los receptores NMDA, ha concedido relevancia al posible papel cooperativo de otras sinapsis en el desarrollo y mantenimiento de la LTP.

Existen evidencias de que, en efecto, la LTP puede ser modulada por sinapsis no glutamatérgicas con efectos metabotrópicos [169,170].

La privación del hipocampo de aferentes colinérgicos bloquea la LTP en el giro dentado [171,172] y el trasplante de tejido fetal colinérgico a esos animales restaura la LTP [173]. La inducción de la LTP es facilitada durante la actividad theta en el hipocampo por un ritmo colinérgico [174,175]. La administración de agonistas muscarínicos es capaz de inducir una potenciación que se desarrolla lentamente, y mimetiza las fases tardías de la LTP [176,177]. Los receptores muscarínicos, relacionados con la proteína G, promueven la activación de proteinocinasas que actúan sinérgicamente con los mecanismos dependientes de NMDA en los procesos de plasticidad sináptica [178].

La administración de antagonistas dopaminérgicos bloquea las fases tardías de la LTP [179,180]. La administración de agonistas, por el contrario, induce una potenciación retrasada que también

depende de la síntesis de proteínas [181]. Asimismo, en este caso, la dopamina parece actuar a través de cinasas como la PKA [119].

La depleción de sodio afecta la LTP en el giro dentado del hipocampo [182,183] y viceversa, la administración de sodio induce una potenciación lenta, similar a la descrita para la acetilcolina y dopamina, que es bloqueada por inhibidores de la síntesis de proteínas [184]. Estas evidencias sostienen la hipótesis de que la sodio ejerce una acción moduladora sobre los procesos de plasticidad sináptica [185].

Otras aminas biógenas como la serotonina [186,187] o la histamina [188] también muestran efectos moduladores sobre la LTP.

Todas estas aferencias, colinérgicas, dopaminérgicas, adrenérgicas, etc., ejercen sus efectos a través de la PKA [170]. De este modo, la activación de aferentes glutamatérgicos de origen cortical (instructivos) induce procesos de plasticidad en el hipocampo que son facilitados por aferentes de origen subcortical (moduladores); un posible ejemplo de cómo interactúan, a nivel celular, el intelecto y las emociones.

Interacción amígdala-hipocampo. Una manifestación de lo anteriormente comentado es la interacción entre la amígdala y el hipocampo en la LTP.

La amígdala es una estructura del sistema límbico que constituye el sustrato neurológico de las emociones [189] y de la memoria emotiva [190,191]. También se ha demostrado que modula el almacenamiento de memoria en otras estructuras cerebrales como el hipocampo [192].

En el ámbito funcional, la estimulación de la amígdala favorece la inducción de la LTP en el hipocampo [193,194]. Recientemente, hemos demostrado que también la fase de mantenimiento tardía de la LTP se beneficia de la estimulación de la amígdala. Este efecto fue bloqueado por antagonistas muscarínicos y noradrenérgicos y por lesión de la fimbria-fórnix, lo que sugiere una mediación del septum en esta acción (Frey, Bergado et al [remetido]; Jas, Almaguer et al [en prensa]). De este modo, una LTP débil y de corta duración, inducida por la estimulación de las aferencias glutamatérgicas, puede ser convertida en una LTP fuerte y duradera por coactivación, dentro de una ventana temporal, de la amígdala. Este modelo funcional explica, en el ámbito celular, las relaciones amígdala-hipocampo en la memoria y la forma en que los factores emocionales-motivacionales influyen en ella.

Plasticidad sináptica y memoria. Se ha considerado a la LTP no sólo como una forma de plasticidad sináptica, sino también como un modelo celular de memoria [195]. Realmente, no existe una prueba definitiva y concluyente [196], pero muchas evidencias coinciden en indicar que dicha relación es verdadera.

Para demostrar que la LTP es un mecanismo celular de memoria se han intentado diversas estrategias. Una de ellas, la saturación, se basa en el siguiente razonamiento: si la LTP es la base funcional de la memoria, inducir una LTP saturada en el hipocampo debe impedir la adquisición de nuevos contenidos de memoria. Los resultados han sido contradictorios [197-199]. La debilidad fundamental de este enfoque es que nada asegura que las sinapsis en estudio estén necesariamente implicadas en el modelo de aprendizaje utilizado. Una aproximación correlacional ha sido, hasta ahora, más fructífera; utilizando la estimulación de la vía perforante como estímulo condicionado en una prueba de evitación activa, se encontraron cambios sinápticos similares a la LTP sólo en aquellos animales que aprenden bien la prueba [200].

Otras evidencias señalan que los receptores de tipo NMDA, cuya importancia para la inducción de la LTP ya hemos comen-

tado, también participan en los procesos de adquisición de nuevos contenidos de memoria [201-203]. Asimismo, la PK-II también interviene en la formación de nuevos trazos de memoria, según demuestran los resultados de trabajos con mutantes deficientes de esta enzima [204]. Finalmente, se han encontrado cambios en la morfología de las sinapsis hipocampales, similares a los que ocurren en la LTP después del entrenamiento [205].

Estas coincidencias entre los mecanismos de la memoria y la plasticidad sináptica sustentan la convicción de que existe una relación funcional entre ambos procesos y validan los modelos conectivistas de aprendizaje.

Plasticidad funcional en otras regiones cerebrales. Los mecanismos de plasticidad sináptica funcional que acabamos de describir en el hipocampo no son exclusivos de esta estructura. Fenómenos de tipo LTP o LTD se han documentado experimentalmente en otras regiones del SNC.

Plasticidad cortical. El estudio de potenciales de campo inducidos por estimulación de la sustancia blanca subyacente ha demostrado LTP en diversas regiones corticales [206].

Las neuronas en la corteza muestran dos patrones de organización; un patrón laminar clásico que queda determinado prenatalmente y una organización funcional de tipo columnar que se desarrolla posnatalmente a partir de la estimulación que recibe [207] según la experiencia particular del individuo.

Los estudios en la corteza visual han documentado la importancia de procesos plásticos en el desarrollo de las capacidades funcionales de este sistema [208]. Mecanismos similares operan en otras áreas como la corteza somatosensorial y motora [209,210], auditiva [211,212] y áreas de asociación [213,214]. Las sinapsis talamocorticales e intracorticales que se establecen durante la embriogénesis son funcionalmente inmaduras, tal vez silentes, y su maduración dependiente de la estimulación parece implicar mecanismos similares, si no idénticos, a los descritos en la LTP hipocampal.

La activación de receptores NMDA y metabotrópicos [213,214], la entrada de Ca^{2+} [215], la activación de proteinocinasas [216] y de factores de transcripción de proto-oncogenes [217], seguida de la síntesis de proteínas funcionales [218] y, a más largo plazo, cambios morfológicos en las espinas dendríticas y formación de nuevas sinapsis [210], forman parte de la cadena funcional de la plasticidad cortical. A estas coincidencias se añade que los procesos de plasticidad cortical también son modulados heterosinápticamente por aferencias subcorticales [219].

El desarrollo de remodelaciones neuroplásticas puede modificar la representación cortical de funciones. Por ejemplo, la región ectosilviana de la corteza de asociación parietal es un área de relación polimodal con regiones visuales, auditivas y somatosensoriales. Tras la privación visual bilateral, la representación visual ectosilviana es 'tomada' por aferencias de las otras modalidades [220].

Pero también las sinapsis corticales muestran formas de plasticidad sináptica, incluyendo, además de LTP, la LTD e interacciones a corto plazo [209]. Estas pueden intervenir en la recuperación de funciones perdidas por daño o degeneración sin que, necesariamente, se produzcan modificaciones importantes en la cartografía cerebral de esas funciones [221].

En un interesante estudio se comprobó que la reorganización de la corteza auditiva por estímulos acústicos se potenciaba de forma significativa cuando se apareaba con estimulación eléctrica del núcleo basal magnocelular, proveedor de inervación colinérgica a la corteza [222]. Ello indica que los mecanismos de plasticidad cortical,

tal como describimos para la LTP hipocampal, son también modulables por señales metabotrópicas subcorticales; una analogía que confirma la universalidad de los mecanismos de neuroplasticidad.

Las capacidades plásticas corticales disminuyen con la edad, sobre todo en las áreas cerebrales primarias, pero siempre se conserva algún grado de plasticidad, especialmente en las áreas asociativas [223]. Al margen de las implicaciones terapéuticas de este hecho, esta diferencia podría contribuir a resolver la vieja disputa entre localizacionistas y antilocalizacionistas [224].

El cierre aparente de las capacidades neuroplásticas corticales ha dado origen al concepto de los períodos críticos del desarrollo después de los cuales se pierde la posibilidad de modificación funcional. Los períodos críticos no son absolutos y pueden reabrirse cuando se activan mecanismos neuroplásticos. Aunque no se conocen con exactitud los agentes moduladores, existen evidencias de que los receptores glutamatérgicos post-sinápticos también son importantes en esta función [225].

El estudio de los determinantes celulares y moleculares de los períodos críticos es un objetivo importante de la Neurología Restaurativa.

Plasticidad en otras estructuras. También se ha descrito LTP en la amígdala [226]. La estimulación eléctrica de núcleos talámicos produce potenciales evocados en la región lateral de la amígdala [227]; esta aferencia desarrolla una LTP [228] relacionada con el desarrollo de reacciones de miedo condicionado [229] y podría constituir la base de una memoria emocional. La proyección del hipocampo a la amígdala es potenciabile por mecanismos dependientes de NMDA [230].

En el cerebelo se han descrito fenómenos plásticos de tipo LTD relacionados con el aprendizaje motor [231] dependientes de receptores metabotrópicos [232] y que pueden conducir a cambios morfológicos en las dendritas de las células estrelladas [233].

La LTP constituye la forma normal de plasticidad dependiente del uso en el estriado [234], en el accumbens (donde depende de receptores NMDA) [235,236] y en la médula espinal [237,238].

Sinapsis silentes

La existencia de sinapsis no funcionales, llamadas sinapsis silentes, se ha demostrado en especies tan lejanas como peces y mamíferos [239,240]. Las sinapsis silentes representan una reserva funcional que puede ser importante para la expresión de fenómenos neuroplásticos [239].

Estudios recientes han demostrado que, en el hipocampo, estas sinapsis sólo expresan receptores NMDA, circunstancia que las hace no funcionales a los valores normales de potencial de membrana en reposo [240]. Su conversión en sinapsis activas implica la aparición de receptores AMPA funcionales, bien por síntesis *de novo* o por alguna modificación molecular que los capacita [240,241]. La activación repetitiva, tal como ocurre en la inducción de la LTP, desencadena este proceso. De modo que la LTP puede ser considerada como un mecanismo de maduración sináptica [241,242], tanto en el adulto como durante la ontogénesis [243]. Takumi et al [244] han calculado que en la región CA1 del hipocampo adulto aproximadamente el 25% son sinapsis silentes y que la coexpresión de receptores NMDA y AMPA se relaciona con un aumento del tamaño de la densidad post-sináptica, correspondiendo a un cambio funcional.

La existencia de sinapsis silentes y su maduración por incorporación de receptores AMPA, mediada por activación repetitiva de NMDA, también ha sido demostrada en la corteza cerebral [245].

El mecanismo de activación de sinapsis silentes muestra similitudes llamativas con la LTP. Ambas comienzan con la activación de receptores NMDA y terminan (¿terminan?) con la incorporación de receptores AMPA a la membrana [156]. En qué medida la activación de sinapsis silentes podría ser parte del fenómeno de potenciación sináptica es una pregunta razonable que deberá resolverse en el futuro. Lo que queremos resaltar es el hecho de que parece existir un *continuum* de modificaciones, desde sutiles cambios de eficacia sináptica hasta la formación de nuevas sinapsis, pasando por la activación de contactos silentes, sustentados por mecanismos moleculares comunes. Esta simplicidad dentro de la aparente complejidad es uno de los fundamentos teóricos de la esperanza de lograr, en breve plazo, una comprensión de los fenómenos neuroplásticos que permita dirigirlos con éxito en el marco de nuevos esquemas terapéuticos y neurorrestaurativos.

Universalidad de los mecanismos de plasticidad

Los mecanismos de neuroplasticidad en adultos son, en esencia, idénticos a los que ocurren durante el desarrollo embrionario del SNC y operan en especies tan lejanas como moluscos y ratas. La conservación ontogenética y filogenética de los mecanismos de neuroplasticidad es una prueba de su valor adaptativo.

Estudios recientes sobre las moléculas de reconocimiento neuronal han revelado funciones similares para estas moléculas durante el desarrollo embrionario y la plasticidad en el adulto [246]. La diferencia fundamental estriba en que durante la primera se establece la circuitería nerviosa, vías y blancos, que es común a todos los miembros de una especie; mientras la segunda garantiza el desarrollo fino de patrones funcionales altamente precisos [247], individualizados según la experiencia particular de cada sujeto.

Desde el punto de vista filogenético se descubren cada día nuevas coincidencias.

En *Aplysia californica*, un molusco, existe LTP [248] que es dependiente de la síntesis de proteínas [249] y está relacionada con la adquisición de reacciones condicionadas simples [250], que son las formas de aprendizaje presentes en estos animales.

La mosca *Drosophila melanogaster* es capaz de aprender y desarrollar procesos neuroplásticos que son mediados por PK-II [251] y AMPc [252].

Finalmente, la universalidad de los mecanismos de plasticidad también se expresa en la identidad de los mismos en regiones diferentes del SNC, como el hipocampo y la corteza [253]. La naturaleza es parsimoniosa y no desdénia mecanismos eficaces por antiguos que sean.

Envejecimiento

Envejecer no es una enfermedad, sin embargo, en el proceso de envejecimiento normal se descubren cambios en las capacidades neuroplásticas que pueden estar relacionadas con las afectaciones de memoria que caracterizan al anciano.

Las ratas viejas aprenden lentamente y olvidan de manera rápida, posiblemente debido al deterioro de sus capacidades neuroplásticas [254]. De forma coincidente, en estos animales la LTP se desarrolla lentamente [255] y decae con rapidez [256,257]. También la LTD se encuentra afectada [258]. El deterioro de estas capacidades obedece, al menos en parte, a déficit en los mecanismos moleculares que los sustentan [259].

También la plasticidad por crecimiento se afecta por el envejecimiento. La colateralización de fibras comisurales y de asociación en el giro dentado, posterior a una lesión de corteza entorrinal está disminuida en ratas viejas [260], así como la colateralización

Tabla II. Factores neurotróficos. La muestra no pretende ser exhaustiva, pues tan sólo recoge una selección de algunos factores proteicos con acción neurotrófica probada. Los grupos o familias de factores tróficos se definen según el grado de homología estructural de sus miembros. Nótese que en todos los casos el mecanismo de acción implica receptores de membrana específicos para cada factor y fosforilación de tirosina (mediada por el propio receptor o por moléculas relacionadas) y la activación de cascadas celulares del tipo MAP-cinasa (una serina-treonina cinasa) y otras con actividad de regulación nuclear. (Se han mantenido las siglas en inglés, impuestas por el uso).

Grupo	Siglas	Nombre	Receptores y cascadas
Neurotrofinas	NGF	Factor de crecimiento nervioso	Tirosinocinasas unidas a membrana (Trk), MAP-cinasa y otros reguladores nucleares
	BDNF	Factor neurotrófico derivado del cerebro	
	NT-3	Neurotrofina 3	
	NT-4/5	Neurotrofina 4/5	
	NT-6	Neurotrofina 6	
Factores hematopoyéticos	CNTF	Factor neurotrófico ciliar	Receptores de membrana sin actividad enzimática intrínseca (CNTF-R α) que atraen a tirosinocinasas solubles, MAP-cinasas y otros reguladores nucleares
	GDNF	Factor neurotrófico derivado de la glía	
Factores de crecimiento	EGF	Factor de crecimiento epidérmico	Tirosinocinasas unidas a membrana (EGFR), MAP-cinasa y otros reguladores nucleares
	FGF	Factor de crecimiento de fibroblastos	
	TGF	Factor de crecimiento transformante	
	TNF	Factor de necrosis tumoral	
	PDGF	Factor de crecimiento derivado de plaquetas	
	IGF	Factor de crecimiento similar a la insulina	

de fibras simpáticas en la misma región, consecutiva a lesión de la fimbria-fornix [261], aunque las fibras colinérgicas septales parecen conservar cierta capacidad de respuesta [38].

La reducción no es absoluta. En ratas viejas no lesionadas se observa una expansión discreta del tercio interno del estrato molecular por colateralización [262].

Enfermedades neurodegenerativas

Los déficit de memoria son más graves en desviaciones patológicas del proceso de envejecimiento, como la demencia de Alzheimer. Las causas de la enfermedad no son conocidas, como tampoco sus consecuencias funcionales, pero entre las hipótesis emitidas algunas atribuyen el deterioro cognitivo a una pérdida de plasticidad [263]. Ratones mutantes para la ApoE muestran una plasticidad sináptica alterada en el hipocampo [264] al igual que animales deficientes en proteína precursora del amiloide [265,266]. Se ha sugerido que la presencia de neuritas anormales en las placas seniles son el resultado de intentos regenerativos fallidos [267].

Otras entidades neurodegenerativas como la enfermedad de Parkinson y la corea de Huntington también se vinculan a formas alteradas de plasticidad [268,269]. Ratones portadores de la forma mutada del gen para la huntingtina muestran una reducción significativa de la LTP [270].

Epilepsia

La neuroplasticidad puede relacionarse con procesos patológicos no solo por defecto. La epilepsia es el ejemplo mejor caracterizado de cómo procesos de plasticidad excesivos y aberrantes pueden afectar la función normal del SNC.

Las crisis epilépticas provocan muerte neuronal por apoptosis

y necrosis que es seguida, en las neuronas que sobreviven, por el desencadenamiento de fenómenos plásticos [271].

Tras crisis epileptógenas en el giro dentado, se produce una extensión y colateralización axonal de las neuronas principales [272] que se expresa por aumentos en la expresión de GAP-43 [18,273] y otros marcadores de crecimiento axonal [274] y dendrítico [275]. Los axones en crecimiento son guiados por moléculas de adhesión de tipo NCAM [276,277].

El crecimiento axonal se dirige fundamentalmente, en forma recurrente, al tercio interno del estrato molecular [278] donde se establecen sinapsis excitatorias, incluso autapsis, que son responsables del estado de hiperexcitabilidad [279-282]. Como ocurre en otros casos de sinaptogénesis reactiva, también se producen cambios en las dendritas y espinas dendríticas que reciben las colaterales recurrentes [283,284].

Esta misma secuencia de acontecimientos se manifiesta en la región CA1 con idénticos resultados [282]. Fenómenos de este tipo ocurren desde etapas tempranas del desarrollo posnatal [285] y existen buenas razones para creer que están realmente implicadas en la epileptogénesis en humanos [286-289].

FACTORES EPIGENÉTICOS MODULADORES DE LA PLASTICIDAD

Los mecanismos de neuroplasticidad pueden contribuir, de modo notable, a la recuperación de funciones nerviosas. La pregunta que se deriva es: ¿cómo podemos estimular, modular y controlar los procesos neuroplásticos para lograr una mejoría más completa? Existe una variada gama de agentes que pueden modificar, de alguna manera, los procesos de neuroplasticidad; aprender a uti-

lizarlos adecuadamente es una de las tareas más importantes de la Neurología Restaurativa.

Factores neurotróficos

Desde el descubrimiento del factor de crecimiento nervioso (NGF, del inglés *Nerve Growth Factor*) la relación de moléculas proteicas de origen natural, capaces de estimular y promover la supervivencia y desarrollo de las células nerviosas ha crecido continuamente. Hoy, además de las neurotrofinas propiamente dichas, se conocen una variada gama de moléculas con capacidades neurotróficas (Tabla II). Estos factores neurotróficos se agrupan en familias según el grado de homología molecular de sus miembros y el tipo de receptor que utilizan para lograr sus efectos tróficos, y muestran un alto grado de conservación filogenética [290], una evidencia evolutiva de su importancia.

La hipótesis neurotrófica atribuye a estos factores una acción principal en la supervivencia de las neuronas. De acuerdo con esta concepción, los factores neurotróficos producidos por el blanco son captados por las terminales presinápticas y transportados retrógradamente por las neuronas. El suministro continuo de un factor neurotrófico específico resulta imprescindible para mantener la vida y expresión fenotípica de las neuronas [291,292]. Esto parece particularmente importante durante el desarrollo, período en el cual un gran número de neuronas muere por apoptosis, aparentemente porque no pudieron proveerse de una cantidad suficiente de factor trófico [293].

Los factores tróficos ejercen sus efectos a través de receptores de membrana que conectan con diferentes cascadas moleculares intracelulares, como la MAP-cinasa, la PKC y la fosfatidil inositol 3-cinasa (PI3-K), capaces de modificar la expresión génica y la síntesis de proteínas (Fig. 4) [294-296]. Ello, a su vez, les capacita para inducir y modular los procesos de neuroplasticidad por crecimiento o funcional. Otros factores con acción neurotrófica, como los factores de crecimiento (EGF, FGF, PDGF e IGF), a través de receptores diferentes, activan las mismas cascadas y pueden ejercer, por lo tanto, acciones similares a las de las neurotrofinas [296].

La producción de factores tróficos en el hipocampo puede ser inducida por crisis epilépticas provocadas [297] o por estímulos inductores de LTP [298-300]. Por otra parte, la administración de algunos factores tróficos, en particular el BDNF, es capaz de inducir potenciación sináptica de tipo LTP [301], lo cual se ha visto reforzado por estudios posteriores con mutantes BDNF(-) [302-304] y otros paradigmas experimentales [305,306]. Los mecanismos de esta LTP inducida por neurotrofinas dependen de la fosforilación de receptores NMDA [307], la activación de proteinocinasas [308,309] y la síntesis de proteínas [156,310], sin olvidar el papel de hipotético mensajero retrógrado en la inducción de cambios presinápticos. En este sitio también el mecanismo involucra fosforilación mediada por MAP-cinasa [309].

Las neurotrofinas también pueden sustentar procesos de plasticidad sináptica indirectamente y reforzar la influencia de aferentes no glutamatérgicas moduladoras de la LTP [255,311].

Los factores tróficos tienen un papel importante en los procesos de plasticidad cortical que conducen a la maduración funcional—dependiente de la experiencia—de las conexiones talamocorticales; ello ha sido bien estudiado en el sistema visual [312-314] y en el somatosensorial [315]. Por esa razón, se les confiere importancia especial en la determinación de los períodos críticos del desarrollo [316].

El BDNF y el NGF potencian la transmisión excitadora en la

corteza visual [317] y la LTP de estas conexiones sinápticas [318]. Se ha propuesto que la actividad visual incrementa la síntesis de BDNF por activación de promotores específicos y esto, a su vez, conduce a elevar la eficacia de la transmisión por mecanismos dependientes de NMDA [319]. Sin embargo, aunque los efectos de las neurotrofinas exógenas son dramáticos, su acción en condiciones fisiológicas es aún dudosa [320].

Evidencias experimentales sugieren una acción neuroprotectora de las neurotrofinas ante distintos insultos que comprometen la integridad y supervivencia de las neuronas, mediante la activación de sistemas enzimáticos implicados en la defensa celular [321-325].

Por todas estas razones, los factores neurotróficos han sido, durante más de una década, la gran esperanza para encontrar un tratamiento efectivo que combatiera las devastadoras consecuencias de las enfermedades neurodegenerativas. Su uso, avalado por experiencias en modelos animales, se ha propuesto para el tratamiento de la demencia de Alzheimer [255,300,311,326-328], la enfermedad de Parkinson [329-331], la esclerosis lateral amiotrófica [332,333] y la corea de Huntington [334], entre otras.

Pero dichas esperanzas no han sido satisfechas por muchas razones. La diversidad de factores con acción neurotrófica específica dificulta encontrar el más adecuado para una población neuronal dada. En la actualidad, se conocen más 20 factores tróficos capaces de sustentar a las neuronas dopaminérgicas de la sustancia *nigra* [329]. ¿Cuál de ellos, o combinación de ellos, es más eficaz? El panorama se complica aún más si consideramos que la sensibilidad a los factores tróficos varía en diferentes etapas del desarrollo, fenómeno conocido como conmutación neurotrófica (*neurotrophin switching*) [335]. En aquellos casos en que la sensibilidad a un factor trófico está demostrada, faltan estudios sistemáticos de dosis-respuesta, lo cual es importante para evitar o minimizar efectos secundarios y porque el propio efecto del factor trófico puede variar, incluso invertirse, dependiendo de la dosis. Por ejemplo, el exceso de BDNF tiene efectos proconvulsivos en regiones límbicas [336] y altas concentraciones de NGF detienen el crecimiento neurítico en neuronas periféricas [337]. Finalmente, comienza a comprenderse que el tipo de efecto trófico de un agente puede modificarse dependiendo de factores ambientales. La inhibición de la fosfolipasa- γ convierte al NGF, un antimitógeno clásico, en un factor mitogénico [338]; otro ejemplo, el BDNF y la NT-4/5 protegen a las células de Purkinje aisladas en cultivo, pero el efecto se invierte si se añaden células granulares [156].

Pero el pecado original de los factores tróficos sigue siendo su naturaleza proteica y, por lo tanto, su incapacidad para atravesar la barrera hematoencefálica. La implantación de células manipuladas genéticamente para producir una neurotrofina [332,334,339-343], o el uso de moléculas modificadas para aumentar su permeabilidad en la barrera hematoencefálica [344], han sido las alternativas más favorecidas.

En menor medida se ha intentado el uso de agentes periféricos que estimulen la producción intracerebral de factores tróficos, aunque algunas evidencias sugieren que agentes químicos [345-348] o fisiológicos (la actividad física) [349] y el estrés [350], pueden tener efectos moduladores de la producción endógena de factores tróficos.

En consecuencia, los ensayos clínicos han sido escasos y sus resultados poco prometedores [351]. Muchas preguntas y factores técnicos deben resolverse antes de que la promesa del uso farmacológico de los factores tróficos se convierta en realidad.

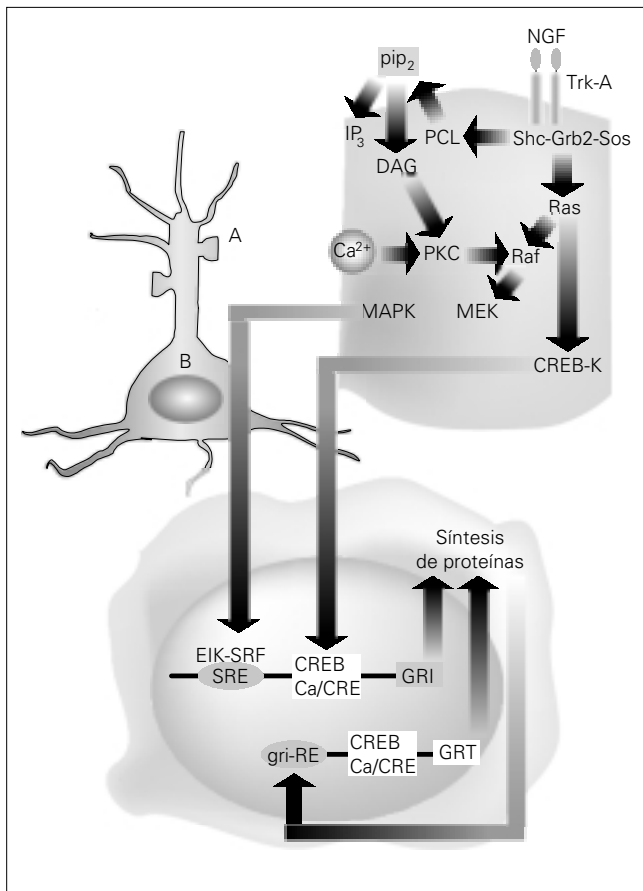


Figura 4. Mecanismo de acción de las neurotrofinas. a) La unión del factor de crecimiento nervioso al receptor tirosinocinasa A provoca su dimerización y autofosforilación en residuos de tirosina. Ello promueve la formación de un complejo tetramérico compuesto además por la proteína Shc fosforilada, el Grb2 y SoS que, de esta forma, son capaces de activar a Ras, una proteína G pequeña, por intercambio de GDP por GTP. En su forma activa, Ras atrae a Raf, una serina-treonina proteinocinasa, hacia la membrana, lo que provoca su activación. Otras serina-treonina cinasas son activadas secuencialmente por fosforilación (MEK, MAPK). Otras cascadas enzimáticas cooperan o complementan la anterior. La activación de la fosfolipasa C conduce a la de proteinocinasa C, que tiene a Raf entre sus sustratos. Por otra parte, Ras puede conducir, indirectamente, a la activación de CREB, un factor de transcripción cooperativo tanto para los genes de respuesta inmediata como para los genes de respuesta tardía. b) Eventos nucleares: la cinasa de la proteína relacionada con microtúbulos y la CREB-K se translocan al núcleo donde activan factores de transcripción de genes de activación inmediata. Los productos de estos genes (gri) son, a su vez, potentes factores de transcripción para genes estructurales de respuesta tardía. Las proteínas sintetizadas son responsables finales de los procesos de crecimiento y diferenciación inducidos por las neurotrofinas. NGF: factor de crecimiento nervioso; Trk-A: tirosinocinasa A, receptor de NGF; Shc: Src homology and collagen; Grb2: Growth factor receptor-bound protein 2; SoS: Son of Sevenless; Ras: proteína G pequeña relacionada con la membrana; Raf: serina-treonina proteinocinasa; MEK: cinasa de MAPK; MAPK: cinasa de proteína relacionada con microtúbulos (MAP, Microtubule Associated Protein); PLC: fosfolipasa C; PIP_2 : fosfolípidos de membrana; IP_3 : inositol-3-fosfato; DAG: diacilglicerol; SRF: Serum Response Factor; Elk: factor de transcripción; SRE: Serum Response Element; CREB: cAMP Regulatory Element-Binding protein; Ca/CRE: Calcium and CREB Response Element; GRI: genes de respuesta inmediata; gri-RE: elemento de respuesta a los genes de respuesta inmediata; GRT: genes de respuesta tardía.

Soporte metabólico

Todos los procesos de neuroplasticidad descritos, tanto los que implican crecimiento como aquellos que tienen un carácter más funcional, implican procesos de remodelación que demandan síntesis de nuevas macromoléculas: proteínas, glicoproteínas y glicolípidos. Teóricamente, es posible que la disponibilidad limitada

de algunos precursores biosintéticos reduzca las potencialidades neuroplásticas. Revisaremos algunos de estos precursores y las evidencias que les confieren atractivo como potenciadores de neuroplasticidad.

Ácido orótico

El ácido orótico es una sustancia natural, precursora de nucleótidos de pirimidina (UMP y CMP). Las concentraciones de pirimidinonucleótidos en el cerebro de los animales adultos son muy bajas [352], lo cual podría limitar la síntesis de macromoléculas (ARN, proteínas, glicoproteínas y glicolípidos) necesarias para la expresión de procesos neuroplásticos.

Estudios *in vitro* han demostrado que una mayor provisión de nucleótidos de pirimidina estimula el desarrollo de neuroblastos [353,354]. De la misma forma, el ácido orótico acelera el crecimiento neurítico [355,356], la migración y diferenciación neuronal [357] y el desarrollo mitocondrial de las neuronas en cultivo [358].

La administración de ácido orótico favorece los procesos neuroplásticos que subyacen en el aprendizaje [359], hecho que favorece particularmente a los animales viejos [360] o pobremente capacitados [361]. Estudios complementarios demostraron que el aprendizaje realmente acelera la síntesis macromolecular y que el ácido orótico favorece estos procesos [362].

Por otra parte, el ácido orótico también beneficia la expresión de procesos de plasticidad sináptica como la LTP [361,363-365].

Dos estudios recientes sugieren que el ácido orótico podría también tener un papel neuroprotector [366,367] en modelos de lesión isquémica del SNC, circunstancia que añade interés al estudio de la posible función de esta sustancia en los procesos de neuroplasticidad, sobre todo si consideramos que esta sustancia posee efectos cardiovasculares que mejoran la tolerancia al ejercicio [368,369] y reducen el daño vascular aterosclerótico [370].

Gangliósidos

Los gangliósidos son glucoesfingolípidos anfílicos que contienen ácido siálico y se encuentran en grandes concentraciones en las membranas sinápticas [371].

In vitro los gangliósidos muestran efectos neuronotróficos, neuritogénicos [261] y neuroprotectores [372], por lo que se consideró que podrían ser beneficiosos en los procesos de reparación nerviosa central y periférica [373,374].

La administración de algunos gangliósidos mejora la memoria [375] y la LTP en animales de experimentación [376], aunque este hecho ha sido cuestionado por resultados posteriores [377]. Su administración en modelos animales mostró efectos neuroprotectores sobre neuronas dopaminérgicas nigrales, colinérgicas septales, así como serotoninérgicas y noradrenérgicas del tallo cerebral [378], tal vez las poblaciones moduladoras más importantes en la Neuropatología actual.

Los gangliósidos pueden administrarse por vía sistémica, pero no oral, pues son destruidos en el tracto digestivo. Se ha medido que sólo el 1-3% de la dosis administrada alcanza el cerebro, aunque en condiciones de trauma, la ruptura de barrera puede aumentar esta proporción [378].

Los estudios clínicos realizados no han confirmado estas expectativas [379]. A pesar de ello, los gangliósidos siguen siendo un grupo interesante por sus propiedades, aunque se requieren estudios moleculares y metabólicos [380] más profundos para una mejor evaluación de su uso como moduladores de neuroplasticidad.

Esteroides

Las hormonas esteroideas incluyen hormonas sexuales, glucocorticoides y mineralocorticoides. El mecanismo clásico de acción de estas hormonas se basa en su interacción con un receptor proteico intranuclear que desenmascara su región de unión al ADN y promueve la transcripción y síntesis de proteínas. Las hormonas esteroideas, por sus propiedades físico-químicas, atraviesan con facilidad la barrera hematoencefálica y la membrana plasmática. Sin embargo, en el cerebro, añaden a su mecanismo clásico de acción genómica efectos extragenómicos que parecen depender de receptores de membrana [381]. Existen receptores nucleares clásicos para los estrógenos en muchas zonas del sistema límbico como el hipocampo y el hipotálamo [382], aunque para su acción neuroplástica los estrógenos requieren la cooperación de receptores de membrana, entre los que se han invocado el receptor IGF-I [383] y los receptores NMDA [381,384].

Los estrógenos tienen efectos neuritogénicos que en algunas células se restringe a los axones [385]; no obstante, su acción más prominente en el hipocampo parece ser el aumento en la densidad de espinas dendríticas [386] por mecanismos dependientes de NMDA [384], que estimulan la formación de nuevas sinapsis y actúan sobre botones terminales preexistentes [387].

Los estrógenos favorecen o estimulan los mecanismos de neuroplasticidad sináptica [388] y por crecimiento [381], probablemente a través de la cascada AMPc-PKA-CREB [389].

La alta sensibilidad a los estrógenos de los mecanismos de neuroplasticidad se evidencia en la demostración de que las capacidades neuroplásticas de ratas hembras varía en diferentes etapas del ciclo estral; así, es alta cuando son elevados los niveles de estrógenos y decae rápidamente cuando éstos disminuyen [390,391].

Los glucocorticoides, las principales hormonas del estrés, han mostrado efectos contrarios y en parte antagonísticos [392]. En el modelo de lesión de corteza entorrinal, la administración de corticosterona reduce la reinervación por colateralización y sinaptogénesis reactiva [393]. Ello sucede a pesar de que los receptores para glucocorticoides son regulados negativamente por la deafferentación [394].

Los efectos antineuroplásticos de los glucocorticoides parecen acentuarse con la edad [4,395].

Recientemente se ha demostrado que puede ocurrir síntesis de esteroides en el propio tejido cerebral. Sustancias como la pregnanolona y la alopregnanolona, denominadas genéricamente neurosteroides, actúan principalmente a través de receptores de membrana. Actualmente se evalúan sus posibles acciones neuroplásticas [296].

Factores ambientales y estilo de vida

Desde hacía tiempo se sospechaba que la privación de estimulación y actividad retarda y compromete el desarrollo del sistema nervioso [396], pero sólo en las décadas recientes se han comenzado a entender los mecanismos y a conocer cómo estos mismos factores pueden operar en la recuperación de funciones del cerebro adulto dañado.

Las primeras evidencias experimentales de plasticidad dependiente del uso o la actividad provienen de los trabajos clásicos de Rosenzweig y Bennet en los años 60. Ratas criadas en ambientes enriquecidos (cajas grandes con abundantes laberintos, escaleras y objetos) mostraron tener una corteza cerebral más gruesa, más contactos sinápticos, mayor número de dendritas y espinas dendríticas [397]. Este experimento fue el inicial

impulso trascendente de un gran número de trabajos experimentales para comprender qué, cuánto y cómo la actividad, la estimulación y la experiencia contribuyen a la formación funcional del sistema nervioso.

Estudios ulteriores demostraron que la vida en ambientes enriquecidos incrementa la talla de las neuronas, la ramificación de sus dendritas y la densidad de las espinas dendríticas, el número de sinapsis por neurona y el tamaño de los contactos sinápticos, así como la vascularización tisular y la talla de astrocitos, oligodendrocitos y las mitocondrias [398].

La permanencia por tiempos más o menos prolongados en ambientes enriquecidos mejora las capacidades de memoria y la actividad de la PKC [399], protege contra las consecuencias de lesiones vasculares provocadas y mejora la expresión de otras proteínas de plasticidad como el BDNF y la PK-II [400]. También se ha comprobado que este procedimiento incrementa el número de espinas dendríticas múltiples en el núcleo caudado [156] y reduce algunas consecuencias negativas del envejecimiento, como la reducción del número de sinapsis en diversas áreas cerebrales [401] y la neurotoxicidad por glucocorticoides [402]. La función glial, expresada en una mayor arborización astrocítica, también se beneficia [398]. La estancia en ambientes enriquecidos también induce cambios neuroplásticos en el cerebro de ratas viejas [398]. Recientemente se ha demostrado que vivir en ambientes enriquecidos estimula la neurogénesis en ratas viejas [57].

Los mecanismos de neuroplasticidad, que conducen a la recuperación por lesión del SNC, también se benefician de ambientes enriquecidos. Tanto en animales jóvenes como adultos, los mecanismos de soporte trófico de tipo glial o por medio de factores neurotróficos (FGF, NGF) muestran mejorías por exposición a ambientes enriquecidos, hecho que se acompaña de un incremento en los procesos de colateralización y sinaptogénesis reactiva en áreas vecinas a la lesión [398].

Es interesante destacar que cambios similares a los que provoca el ambiente enriquecido se han demostrado en animales sometidos a entrenamiento cognitivo o en la recuperación después de daño cerebral [156]; una evidencia de la universalidad de los mecanismos de neuroplasticidad.

Para una rata de laboratorio, vivir en un ambiente enriquecido significa, sobre todo, mayor estimulación sensorial, motora y cognitiva. Todos los experimentos que utilizan el paradigma de ambiente enriquecido vienen a significar que la estimulación neural de cualquier tipo, en cualquier etapa de la vida, estimula mecanismos de plasticidad importantes para la maduración morfofuncional del sistema y su reparación en caso de daño.

En efecto, los mecanismos de plasticidad son modulables por estimulación sensorial. La LTP en hipocampo y corteza piriforme es modulada por estímulos olfatorios [403]. Otras formas de plasticidad funcional pueden modificarse transitoriamente en la corteza somatosensorial por asociación de estímulos táctiles [404]. La LTP está exacerbada y la LTD disminuida en la corteza visual de ratas privadas de luz, situación que puede revertirse después de sólo dos días de exposición a la luz [405]; esta circunstancia también modifica la expresión de proteínas de plasticidad como AP-1 y zif-268 [406]. Cambios equivalentes se describen en áreas motoras [407] y ya algunos de estos conocimientos comienzan a comprobarse en la clínica [408]. El uso determina el número, morfología y tipo de sinapsis, aun con períodos relativamente cortos de activación [409].

La participación exacta de las neurotrofinas está por establecerse, pero existen evidencias de que su producción, en diversas

estructuras nerviosas centrales y periféricas, es modificada por estímulos fisiológicos [410].

Nuevamente se demuestra la participación de mecanismos similares a los anteriormente descritos en la plasticidad inducida por la experiencia, incluida la mediación de receptores NMDA [411], que confirman la universalidad de los mecanismos de neuroplasticidad.

La utilización en la práctica clínica de modelos de estimulación requiere, sin embargo, un mayor conocimiento de los mejores patrones de estimulación, ya que comienza a comprenderse que ligeros cambios en el modo de estimulación [412] y el modo de vida [413] pueden tener efectos muy diferentes en los procesos de plasticidad.

Actividad y ejercicio físico

El ejercicio físico es fuente de desarrollo no sólo del cuerpo, sino que también el cerebro y la mente pueden beneficiarse de la actividad física mediante la inducción de cambios plásticos. Después de lo comentado en el epígrafe anterior este acerto puede parecer obvio. Ciertamente es que la realización de cualquier tarea motora genera patrones de estimulación sensorial propioceptiva y puede ser fuente de modulación neuroplástica en áreas motoras y somatosensoriales. En este caso, nos referimos a efectos más generales e inespecíficos de la actividad física, provocados no sólo por patrones de estimulación sensorial, sino por la interacción de cambios físicos, hormonales y otros que mejoran o potencian los procesos involucrados en las remodelaciones neuroplásticas.

La actividad física, por ejemplo, mejoró la memoria y paralelamente la actividad de PKC en el hipocampo de ratones entrenados en una estera rodante durante ocho semanas [414]. También la expresión de neurotrofinas, en particular el BDNF y su receptor, aumentan en el hipocampo por el ejercicio [415,416]; la producción de nuevas células nerviosas y la LTP en el giro dentado se benefician por el ejercicio [417], y el entrenamiento

repetido ayuda a reparar los daños por lesiones provocadas en ratas [3].

Es posible que la estimulación específica de áreas cerebrales y el ejercicio físico difieran en algunos aspectos con relación a los cambios neuroplásticos que uno y otro producen. Hasta hace sólo unos años, se consideraba que la actividad física modificaba sobre todo la vascularización cerebral y no la densidad sináptica [407,418,419]. Los experimentos citados en el párrafo anterior están ayudando a cambiar esa opinión, pero la combinación de actividad física y estimulación sensorial y motora específica sigue pareciendo la más eficaz para la inducción de procesos de remodelación neuroplástica del cerebro dañado.

CONCLUSIONES

Los mecanismos de la neuroplasticidad son universales. En la escala filogenética de las especies animales, mecanismos basados en patrones de activación y eventos moleculares similares o idénticos participan tanto en la construcción del sistema nervioso durante el desarrollo embrionario, como en su reconstrucción durante la vida posnatal.

Esta reconstrucción puede darse por medio de sutiles modificaciones funcionales, por ejemplo en el aprendizaje, o mediante procesos de crecimiento axonal, dendrítico y la formación de nuevas sinapsis en respuesta al daño.

En esencia, plasticidad por crecimiento y plasticidad funcional no son tampoco procesos separados. Como se ilustra en el caso de la LTP, estos procesos operan formando un *continuum* que va desde modificaciones moleculares en sinapsis existentes hasta la formación de nuevas sinapsis, según el tipo de estímulo aplicado.

La neuroplasticidad comienza a ganar un lugar en los esquemas terapéuticos más modernos de la Neurología Restaurativa [420]. Conocer mejor sus mecanismos y modulación por agentes físicos y químicos será una contribución de enorme utilidad para hacer de ellos un uso más eficiente y eficaz.

BIBLIOGRAFÍA

1. Álvarez-Buylla A, Lois C. Neuronal stem cells in the brain of adult vertebrates. *Stem Cells* (Dayt) 1995; 13: 263-72.
2. Von Steinbüchel N, Pöppel E. Domains of rehabilitation: a theoretical perspective. *Behav Brain Res* 1993; 56: 1-10.
3. Nitta A, Hayashi K, Hasegawa T, Nabeshima T. Development of plasticity of brain function with repeated trainings and passage of time after basal forebrain lesions in rats. *J Neural Transm Gen Sect* 1993; 93: 46.
4. Fuxe K, Díaz R, Cintra A, Bhatnagar M, Tinner B, Gustafsson JA, et al. On the role of glucocorticoid receptors in brain plasticity. *Cell Mol Neurobiol* 1996; 16: 239-58.
5. Hebb DO. *The organization of behaviour*. New York: Wiley & Sons; 1949.
6. Spatz HC. Hebb's concept of synaptic plasticity and neuronal cell assemblies. *Behav Brain Res* 1996; 78: 3-7.
7. Matthies H. The biochemical basis of learning and memory. *Life Sci* 1974; 15: 2017-31.
8. Olson L. Regeneration in the adult central nervous system: experimental repair strategies. *Nat Med* 1997; 3: 1329-35.
9. Deller T, Frotscher M. Lesion-induced plasticity of central neurons: sprouting of single fibers in the rat hippocampus after unilateral entorhinal cortex lesion. *Prog Neurobiol* 1997; 53: 687-727.
10. Kapfhammer JP, Schwab ME. Inverse patterns of myelination and GAP-43 expression in the adult CNS: neurite growth inhibitors as regulators of neuronal plasticity. *J Comp Neurol* 1994; 340: 194-206.
11. Kapfhammer JP, Schwab ME. Increased expression of the growth-associated protein GAP-43 in the myelin-free rat spinal cord. *Eur J Neurosci* 1994; 6: 403-11.
12. Colman BW, Earley EM, Shahar A, Dudek FE, Ide CF. Factors influencing mossy fiber collateral sprouting in organotypic slice cultures of neonatal mouse hippocampus. *J Comp Neurol* 1995; 362: 209-22.
13. McNamara RK, Routtenberg A. NMDA receptor blockade prevents kainate induction of protein F1/GAP-43 mRNA in hippocampal granule cells and subsequent mossy fiber sprouting in the rat. *Mol Brain Res* 1995; 33: 22-8.
14. Bendotti C, Pende M, Guglielmetti F, Samanin R. Cycloheximide inhibits kainic acid-induced GAP-43 mRNA in dentate granule cells in rats. *Neuroreport* 1996; 7: 2539-42.
15. Benowitz LI, Routtenberg A. A membrane phosphoprotein associated with neural development, axonal regeneration, phospholipid metabolism, and synaptic plasticity. *Trends Neurosci* 1987; 10: 527-32.
16. Benowitz LI, Routtenberg A. GAP-43: an intrinsic determinant of neuronal development and plasticity. *Trends Neurosci* 1997; 20: 84-91.
17. Clayton GH, Mahalik TJ, Finger TE. Expression of GAP-43 mRNA in normally developing and transplanted neurons from the rat ventral mesencephalon. *J Comp Neurol* 1994; 347: 470-80.
18. Bendotti C, Pende M, Samanin R. Expression of GAP-43 in the granule cells of rat hippocampus after seizure-induced sprouting of mossy fibres: in situ hybridization and immunocytochemical studies. *Eur J Neurosci* 1994; 6: 509-15.
19. Bendotti C, Baldessari S, Samanin R, De Biasi S. Ultrastructural immunolocalization of GAP-43 in the sprouted mossy fibres of kainic acid lesioned rats. *Neuroreport* 1994; 5: 2645-8.
20. Routtenberg A. Protein kinase C activation leading to protein F1 phosphorylation may regulate synaptic plasticity by presynaptic terminal growth. *Behav Neural Biol* 1985; 44: 186-200.
21. Boschert U, O'Shaughnessy C, Dickinson R, Tessari M, Bendotti C, Catsicas S, et al. Developmental and plasticity-related differential expression of two SNAP-25 isoforms in the rat brain. *J Comp Neurol* 1996; 367: 177-93.
22. Melloni RH Jr, De Gennaro LJ. Temporal onset of synapsin I gene expression coincides with neuronal differentiation during the development of the nervous system. *J Comp Neurol* 1994; 342: 449-62.

23. Melloni RH Jr, Apostolides PJ, Hamos JE, De Gennaro LJ. Dynamics of synapsin I gene expression during the establishment and restoration of functional synapses in the rat hippocampus. *Neuroscience* 1994; 58: 683-703.
24. Jensen MB, González B, Castellano B, Zimmer J. Microglial and astroglial reactions to anterograde axonal degeneration: a histochemical and immunocytochemical study of the adult rat fascia dentata after entorhinal perforant path lesions. *Exp Brain Res* 1994; 98: 245-60.
25. Fagan AM, Gage FH. Mechanisms of sprouting in the adult central nervous system: cellular responses in areas of terminal degeneration and reinnervation in the rat hippocampus. *Neuroscience* 1994; 58: 705-25.
26. Beck KD, Lamballe F, Klein R, Barbacid M, Schaufwecker PE, McNeill TH, et al. Induction of noncatalytic TrkB neurotrophin receptors during axonal sprouting in the adult hippocampus. *J Neurosci* 1993; 13: 4001-14.
27. Cremer H, Chazal G, Carleton A, Goridis C, Vincent JD, Lledo PM. Long-term but not short-term plasticity at mossy fiber synapses is impaired in neural cell adhesion molecule-deficient mice. *Proc Natl Acad Sci U S A* 1998; 95: 13242-7.
28. Styren SD, Lagenaur CF, Miller PD, DeKosky ST. Rapid expression and transport of embryonic N-CAM in dentate gyrus following entorhinal cortex lesion: ultrastructural analysis. *J Comp Neurol* 1994; 349: 486-92.
29. Miller PD, Styren SD, Lagenaur CF, DeKosky ST. Embryonic neural cell adhesion molecule (N-CAM) is elevated in the denervated rat dentate gyrus. *J Neurosci* 1994; 14: 4217-25.
30. Jucker M, Mondadori C, Mohajeri H, Bartsch U, Schachner M. Transient upregulation of NCAM mRNA in astrocytes in response to entorhinal cortex lesions and ischemia. *Mol Brain Res* 1995; 28: 149-56.
31. Frotscher M, Heimrich B, Deller T. Sprouting in the hippocampus is layer-specific. *Trends Neurosci* 1997; 20: 218-23.
32. Frotscher M, Heimrich B, Deller T, Nitsch R. Understanding the cortex through the hippocampus: lamina-specific connections of the rat hippocampal neurons. *J Anat* 1995; 187: 539-45.
33. Frotscher M, Heimrich B. Formation of layer-specific fiber projections to the hippocampus in vitro. *Proc Natl Acad Sci U S A* 1993; 90: 10400-3.
34. Peterson GM. Sprouting of central noradrenergic fibers in the dentate gyrus following combined lesions of its entorhinal and septal afferents. *Hippocampus* 1994; 4: 635-48.
35. Jackisch R, Neufang B, Hertting G, Jeltsch H, Kelche C, Will B, et al. Sympathetic sprouting: time course of changes of noradrenergic, cholinergic, and serotonergic markers in the denervated rat hippocampus. *J Neurochem* 1995; 65: 329-37.
36. Deller T, Frotscher M, Nitsch R. Morphological evidence for the sprouting of inhibitory commissural fibers in response to the lesion of the excitatory entorhinal input to the rat dentate gyrus. *J Neurosci* 1995; 15: 6868-78.
37. Ramírez JJ, McQuilkin M, Carrigan T, MacDonald K, Kelley MS. Progressive entorhinal cortex lesions accelerate hippocampal sprouting and spare spatial memory in rats. *Proc Natl Acad Sci U S A* 1996; 93: 15512-7.
38. Kugler P, Schleicher A, Zilles K, Horváth E. Acetylcholinesterase activity and post-lesional plasticity in the hippocampus of young and aged rats. *Neuroscience* 1993; 55: 91-103.
39. Alonso JR, Sang H, Amaral DG. Cholinergic innervation of the primate hippocampal formation. II. Effects of fimbria/fornix transection. *J Comp Neurol* 1996; 375: 527-51.
40. Anthes DL, LeBoutillier JC, Petit TL. Structure and plasticity of newly formed adult synapses: a morphometric study in the rat hippocampus. *Brain Res* 1993; 626: 50-62.
41. Hoff SF. Lesion-induced transneuronal plasticity in the adult rat hippocampus. *Neuroscience* 1986; 19: 1227-33.
42. Hamori J. Morphological plasticity of postsynaptic neurones in reactive synaptogenesis. *J Exp Biol* 1990; 153: 251-60.
43. Burry RW. Protein synthesis requirement for the formation of synaptic elements. *Brain Res* 1985; 344: 109-19.
44. Gazzaley AH, Benson DL, Huntley GW, Morrison JH. Differential subcellular regulation of NMDAR1 protein and mRNA in dendrites of dentate gyrus granule cells after perforant path transection. *J Neurosci* 1997; 17: 2006-17.
45. Steward O. The process of reinnervation in the dentate gyrus of adult rats: gene expression by neurons during the period of lesion-induced growth. *J Comp Neurol* 1995; 359: 391-411.
46. Díez-Guerra FJ, Ávila J. An increase in phosphorylation of microtubule-associated protein 2 accompanies dendrite extension during the differentiation of cultured hippocampal neurones. *Eur J Biochem* 1995; 227: 68-77.
47. Díez-Guerra FJ, Ávila J. MAP2 phosphorylation parallels dendrite arborization in hippocampal neurones in culture. *Neuroreport* 1993; 4: 419-22.
48. Álvarez-Buylla A, Ling CY, Yu WS. Contribution of neurons born during embryonic, juvenile, and adult life to the brain of adult canaries: regional specificity and delayed birth of neurons in the song-control nuclei. *J Comp Neurol* 1994; 347: 233-48.
49. Bottjer SW, Arnold AP. Developmental plasticity in neural circuits for a learned behavior. *Annu Rev Neurosci* 1997; 20: 459-81.
50. Seki T, Arai Y. Age-related production of new granule cells in the adult dentate gyrus. *Neuroreport* 1995; 6: 2479-82.
51. Palmer TD, Takahashi J, Gage FH. The adult rat hippocampus contains primordial neural stem cells. *Mol Cell Neurosci* 1997; 8: 389-404.
52. Brezun JM, Daszuta A. Depletion in serotonin decreases neurogenesis in the dentate gyrus and the subventricular zone of adult rats. *Neuroscience* 1999; 89: 999-1002.
53. Madeira MD, Cadete-Leite A, Andrade JP, Paula-Barbosa MM. Effects of hypothyroidism upon the granular layer of the dentate gyrus in male and female adult rats: a morphometric study. *J Comp Neurol* 1991; 314: 171-86.
54. Parent JM, Yu TW, Leibowitz RT, Geshwind DH, Sloviter RS, Lowenstein DH. Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci* 1997; 17: 3727-38.
55. Parent JM, Janumpalli S, McNamara JO, Lowenstein DH. Increased dentate granule cells neurogenesis following amygdala kindling in the adult rat. *Neurosci Lett* 1998; 247: 9-12.
56. Seki T, Arai Y. Highly polysialylated neural cell adhesion molecule (NCAM-H) is expressed by newly generated granule cells in the dentate gyrus of the adult rat. *J Neurosci* 1993; 13: 2351-8.
57. Kempermann G, Kuhn HG, Gage FH. Experience-induced neurogenesis in the senescent dentate gyrus. *J Neurosci* 1998; 18: 3206-12.
58. Lim DA, Fishell GJ, Álvarez-Buylla A. Postnatal mouse subventricular zone neuronal precursors can migrate and differentiate within multiple levels of the developing neuraxis. *Proc Natl Acad Sci U S A* 1997; 94: 14832-6.
59. Brewer GJ. Regeneration and proliferation of embryonic and adult rat hippocampal neurons in culture. *Exp Neurol* 1999; 159: 237-47.
60. Eckenhoff MF, Rakic P. Nature and fate of proliferative cells in the hippocampal dentate gyrus during the life span of the rhesus monkey. *J Neurosci* 1988; 8: 2729-47.
61. Matthies H. [Cellular mechanisms of learning processes and the shaping of memory]. *Z Psychol* 1976; 184: 308-28.
62. Matthies H. [Neurobiological principles of higher nervous activity]. *Z Arztl Fortbild (Jena)* 1973; 67: 316-9.
63. Matthies H. [Principles of neuronal information storage. Theories, methodology, experiments]. *Z Psychol* 1986; 194: 285-92.
64. Matthies H. From molecular mechanisms to behavior. *Activ Nerv Suppl (Prague)* 1988; 30: 2-17.
65. Matthies H. In search of cellular mechanisms of memory. *Prog Neurobiol* 1989; 32: 277-349.
66. Shaw CA, Lanius RA, Van den Doel K. The origin of synaptic neuroplasticity: crucial molecules or a dynamical cascade. *Brain Res Rev* 1994; 19: 241-63.
67. López Planes J, Almaguer Melian W, Jas García J, Bergado Rosado J. Influencia de la frecuencia de estimulación sobre procesos de plasticidad sináptica en el giro dentado de la rata. *Arch Neurocién (Mex)* 1999; 4: 9-20.
68. Kamiya H, Zucker RS. Residual Ca^{2+} and short-term synaptic plasticity. *Nature* 1994; 371: 603-6.
69. Rosahl TW, Geppert M, Spillane D, Herz J, Hammer RE, Malenka RC, et al. Short-term synaptic plasticity is altered in mice lacking synapsin I. *Cell* 1993; 75: 661-70.
70. Bliss TV, Gardner-Medwin AR. Long-lasting potentiation of synaptic transmission in the dentate area of the unanaesthetized rabbit following stimulation of the perforant path. *J Physiol* 1973; 232: 357-74.
71. Bliss TV, Lomo T. Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol* 1973; 232: 331-56.
72. Debanne D, Gähwiler BH, Thompson SM. Bidirectional associative plasticity of unitary CA3-CA1 EPSPs in the rat hippocampus in vitro. *J Neurophysiol* 1997; 77: 2851-5.
73. Coussens CM, Teyler TJ. Protein kinase and phosphatase activity regulate the form of synaptic plasticity expressed. *Synapse* 1996; 24: 97-103.
74. Malenka RC, Nicoll RA. NMDA-receptor-dependent synaptic plasticity: multiple forms and mechanisms. *Trends Neurosci* 1993; 16: 521-7.
75. Bernard CL, Hirsch JC, Khazipov R, Ben-Ari Y, Gozlan H. Redox modulation of synaptic responses and plasticity in rat CA1 hippocampal neurons. *Exp Brain Res* 1997; 113: 343-52.
76. Bear MF, Malenka RC. Synaptic plasticity: LTP and LTD. *Curr Opin Neurobiol* 1994; 4: 389-99.
77. Mulkey RM, Herron CE, Malenka RC. An essential role for protein phosphatases in hippocampal long-term depression. *Science* 1993; 261: 1051-5.

78. Christie BR, Kerr DS, Abraham WC. Flip side of synaptic plasticity: long-term depression mechanisms in the hippocampus. *Hippocampus* 1994; 4: 127-35.
79. Michaelis EK. Molecular biology of glutamate receptors in the central nervous system and their role in excitotoxicity, oxidative stress and aging. *Prog Neurobiol* 1998; 54: 369-415.
80. Ozawa S, Kamiya H, Tsuzuki K. Glutamate receptors in the mammalian central nervous system. *Prog Neurobiol* 1998; 54: 581-618.
81. Collingridge GL. Long-term potentiation in the hippocampus: mechanism of initiation and modulation by neurotransmitters. *Trends Pharmacol Sci* 1985; 407-11.
82. Morris RGM, Anderson E, Lynch GS, Baudry M. Selective impairment of learning and blockade of long-term potentiation by an N-methyl-D-aspartate receptor antagonist, AP5. *Nature* 1986; 319: 774-6.
83. Anwyl R. Synaptic plasticity: a molecular switch for memory. *Curr Biol* 1994; 4: 854-6.
84. Bashir ZI, Collingridge GL. Synaptic plasticity: long-term potentiation in the hippocampus. *Curr Opin Neurobiol* 1992; 2: 328-35.
85. Balschun D, Manahan-Vaughan D, Wagner T, Behnisch T, Reymann KG, Wetzel W. A specific role for group I mGluRs in hippocampal LTP and hippocampus-dependent spatial learning. *Learn Mem* 1999; 6: 138-52.
86. Richter-Levin G, Errington ML, Maegawa H, Bliss TVP. Activation of metabotropic glutamate receptors is necessary for long-term potentiation in the dentate gyrus and for spatial learning. *Neuropharmacology* 1994; 33: 853-7.
87. Zamanillo D, Sprengel R, Hvalby O, Jensen V, Burnashev N, Rozov A, et al. Importance of AMPA receptors for hippocampal synaptic plasticity but not for spatial learning. *Science* 1999; 284: 1805-11.
88. Asztely F, Gustafsson B. Ionotropic glutamate receptors. Their possible role in the expression of hippocampal synaptic plasticity. *Mol Neurobiol* 1996; 12: 1-11.
89. Segal M. Calcium and neuronal plasticity. *Isr J Med Sci* 1993; 29: 543-8.
90. Pelletier MR, Hablitz JJ. Tetraethylammonium-induced synaptic plasticity in rat neocortex. *Cereb Cortex* 1996; 6: 771-80.
91. Teyler TJ, Cavus I, Coussens C, DiScenna P, Grover L, Lee YP, et al. Multideterminant role of calcium in hippocampal synaptic plasticity. *Hippocampus* 1994; 4: 623-34.
92. Teyler TJ, Cavus I, Coussens C. Synaptic plasticity in the hippocampal slice: functional consequences. *J Neurosci Methods* 1995; 59: 11-7.
93. Dosemeci A, Choi C. Ca^{2+} -independent autophosphorylation of postsynaptic density-associated Ca^{2+} /calmodulin-dependent protein kinase. *Neurochem Res* 1997; 22: 1151-7.
94. Fukunaga K, Miyamoto E. Current studies on a working model of CaM kinase II in hippocampal long-term potentiation and memory. *Jpn J Pharmacol* 1999; 79: 7-15.
95. Fukunaga K, Muller D, Miyamoto E. CaM kinase II in long-term potentiation. *Neurochem Int* 1996; 28: 343-58.
96. Giese KP, Fedorov NB, Filipkowski RK, Silva AJ. Autophosphorylation at Thr²⁸⁶ of the α calcium-calmodulin kinase II in LTP and learning. *Science* 1998; 279: 870-3.
97. Reymann KG, Brödemann R, Kase H, Matthies H. Inhibitors of calmodulin and protein kinase C block different phases of hippocampal long-term potentiation. *Brain Res* 1988; 461: 388-92.
98. Stevens CF, Tonegawa S, Wang Y. The role of calcium-calmodulin kinase II in three forms of synaptic plasticity. *Curr Biol* 1994; 4: 687-93.
99. Silva AJ, Stevens CF, Tonegawa S, Wang Y. Deficient hippocampal long-term potentiation in α -calcium-calmodulin kinase II mutant mice. *Science* 1992; 257: 201-6.
100. Leonard AS, Lim IA, Hemsworth DE, Horne MC, Hell JW. Calcium/calmodulin-dependent protein kinase II is associated with the N-methyl-D-aspartate receptor. *Proc Natl Acad Sci U S A* 1999; 96: 3239-44.
101. Derkach V, Barria A, Soderling TR. Ca^{2+} /calmodulin-kinase II enhances channel conductance of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionate type glutamate receptors. *Proc Natl Acad Sci U S A* 1999; 96: 3269-74.
102. Lovinger DM, Wong KL, Murakami K, Routtenberg A. Protein kinase C inhibitors eliminate hippocampal long-term potentiation. *Brain Res* 1987; 436: 177-83.
103. Reymann KG, Frey U, Jork R, Matthies H. Polymyxin B, an inhibitor of protein kinase C, prevents the maintenance of synaptic long-term potentiation in hippocampal CA1 neurons. *Brain Res* 1988; 440: 305-14.
104. Colley PA, Sheu FS, Routtenberg A. Inhibition of protein kinase C blocks two components of LTP persistence, leaving initial potentiation intact. *J Neurosci* 1990; 10: 3353-60.
105. Malenka RC, Madison DV, Nicoll RA. Potentiation of synaptic transmission in the hippocampus by phorbol esters. *Nature* 1986; 321: 175-7.
106. Asztely F, Hanse E, Gustafsson B. The early decay of long-term potentiation in the hippocampal CA1 region in vitro is reduced by activators of protein kinase C. *Brain Res* 1990; 521: 355-8.
107. Klann E, Chen SJ, Sweatt JD. Mechanism of protein kinase C activation during the induction and maintenance of long-term potentiation probed using a selective peptide substrate. *Proc Natl Acad Sci U S A* 1993; 90: 8337-41.
108. Klann E, Chen SJ, Sweatt JD. Increased phosphorylation of a 17-kDa protein kinase C substrate (P17) in long-term potentiation. *J Neurochem* 1992; 58: 1576-9.
109. Lovinger DM, Routtenberg A. Synapse-specific protein kinase C activation enhances maintenance of long-term potentiation in rat hippocampus. *J Physiol* 1988; 400: 321-33.
110. Linden DJ, Routtenberg A. The role of protein kinase C in long-term potentiation: a testable model. *Brain Res Brain Res Rev* 1989; 14: 279-96.
111. Angenstein F, Hirschfelder M, Staak S. Activation of metabotropic glutamate receptors increases endogenous protein kinase C substrate phosphorylation in adult hippocampal slices. *Brain Res* 1997; 745: 46-54.
112. Chen SJ, Sweatt JD, Klann E. Enhanced phosphorylation of the postsynaptic protein kinase C substrate RC3/neurogranin during long-term potentiation. *Brain Res* 1997; 749: 181-7.
113. Cohen RW, Margulies JE, Coulter PM II, Watson JB. Functional consequences of expression of the neuron-specific, protein kinase C substrate RC3 (neurogranin) in *Xenopus* oocytes. *Brain Res* 1993; 627: 147-52.
114. Fedorov NB, Pasinelli P, Oestreicher AB, DeGraan PNE, Reymann KG. Antibodies to postsynaptic PKC substrate neurogranin prevent long-term potentiation in hippocampal CA1 neurons. *Eur J Neurosci* 1995; 7: 819-22.
115. Pontzer NJ, Chandler LJ, Stevens BR, Crews FT. Receptors, phosphoinositide hydrolysis and plasticity of nerve cells. *Prog Brain Res* 1990; 86: 221-5.
116. Huang YY, Kandel ER. Recruitment of long-lasting and protein kinase A-dependent long-term potentiation in the CA1 region of hippocampus requires repeated tetanization. *Learn Mem* 1994; 1: 74-82.
117. Xia Z, Choi EJ, Storm DR, Blazynski C. Do the calmodulin-stimulated adenylyl cyclases play a role in neuroplasticity? *Behav Brain Sci* 1995; 18: 429-40.
118. Chetkovich DM, Sweatt JD. NMDA receptor activation increases cyclic AMP in area CA1 of the hippocampus via calcium/calmodulin stimulation of adenylyl cyclase. *J Neurochem* 1993; 61: 1933-42.
119. Matthies H, Reymann KG. Protein kinase A inhibitors prevent the maintenance of hippocampal long-term potentiation. *Neuroreport* 1993; 4: 712-4.
120. Abel T, Nguyen PV, Barad M, Deuel TAS, Kandel ER. Genetic demonstration of a role for PKA in the late phase of LTP and in hippocampus-based long-term memory. *Cell* 1997; 88: 615-26.
121. Brandon EP, Zhuo M, Huang YY, Qi M, Gerhold KA, Burton KA, et al. Hippocampal long-term depression and depotentiation are defective in mice carrying a targeted disruption of the gene encoding the R1b subunit of cAMP-dependent protein kinase. *Proc Natl Acad Sci U S A* 1995; 92: 8851-5.
122. Qi M, Zhuo M, Skolhegg BS, Brandon EP, Kandel ER, McKnight GS, et al. Impaired hippocampal plasticity in mice lacking the Cb β catalytic subunit of cAMP-dependent protein kinase. *Proc Natl Acad Sci U S A* 1996; 93: 1571-6.
123. Frey U, Huang YY, Kandel ER. Effects of cAMP simulate a late stage of LTP in hippocampal CA1 neurons. *Science* 1993; 260: 1661-4.
124. Hell JW, Yokoyama CT, Breeze LJ, Chavkin C, Catterall WA. Phosphorylation of presynaptic and postsynaptic calcium channels by cAMP-dependent protein kinase in hippocampal neurons. *EMBO J* 1995; 14: 3036-44.
125. Nguyen PV, Kandel ER. A macromolecular synthesis-dependent late phase of long-term potentiation requiring cAMP in the medial perforant pathway of rat hippocampal slices. *J Neurosci* 1996; 16: 3189-98.
126. Nguyen PV, Kandel ER. Brief theta-burst stimulation induces a transcription-dependent late phase of LTP requiring cAMP in area CA1 of the mouse hippocampus. *Learn Mem* 1997; 4: 230-43.
127. Impey S, Mark M, Villacres EC, Poser S, Chavkin C, Storm DR. Induction of CRE-mediated gene expression by stimuli that generate long-lasting LTP in area CA1 of the hippocampus. *Neuron* 1996; 16: 973-82.
128. Schulman H. Protein phosphorylation in neuronal plasticity and gene expression. *Curr Opin Neurobiol* 1995; 5: 375-81.
129. Roche KW, Tingley WG, Huganir RL. Glutamate receptor phosphorylation and synaptic plasticity. *Curr Opin Neurobiol* 1994; 4: 383-8.
130. Frey U, Krug M, Reymann KG, Matthies H. Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Res* 1988; 452: 57-65.
131. Krug M, Lössner B, Ott T. Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 1984; 13: 39-42.
132. Otani S, Abraham WC. Inhibition of protein synthesis in the dentate gyrus, but not the entorhinal cortex, blocks maintenance of long-term potentiation in rats. *Neurosci Lett* 1989; 106: 175-80.

133. Frey U, Krug M, Brödemann R, Reymann K, Matthies H. Long-term potentiation induced in dendrites separated from rat's CA1 pyramidal somata does not establish a late phase. *Neurosci Lett* 1989; 97: 135-9.
134. Matthies H, Frey U, Reymann K, Krug M, Jork R, Schroeder H. Different mechanisms and multiple stages of LTP. *Adv Exp Med Biol* 1990; 268: 359-68.
135. Matthies H, Reymann KG, Krug M, Frey U, Loessner B, Popov N. Multiple stages of posttetanic LTP and memory. In Rahmann, ed. *Fundamentals of memory formation: neuronal plasticity and brain function*. Stuttgart: Gustav Fischer Verlag; 1989; 395-6.
136. Frey S, Schweigert C, Krug M, Lossner B. Long-term potentiation induced changes in protein synthesis of hippocampal subfields of freely moving rats: time-course. *Biomed Biochim Acta* 1991; 50: 1231-40.
137. Lössner B, Schweigert C, Pchalek V, Krug M, Frey S, Matthies H. Training -and LTP- induced changes of protein synthesis in rat hippocampus. *Neurosci Suppl* 1987; 22: 9512-2.
138. Bishop JM, Capobianco AJ, Doyle HJ, Finney RE, McMahon M, Robbins SM, et al. Proto-oncogenes and plasticity in cell signaling. *Cold Spring Harbor Symp Quant Biol* 1994; 59: 165-72.
139. Roberts LA, Higgins MJ, O'Shaughnessy CT, Stone TW, Morris BJ. Changes in hippocampal gene expression associated with the induction of long-term potentiation. *Mol Brain Res* 1996; 42: 123-7.
140. Abraham WC, Dragunow M, Tate WP. The role of immediate early genes in the stabilization of long-term potentiation. *Mol Neurobiol* 1991; 5: 297-314.
141. Chineastra P, Leinekugel X, Ben Ari Y, Pollard H. Use of hippocampal slices to study mRNA changes in relation to synaptic plasticity. *Neuroreport* 1994; 5: 1461-5.
142. Inokuchi K, Murayama A, Ozawa F. mRNA differential display reveals Krox-20 as a neural plasticity-regulated gene in the rat hippocampus. *Biochem Biophys Res Commun* 1996; 221: 430-6.
143. Nedivi E, Hevroni D, Naot D, Israeli D, Citri Y. Numerous candidate plasticity-related genes revealed by differential cDNA cloning. *Nature* 1993; 363: 718-22.
144. Vogt K, Streit J, Dityatev A, Lüscher HR. Synaptic plasticity in dissociated hippocampal cultures: pre- and postsynaptic contributions. *Eur J Neurosci* 1997; 9: 1078-82.
145. Desmond NL, Weinberg RJ. Enhanced expression of AMPA receptor protein at perforated axospinous synapses. *Neuroreport* 1998; 9: 857-60.
146. Frey U, Morris RGM. Synaptic tagging and long-term potentiation. *Nature* 1997; 385: 533-6.
147. Frey U, Morris RGM. Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 1997; 21: 181-8.
148. Geinisman Y, Morrell F, De Toledo-Morrell L. Perforated synapses on double-headed dendritic spines: a possible structural substrate of synaptic plasticity. *Brain Res* 1989; 480: 326-9.
149. Geinisman Y. Perforated axospinous synapses with multiple, completely partitioned transmission zones: probable structural intermediates in synaptic plasticity. *Hippocampus* 1993; 3: 417-34.
150. Rusakov DA, Richter-Levin G, Stewart MG, Bliss TV. Reduction in spine density associated with long-term potentiation in the dentate gyrus suggests a spine fusion-and-branching model of potentiation. *Hippocampus* 1997; 7: 489-500.
151. Fields RD, Itoh K. Neural cell adhesion molecules in activity-dependent development and synaptic plasticity. *Trends Neurosci* 1996; 19: 473-80.
152. Muller D, Wang C, Skibo G, Toni N, Cremer H, Calaora V, et al. PSA-NCAM is required for activity-induced synaptic plasticity. *Neuron* 1996; 17: 413-22.
153. Roberts LA, Morris BJ, O'Shaughnessy CT. Involvement of two isoforms of SNAP-25 in the expression of long-term potentiation in the rat hippocampus. *Neuroreport* 1998; 9: 33-6.
154. Steffensen SC, Wilson MC, Henriksen SJ. Coloboma contiguous gene deletion encompassing SNAP alters hippocampal plasticity. *Synapse* 1996; 22: 281-9.
155. Hoffman KB, Pinkstaff JK, Gall CM, Lynch G. Seizure induced synthesis of fibronectin is rapid and age dependent: implications for long-term potentiation and sprouting. *Brain Res* 1998; 812: 209-15.
156. Klintsova AY, Greenough WT. Synaptic plasticity in cortical systems. *Curr Opin Neurobiol* 1999; 9: 203-8.
157. Bliss TVP, Douglas RM, Errington ML, Lynch MA. Correlation between long-term potentiation and release of endogenous amino acids from dentate gyrus of anesthetized rats. *J Physiol* 1986; 377: 391-408.
158. Errington ML, Lynch MA, Bliss TVP. Long-term potentiation in the dentate gyrus: induction and increased glutamate release are blocked by D(-) aminophosphonovalerate. *Neuroscience* 1987; 20: 279-84.
159. Richter-Levin G, Canevari L, Bliss TVP. Long-term potentiation and glutamate release in the dentate gyrus: links to spatial learning. *Behav Brain Res* 1995; 66: 37-40.
160. Helme-Guizon A, Davis S, Israel M, Lesbats B, Mallet J, Laroche S, et al. Increase in syntaxin 1B and glutamate release in mossy fibre terminals following induction of LTP in the dentate gyrus: a candidate molecular mechanism underlying trans-synaptic plasticity. *Eur J Neurosci* 1998; 10: 2231-7.
161. Stevens CF, Wang Y. Changes in reliability of synaptic function as a mechanism for plasticity. *Nature* 1994; 371: 704-7.
162. Brundage JM, Dunwiddie TV. Modulation of excitatory synaptic transmission by adenosine released from single hippocampal pyramidal neurons. *J Neurosci* 1996; 16: 5603-12.
163. Dragunow M, Beilharz E, Mason B, Lawlor P, Abraham W, Gluckman P. Brain-derived neurotrophic factor expression after long-term potentiation. *Neurosci Lett* 1993; 160: 232-6.
164. Hawkins RD, Zhuo M, Arancio O. Nitric oxide and carbon monoxide as possible retrograde messengers in hippocampal long-term potentiation. *J Neurobiol* 1994; 25: 652-65.
165. Huang EP. Synaptic plasticity: a role for nitric oxide in LTP. *Curr Biol* 1997; 7: R141-3.
166. Dinerman JL, Dawson TM, Schell MJ, Snowman A, Snyder SH. Endothelial nitric oxide synthase localized to hippocampal pyramidal cells: implications for synaptic plasticity. *Proc Natl Acad Sci U S A* 1994; 91: 4214-8.
167. Kato K, Zorumski CF. Platelet-activating factor as a potential retrograde messenger. *J Lipid Mediat Cell Signal* 1996; 14: 341-8.
168. Luo Y, Vallano ML. Arachidonic acid, but not sodium nitroprusside, stimulates presynaptic protein kinase C and phosphorylation of GAP-43 in rat hippocampal slices and synaptosomes. *J Neurochem* 1995; 64: 1808-18.
169. Gereau RW, Conn PJ. A cyclic AMP-dependent form of associative synaptic plasticity induced by coactivation of β -adrenergic receptors and metabotropic glutamate receptors in rat hippocampus. *J Neurosci* 1994; 14: 3310-8.
170. Haas HL, Sergueeva OA, Vorobjev VS, Sharonova IN. Subcortical modulation of synaptic plasticity in the hippocampus. *Behav Brain Res* 1995; 66: 41-4.
171. Buzsáki G, Gage FH. Absence of long-term potentiation in the subcortically deafferented dentate gyrus. *Brain Res* 1989; 484: 94-101.
172. Bergado JA, Moreno H, Núñez N. Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biol Res* 1996; 29: 197-202.
173. Bergado JA, Moreno H, Soto J, Castellano O, Castillo L. Septal fetal tissue transplants restore long-term potentiation in the dentate gyrus of fimbria-fornix-lesioned rats. *J Neural Transplant Plast* 1997; 6: 31-40.
174. Huerta PT, Lisman JE. Heightened synaptic plasticity of hippocampal CA1 neurons during a cholinergically induced rhythmic state. *Nature* 1993; 364: 723-5.
175. Huerta PT, Lisman JE. Bidirectional synaptic plasticity induced by a single burst during cholinergic theta oscillation in CA1 in vitro. *Neuron* 1995; 15: 1053-63.
176. Auerbach JM, Segal M. A novel cholinergic induction of long-term potentiation in rat hippocampus. *J Neurophysiol* 1994; 72: 2034-40.
177. Segal M, Auerbach JM. Muscarinic receptors involved in hippocampal plasticity. *Life Sci* 1997; 60: 1085-91.
178. Jerusalinsky D, Kornisiuk E, Izquierdo I. Cholinergic neurotransmission and synaptic plasticity concerning memory processing. *Neurochem Res* 1997; 22: 507-15.
179. Frey U, Schroeder H, Matthies H. Dopaminergic antagonists prevent long-term maintenance of posttetanic LTP in the CA1 region of rat hippocampal slices. *Brain Res* 1990; 522: 69-75.
180. Jibiki I, Wakita S, Kubota T, Kurokawa K, Fukushima T, Yamaguchi N. Haloperidol-induced blockade of induction of long-term potentiation in perforant path-dentate gyrus pathway in chronically prepared rabbits. *Pharmacol Biochem Behav* 1993; 46: 847-52.
181. Huang YY, Kandel ER. D1/D5 receptor agonists induce a protein synthesis-dependent late potentiation in the CA1 region of the hippocampus. *Proc Natl Acad Sci U S A* 1995; 92: 2446-50.
182. Stanton PK, Sarvey JM. Norepinephrine regulates long-term potentiation of both the population spike and dendritic EPSP in hippocampal dentate gyrus. *Brain Res Bull* 1987; 18: 115-9.
183. Robinson GB, Racine RJ. Long-term potentiation in the dentate gyrus: effects of noradrenaline depletion in the awake rat. *Brain Res* 1985; 325: 71-8.
184. Stanton PK, Sarvey JM. Blockade of norepinephrine-induced long-lasting potentiation in the hippocampal dentate gyrus by an inhibitor of protein synthesis. *Brain Res* 1985; 361: 276-83.
185. Katsuki H, Izumi Y, Zorumski CF. Noradrenergic regulation of synaptic plasticity in the hippocampal CA1 region. *J Neurophysiol* 1997; 77: 3013-20.
186. Klancnik JM, Phillips AG. Modulation of synaptic plasticity in the dentate gyrus of the rat by electrical stimulation of the median raphe nucleus. *Brain Res* 1991; 557: 236-40.

187. Sakai N, Tanaka C. Inhibitory modulation of long-term potentiation via the 5-HT_{1A} receptor in slices of the rat hippocampal dentate gyrus. *Brain Res* 1993; 613: 326-30.
188. Brown RE, Fedorov NB, Haas HL, Reymann KG. Histaminergic modulation of synaptic plasticity in area CA1 of rat hippocampal slices. *Neuropharmacology* 1995; 34: 181-90.
189. Aggleton JP. The contribution of the amygdala to normal and abnormal emotional states. *Trends Neurosci* 1993; 16: 328-33.
190. LeDoux JE. The amygdala: contribution to fear and stress. *Semin Neurosci* 1994; 6: 237.
191. LeDoux JE. Emotional memory systems in the brain. *Behav Brain Res* 1993; 58: 69-79.
192. McGaugh JL, Cahill L, Roozendaal B. Involvement of the amygdala in memory storage: interaction with other brain systems. *Proc Natl Acad Sci U S A* 1996; 93: 13508-14.
193. Ikegaya Y, Saito H, Abe K. The basomedial and basolateral amygdaloid nuclei contribute to the induction of long-term potentiation in the dentate gyrus in vivo. *Eur J Neurosci* 1996; 8: 1833-9.
194. Ikegaya Y, Saito H, Abe K. High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 1995; 22: 203-7.
195. Teyler TJ, Fountain SB. Neuronal plasticity in the mammalian brain: relevance to behavioral learning and memory. *Child Dev* 1987; 58: 698-712.
196. Stevens CF. A million dollar question: does LTP= memory. *Neuron* 1998; 20: 1-2.
197. Bliss TVP. The saturation debate. *Science* 1998; 281: 1975-6.
198. Moser EI, Moser MB. Is learning blocked by saturation of synaptic weights in the hippocampus? *Neurosci Biobehav Rev* 1999; 23: 661-72.
199. Barnes CA, Jung MW, McNaughton BL, Korol DL, Andreasson K, Worley PF. LTP saturation and spatial learning disruption: effects of task variables and saturation levels. *J Neurosci* 1994; 14: 5793-806.
200. Matthies H, Ruethrich H, Ott T, Matthies HK, Matthies R. Low frequency perforant path stimulation as a conditioned stimulus demonstrates correlations between long-term synaptic potentiation and learning. *Physiol Behav* 1986; 36: 811-21.
201. Wilson MA, Tonegawa S. Synaptic plasticity, place cells and spatial memory: study with second generation knockouts. *Trends Neurosci* 1997; 20: 102-6.
202. Morris RG. Synaptic plasticity and learning: selective impairment of learning rats and blockade of long-term potentiation in vivo by the N-methyl-D-aspartate receptor antagonist AP5. *J Neurosci* 1989; 9: 3040-57.
203. Tsien JZ, Huerta PT, Tonegawa S. The essential role of hippocampal CA1 NMDA receptor-dependent synaptic plasticity in spatial memory. *Cell* 1996; 87: 1327-38.
204. Silva AJ, Rosahl TW, Chapman PF, Marowitz Z, Friedman E, Frankland PW, et al. Impaired learning in mice with abnormal short-lived plasticity. *Curr Biol* 1996; 6: 1509-18.
205. Van Reempts J, Dikova M, Werbrouck L, Clincke G, Borgers M. Synaptic plasticity in rat hippocampus associated with learning. *Behav Brain Res* 1992; 51: 179-83.
206. Voronin LL. [Synaptic plasticity at the levels of the archicortex and neocortex]. *Neirofiziologija* 1984; 16: 651-65.
207. Toyama K, Komatsu Y, Yamamoto N, Kurotani T, Yamada K. In vitro approach to visual cortical development and plasticity. *Neurosci Res* 1991; 12: 57-71.
208. Hubel DH, Wiesel TN. Early exploration of the visual cortex. *Neuron* 1998; 20: 401-12.
209. Castro-Alamancos MA, Donoghue JP, Connors BW. Different forms of synaptic plasticity in somatosensory and motor areas of the neocortex. *J Neurosci* 1995; 15: 5324-33.
210. Jones TA, Kleim JA, Greenough WT. Synaptogenesis and dendritic growth in the cortex opposite unilateral sensorimotor cortex damage in adult rats: a quantitative electron microscopic examination. *Brain Res* 1996; 733: 142-8.
211. Cruikshank SJ, Weinberger NM. Receptive-field plasticity in the adult auditory cortex induced by Hebbian covariance. *J Neurosci* 1996; 16: 861-75.
212. Cruikshank SJ, Weinberger NM. Evidence for the Hebbian hypothesis in experience-dependent physiological plasticity of neocortex: a critical review. *Brain Res Rev* 1996; 22: 191-228.
213. Hedberg TG, Stanton PK. Long-term plasticity in cingulate cortex requires both NMDA and metabotropic glutamate receptor activation. *Eur J Pharmacol* 1996; 310: 19-27.
214. Kaczmarek L, Kossut M, Skangiel-Kramska J. Glutamate receptors in cortical plasticity: molecular and cellular biology. *Physiol Rev* 1997; 77: 217-55.
215. Komatsu Y. Plasticity of excitatory synaptic transmission in kitten visual cortex depends on voltage-dependent Ca²⁺ channels but not on NMDA receptors. *Neurosci Res* 1994; 20: 209-12.
216. Gordon JA, Cioffi D, Silva AJ, Stryker MP. Deficient plasticity in the primary visual cortex of a -calcium/calmodulin- dependent protein kinase II mutant mice. *Neuron* 1996; 17: 491-9.
217. Deisseroth K, Bito H, Tsien RW. Signaling from synapse to nucleus: postsynaptic CREB phosphorylation during multiple forms of hippocampal synaptic plasticity. *Neuron* 1996; 16: 89-101.
218. Kaminska B, Mosieniak G, Gierdalski M, Kossut M, Kaczmarek L. Elevated AP-1 transcription factor DNA binding activity at the onset of functional plasticity during development of rat sensory cortical areas. *Mol Brain Res* 1995; 33: 295-304.
219. Maho C, Hars B, Edeline JM, Hennevin E. Conditioned changes in the basal forebrain: relations with learning-induced cortical plasticity. *Psychobiology* 1995; 23: 10-25.
220. Rauschecker JP. Developmental plasticity and memory. *Behav Brain Res* 1995; 66: 7-12.
221. Rajan R. Receptor organ damage causes loss of cortical surround inhibition without topographic map plasticity. *Nat Neurosci* 1998; 1: 138-43.
222. Kilgard MP, Merzenich MM. Cortical map reorganization enabled by nucleus basalis activity. *Science* 1998; 279: 1714-8.
223. Singer W. Development and plasticity of cortical processing architectures. *Science* 1995; 270: 758-64.
224. Sermasi E, Tropea D, Domenici L. A new form of synaptic plasticity is transiently expressed in the developing rat visual cortex: a modulatory role for visual experience and brain-derived neurotrophic factor. *Neuroscience* 1999; 91: 163-73.
225. Schlaggar BL, Fox K, O'Leary DDM. Postsynaptic control of plasticity in developing somatosensory cortex. *Nature* 1993; 364: 623-6.
226. Maren S. Synaptic transmission and plasticity in the amygdala: an emerging physiology of fear conditioning circuits. *Mol Neurobiol* 1996; 13: 1-22.
227. Clugnet MC, LeDoux JE. Synaptic plasticity in fear conditioning circuits: induction of LTP in the lateral nucleus of the amygdala by stimulation of the medial geniculate body. *J Neurosci* 1990; 10: 2818-24.
228. Rogan MT, LeDoux JE. LTP is accompanied by commensurate enhancement of auditory-evoked responses in a fear conditioning circuit. *Neuron* 1995; 15: 127-36.
229. Rogan MT, Staubli UV, LeDoux JE. Fear conditioning induces associative long-term potentiation in the amygdala. *Nature* 1997; 390: 604-7.
230. Maren S, Fanselow MS. Synaptic plasticity in the basolateral amygdala induced by hippocampal formation stimulation in vivo. *J Neurosci* 1995; 15: 7548-64.
231. Meftah EM, Rispal-Padel L. Synaptic plasticity in the thalamo-cortical pathway as one of the neurobiological correlates of forelimb flexion conditioning: electrophysiological investigation in the cat. *J Neurophysiol* 1994; 72: 2631-47.
232. Pekhletski R, Gerlai R, Overstreet LS, Huang XP, Agopyan N, Slater NT, et al. Impaired cerebellar synaptic plasticity and motor performance in mice lacking the mGluR4 subtype of metabotropic glutamate receptor. *J Neurosci* 1996; 16: 6364-73.
233. Kleim JA, Swain RA, Czerlanis NC, Kelly JL, Pipitone MA, Greenough WT. Learning-dependent dendritic hypertrophy of cerebellar stellate cells: plasticity of local circuit neurons. *Neurobiol Learn Mem* 1997; 67: 29-33.
234. Charpier S, Deniau JM. In vivo activity-dependent plasticity at cortico-striatal connections: evidence for physiological long-term potentiation. *Proc Natl Acad Sci U S A* 1997; 94: 7036-40.
235. Pennartz CMA, Ameerun RF, Groenewegen HJ, Lopes Da Silva FH. Synaptic plasticity in an in vitro slice preparation of the rat nucleus accumbens. *Eur J Neurosci* 1993; 5: 107-17.
236. Mulder AB, Arts MP, Da Silva FH. Short- and long-term plasticity of the hippocampus to nucleus accumbens and prefrontal cortex pathways in the rat, in vivo. *Eur J Neurosci* 1997; 9: 1603-11.
237. Pockett S. Long-term potentiation and depression in the intermediate gray matter of rat spinal cord in vitro. *Neuroscience* 1995; 67: 791-8.
238. Pockett S, Figurov A. Long-term potentiation and depression in the ventral horn of rat spinal cord in vitro. *Neuroreport* 1993; 4: 97-9.
239. Charpier S, Behrends JC, Triller A, Faber DS, Korn H. 'Latent' inhibitory connections become functional during activity-dependent plasticity. *Proc Natl Acad Sci U S A* 1995; 92: 117-20.
240. Liao D, Hessler NA, Malinow R. Activation of postsynaptically silent synapses during pairing-induced LTP in CA1 region of hippocampal slice. *Nature* 1995; 375: 400-4.
241. Isaac JTR, Nicoll RA, Malenka RC. Evidence for silent synapses: implications for the expression of LTP. *Neuron* 1995; 15: 427-34.
242. Nicoll RA, Malenka RC. Expression mechanisms underlying NMDA receptor-dependent long-term potentiation. *Ann N Y Acad Sci* 1999; 868: 515-25.
243. Durand GM, Konnerth A. Long-term potentiation as a mechanism of

- functional synapse induction in the developing hippocampus. *J Physiol (Paris)* 1996; 90: 313-5.
244. Takumi Y, Ramírez-León V, Laake P, Ottersen OP. Different modes of expression of AMPA and NMDA receptors in hippocampal synapses. *Nat Neurosci* 1999; 2: 619-24.
 245. Rumpel S, Hatt H, Gottmann K. Silent synapses in the developing rat visual cortex: evidence for postsynaptic expression of synaptic plasticity. *J Neurosci* 1998; 18: 8863-74.
 246. Schachner M. Neural recognition molecules and synaptic plasticity. *Curr Opin Cell Biol* 1997; 9: 627-34.
 247. Doherty P, Fazeli MS, Walsh FS. The neural cell adhesion molecule and synaptic plasticity. *J Neurobiol* 1995; 26: 437-46.
 248. Glanzman DL. Postsynaptic regulation of the development and long-term plasticity of aplysia sensorimotor synapses in cell culture. *J Neurobiol* 1994; 25: 666-93.
 249. Sekino Y, Kuroda Y. Synaptic plasticity and gene products. *Nippon Rinsho* 1992; 50: 2796-807.
 250. Schacher S, Wu F, Sun ZY. Pathway-specific synaptic plasticity: activity-dependent enhancement and suppression of long-term heterosynaptic facilitation at converging inputs on a single target. *J Neurosci* 1997; 17: 597-606.
 251. Wang J, Renger JJ, Griffith LC, Greenspan RJ, Wu CF. Concomitant alterations of physiological and developmental plasticity in *Drosophila* CaM kinase II-inhibited synapses. *Neuron* 1994; 13: 1373-84.
 252. Zhong Y, Wu CF. Altered synaptic plasticity in *drosophila* memory mutants with a defective cyclic AMP cascade. *Science* 1990; 251: 198-201.
 253. Kirkwood A, Dudek SM, Gold JT, Aizenman CD, Bear MF. Common forms of synaptic plasticity in the hippocampus and neocortex in vitro. *Science* 1993; 260: 1518-21.
 254. Shen JM, Barnes CA, McNaughton BL, Skaggs WE, Weaver KL. The effect of aging on experience-dependent plasticity of hippocampal place cells. *J Neurosci* 1997; 17: 6769-82.
 255. Bergado JA, Fernández CI, Gómez-Soria A, González O. Chronic intraventricular infusion with NGF improves LTP in old cognitively-impaired rats. *Brain Res* 1997; 770: 1-9.
 256. Barnes CA, McNaughton BL. An age comparison of the rates of acquisition and forgetting. *Behav Neurosci* 1985; 99: 1040-8.
 257. De Toledo-Morrell L, Geinisman Y, Morrell F. Age-dependent alterations in hippocampal synaptic plasticity: relation to memory disorders. *Neurobiol Aging* 1988; 9: 581-90.
 258. Kamal A, Biessels GJ, Gispen WH, Urban IJA. Increasing age reduces expression of long-term depression and dynamic range of transmission plasticity in CA1 field of the rat hippocampus. *Neuroscience* 1998; 83: 707-15.
 259. Kirkwood A, Silva A, Bear MF. Age-dependent decrease of synaptic plasticity in the neocortex of α CaMKII mutant mice. *Proc Natl Acad Sci U S A* 1997; 94: 3380-3.
 260. Schauwecker PE, Cheng HW, Serquinia RMP, Mori N, McNeill TH. Lesion-induced sprouting of commissural/associational axons and induction of GAP-43 mRNA in hilar and CA3 pyramidal neurons in the hippocampus are diminished in aged rats. *J Neurosci* 1995; 15: 2462-70.
 261. Scott SA, Liang S, Weingartner JA, Crutcher KA. Increased NGF-like activity in young but not aged rat hippocampus after septal lesions. *Neurobiol Aging* 1994; 15: 337-46.
 262. Nagahara AH, Nicolle MM, Gallagher M. Alterations in [3H]-kainate receptor binding in the hippocampal formation of aged Long-Evans rats. *Hippocampus* 1993; 3: 269-77.
 263. Neill D. Alzheimer's disease: maladaptive synaptoplasticity hypothesis. *Neurodegeneration* 1995; 4: 217-32.
 264. Krugers HJ, Mulder M, Korf J, Havekes L, De Kloet ER, Joëls M. Altered synaptic plasticity in hippocampal CA1 area of apolipoprotein E deficient mice. *Neuroreport* 1997; 8: 2505-10.
 265. Seabrook GR, Smith DW, Bowery BJ, Easter A, Reynolds T, Fitzjohn SM, et al. Mechanisms contributing to the deficits in hippocampal synaptic plasticity in mice lacking amyloid precursor protein. *Neuropharmacology* 1999; 38: 349-59.
 266. Sugaya K, Chouinard M, Greene R, Robbins M, Personett D, Kent C, et al. Molecular indices of neuronal and glial plasticity in the hippocampal formation in a rodent model of age-induced spatial learning impairment. *J Neurosci* 1996; 16: 3427-43.
 267. Masliah E, Honer WG, Mallory M, Voigt M, Kushner P, Hansen L, et al. Topographical distribution of synaptic-associated proteins in the neuritic plaques of Alzheimer's disease hippocampus. *Acta Neuropathol (Berl)* 1994; 87: 135-42.
 268. Calabresi P, Pisani A, Mercuri NB, Bernardi G. The corticostriatal projection: from synaptic plasticity to dysfunctions of the basal ganglia. *Trends Neurosci* 1996; 19: 19-24.
 269. Calabresi P, Pisani A, Centonze D, Bernardi G. Synaptic plasticity and physiological interactions between dopamine and glutamate in the striatum. *Neurosci Biobehav Rev* 1997; 21: 519-23.
 270. Usdin MT, Shelbourne PF, Myers RM, Madison DV. Impaired synaptic plasticity in mice carrying the Huntington's disease mutation. *Hum Mol Genet* 1999; 8: 839-46.
 271. Represa A, Niquet J, Pollard H, Ben-Ari Y. Cell death, gliosis, and synaptic remodeling in the hippocampus of epileptic rats. *J Neurobiol* 1995; 26: 413-25.
 272. Represa A, Jorquera I, Le Gal La Salle G, Ben-Ari Y. Epilepsy induced collateral sprouting of hippocampal mossy fibers: does it induce the development of ectopic synapses with granule cell dendrites? *Hippocampus* 1993; 3: 257-68.
 273. Meberg PJ, Gall CM, Routtenberg A. Induction of F1/GAP-43 gene: expression in hippocampal granule cells after seizures. *Mol Brain Res* 1993; 17: 295-9.
 274. Represa A, Pollard H, Moreau J, Ghilini G, Khrestchatsky M, Ben-Ari Y. Mossy fiber sprouting in epileptic rats is associated with a transient increased expression of α -tubulin. *Neurosci Lett* 1993; 156: 149-52.
 275. Pollard H, Khrestchatsky M, Moreau J, Ben-Ari Y, Represa A. Correlation between reactive sprouting and microtubule protein expression in epileptic hippocampus. *Neuroscience* 1994; 61: 773-87.
 276. Niquet J, Jorquera I, Ben-Ari Y, Represa A. NCAM immunoreactivity on mossy fibers and reactive astrocytes in the hippocampus of epileptic rats. *Brain Res* 1993; 626: 106-16.
 277. Wang S, Lees GJ, Bock E, Hamberger A, Haglid KG. Biphasic changes in NCAM level after an NMDA lesion to the hippocampal formation: a quantitative dot-immunobinding assay. *J Neurosci Res* 1992; 33: 626-30.
 278. Okazaki MM, Evenson DA, Nadler JV. Hippocampal mossy fiber sprouting and synapse formation after status epilepticus in rats: visualization after retrograde transport of biocytin. *J Comp Neurol* 1995; 352: 515-34.
 279. Golarai G, Sutula TP. Functional alterations in the dentate gyrus after induction of long-term potentiation, kindling, and mossy fiber sprouting. *J Neurophysiol* 1996; 75: 343-53.
 280. Golarai G, Cavazos JE, Sutula TP. Activation of the dentate gyrus by pentylentetrazol evoked seizures induces mossy fiber synaptic reorganization. *Brain Res* 1992; 593: 257-64.
 281. Dudek FE, Obenaus A, Schweitzer JS, Wuarin JP. Functional significance of hippocampal plasticity in epileptic brain: electrophysiological changes of the dentate granule cells associated with mossy fiber sprouting. *Hippocampus* 1994; 4: 259-65.
 282. Pérez Y, Morin F, Beaulieu C, Lacaille JC. Axonal sprouting of CA1 pyramidal cells in hyperexcitable hippocampal slices of kainate-treated rats. *Eur J Neurosci* 1996; 8: 736-48.
 283. Bundman MC, Gall CM. Ultrastructural plasticity of the dentate gyrus granule cells following recurrent limbic seizures. II. Alterations in somatic synapses. *Hippocampus* 1994; 4: 611-22.
 284. Bundman MC, Pico RM, Gall CM. Ultrastructural plasticity of the dentate gyrus granule cells following recurrent limbic seizures. I. Increase in somatic spines. *Hippocampus* 1994; 4: 601-10.
 285. Qiao X, Noebels JL. Developmental analysis of hippocampal mossy fiber outgrowth in a mutant mouse with inherited spike-wave seizures. *J Neurosci* 1993; 13: 4622-35.
 286. Mathern GW, Leite JP, Pretorius JK, Quinn B, Peacock WJ, Babb TL. Children with severe epilepsy: evidence of hippocampal neuron losses and aberrant mossy fiber sprouting during postnatal granule cell migration and differentiation. *Dev Brain Res* 1994; 78: 70-80.
 287. Mathern GW, Pretorius JK, Babb TL. Quantified patterns of mossy fiber sprouting and neuron densities in hippocampal and lesional seizures. *J Neurosurg* 1995; 82: 211-9.
 288. Mathern GW, Babb TL, Micevych PE, Blanco CE, Pretorius JK. Granule cell mRNA levels for BDNF, NGF, and NT-3 correlate with neuron losses or supragranular mossy fiber sprouting in the chronically damaged and epileptic human hippocampus. *Mol Chem Neuropathol* 1997; 30: 53-76.
 289. Cavazos JE, Das I, Sutula TP. Neuronal loss induced in limbic pathways by kindling: evidence for induction of hippocampal sclerosis by repeated brief seizures. *J Neurosci* 1994; 14: 3106-21.
 290. McKay SE, Purcell AL, Carew TJ. Regulation of synaptic function by neurotrophic factors in vertebrates and invertebrates: implications for development and learning. *Learn Mem* 1999; 6: 193-215.
 291. Davies AM. The neurotrophic hypothesis: where does it stand? *Philos Trans R Soc Lond B Biol Sci* 1996; 351: 389-94.
 292. Barde YA. Neurotrophins: a family of proteins supporting the survival of neurons. *Prog Clin Biol Res* 1994; 390: 45-56.
 293. Henderson CE. Role of neurotrophic factors in neuronal development. *Curr Opin Neurobiol* 1996; 6: 64-70.
 294. Barbacid M. The Trk family of neurotrophin receptors. *J Neurobiol* 1994; 25: 1386-403.
 295. Segal RA, Greenberg ME. Intracellular signaling pathways activated by neurotrophic factors. *Annu Rev Neurosci* 1996; 19: 463-89.
 296. Vogt Weisenhorn DM, Roback J, Young AN, Wainer BH. Cellular

- aspects of trophic actions in the nervous system. *Int Rev Cytol* 1999; 189: 177-265.
297. Humpel C, Wetmore C, Olson L. Regulation of brain-derived neurotrophic factor messenger RNA and protein at the cellular level in pentylenetetrazol-induced epileptic seizures. *Neuroscience* 1993; 53: 909-18.
 298. Patterson SL, Grover LM, Schwartzkroin PA, Bothwell M. Neurotrophin expression in rat hippocampal slices: a stimulus paradigm inducing LTP in CA1 evokes increases in BDNF and NT-3 mRNAs. *Neuron* 1992; 9: 1081-8.
 299. Castren E, Pitkanen M, Sirvio J, Parsadanian A, Lindholm D, Thoenen H, et al. The induction of LTP increases BDNF and NGF mRNA but decreases NT-3 mRNA in the dentate gyrus. *Neuroreport* 1993; 4: 895-8.
 300. Dragunow M, Hughes P, Mason-Parker SE, Lawlor P, Abraham WC. TrkB expression in dentate granule cells is associated with a late phase of long-term potentiation. *Brain Res Mol Brain Res* 1997; 46: 274-80.
 301. Kang HJ, Schuman EM. Neurotrophin-induced modulation of synaptic transmission in the adult hippocampus. *J Physiol (Paris)* 1995; 89: 11-22.
 302. Korte M, Carroll P, Wolf E, Brem G, Thoenen H, Bonhoeffer T. Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc Natl Acad Sci U S A* 1995; 92: 8856-60.
 303. Korte M, Staiger V, Griesbeck O, Thoenen H, Bonhoeffer T. The involvement of brain-derived neurotrophic factor in hippocampal long-term potentiation revealed by gene targeting experiments. *J Physiol (Paris)* 1996; 90: 157-64.
 304. Korte M, Griesbeck O, Gravel C, Carroll P, Staiger V, Thoenen H, et al. Virus-mediated gene transfer into hippocampal CA1 region restores long-term potentiation in brain-derived neurotrophic factor mutant mice. *Proc Natl Acad Sci U S A* 1996; 93: 12547-52.
 305. Lu B, Chow A. Neurotrophins and hippocampal synaptic transmission and plasticity. *J Neurosci Res* 1999; 58: 76-87.
 306. Chen G, Kolbeck R, Barde YA, Bonhoeffer T, Kossel A. Relative contribution of endogenous neurotrophins in hippocampal long-term potentiation. *J Neurosci* 1999; 19: 7983-90.
 307. Suen PC, Wu K, Levine ES, Mount HT, Xu JL, Lin SY, et al. Brain-derived neurotrophic factor rapidly enhances phosphorylation of the postsynaptic N-methyl-D-aspartate receptor subunit 1. *Proc Natl Acad Sci U S A* 1997; 94: 8191-5.
 308. Blanquet PR, Lamour Y. Brain-derived neurotrophic factor increases Ca²⁺/calmodulin-dependent protein kinase 2 activity in hippocampus. *J Biol Chem* 1997; 272: 24133-6.
 309. Jovanovic JN, Benfenati F, Siow YL, Sihra TS, Sanghera JS, Pelech SL, et al. Neurotrophins stimulate phosphorylation of synapsin I by MAP kinase and regulate synapsin I-actin interactions. *Proc Natl Acad Sci U S A* 1996; 93: 3679-83.
 310. Kang HJ, Schuman EM. A requirement for local protein synthesis in neurotrophin-induced hippocampal synaptic plasticity. *Science* 1996; 273: 1402-6.
 311. Bergado JA, Gómez-Soria A, Cruz R, Fernández CI. Nerve growth factor improves evoked potentials and long-term potentiation in the dentate gyrus of presenile rats. *Eur J Pharmacol* 1998; 345: 181-4.
 312. Berardi N, Maffei L. From visual experience to visual function: roles of neurotrophins. *J Neurobiol* 1999; 41: 119-26.
 313. Akaneya Y, Tsumoto T, Hatanaka H. Brain-derived neurotrophic factor blocks long-term depression in rat visual cortex. *J Neurophysiol* 1996; 76: 4198-201.
 314. Galuske RA, Kim DS, Castren E, Thoenen H, Singer W. Brain-derived neurotrophic factor reversed experience-dependent synaptic modifications in kitten visual cortex. *Eur J Neurosci* 1996; 8: 1554-9.
 315. Prakash N, Cohen-Cory S, Frostig RD. RAPID and opposite effects of BDNF and NGF on the functional organization of the adult cortex in vivo. *Nature* 1996; 381: 702-6.
 316. Huang ZJ, Kirkwood A, Pizzorusso T, Porciatti V, Morales B, Bear MF, et al. BDNF regulates the maturation of inhibition and the critical period of plasticity in mouse visual cortex. *Cell* 1999; 98: 739-55.
 317. Carmignoto G, Pizzorusso T, Tia S, Vicini S. Brain-derived neurotrophic factor and nerve growth factor potentiate excitatory synaptic transmission in the rat visual cortex. *J Physiol (Lond)* 1997; 498: 153-64.
 318. Akaneya Y, Tsumoto T, Kinoshita S, Hatanaka H. Brain-derived neurotrophic factor enhances long-term potentiation in rat visual cortex. *J Neurosci* 1997; 17: 6707-16.
 319. Black IB. Trophic regulation of synaptic plasticity. *J Neurobiol* 1999; 41: 108-18.
 320. Cellerino A, Maffei L. The action of neurotrophins in the development and plasticity of the visual cortex. *Prog Neurobiol* 1996; 49: 53-71.
 321. Jackson GR, Werbach-Pérez K, Pan Z, Sampath D, Pérez-Polo JR. Neurotrophin regulation of energy homeostasis in the central nervous system. *Dev Neurosci* 1994; 16: 285-90.
 322. Kirschner PB, Jenkins BG, Schulz JB, Finkelstein SP, Matthews RT, Rosen BR, et al. NGF, BDNF and NT-5, but not NT-3 protect against MPP⁺ toxicity and oxidative stress in neonatal animals. *Brain Res* 1996; 713: 178-85.
 323. Mocchetti I, Wrathall JR. Neurotrophic factors in central nervous system trauma. *J Neurotrauma* 1995; 12: 853-70.
 324. Cruz-Aguado R, Fernández CI, Díaz CM, González O, Antúnez I, Bergado J. Effects of nerve growth factor on brain glutathione-related enzymes from aged rats. *Fundam Clin Pharmacol* 1998; 12: 546-52.
 325. Cruz-Aguado R, Francis-Turner L, Díaz-Suárez CM, Bergado J. NGF prevents changes in rat brain glutathione-related enzymes following transection of the septohippocampal pathway. *Neurochem Int* 1999; 34: 125-30.
 326. Ferrer I, Marín C, Rey MJ, Ribalta T, Goutan E, Blanco R, et al. BDNF and full-length and truncated TrkB expression in Alzheimer's disease. Implications in therapeutic strategies. *J Neuropathol Exp Neurol* 1999; 58: 729-39.
 327. Knusel B, Gao H. Neurotrophins and Alzheimer's disease: beyond the cholinergic neurons. *Life Sci* 1996; 58: 2019-27.
 328. Sofroniew MV, Cooper JD, Svendsen CN, Crossman P, Ip NY, Lindsay RM, et al. Atrophy but not death of adult septal cholinergic neurons after ablation of target capacity to produce mRNAs for NGF, BDNF, and NT3. *J Neurosci* 1993; 13: 5263-76.
 329. Collier TJ, Sortwell CE. Therapeutic potential of nerve growth factors in Parkinson's disease. *Drugs Aging* 1999; 14: 261-87.
 330. Hyman C, Juhasz M, Jackson C, Wright P, Ip NY, Lindsay RM. Overlapping and distinct actions of the neurotrophins BDNF, NT-3, and NT-4/5 on cultured dopaminergic and GABAergic neurons of the ventral mesencephalon. *J Neurosci* 1994; 14: 335-47.
 331. Walton KM. GDNF: a novel factor with therapeutic potential for neurodegenerative disorders. *Mol Neurobiol* 1999; 19: 43-59.
 332. Lindsay RM. Trophic protection of motor neurons: clinical potential in motor neuron diseases. *J Neurol* 1994; 242: 8-11.
 333. Junger H, Varon S. Neurotrophin-4 (NT-4) and glial cell line-derived neurotrophic factor (GDNF) promote the survival of corticospinal motor neurons of neonatal rats in vitro. *Brain Res* 1997; 762: 56-60.
 334. Kordower JH, Isacson O, Emerich DF. Cellular delivery of trophic factors for the treatment of Huntington's disease: is neuroprotection possible? *Exp Neurol* 1999; 159: 4-20.
 335. Davies AM. Neurotrophin switching: where does it stand? *Curr Opin Neurobiol* 1997; 7: 110-8.
 336. Croll SD, Suri C, Compton DL, Simmons MV, Yancopoulos GD, Lindsay RM, et al. Brain-derived neurotrophic factor transgenic mice exhibit passive avoidance deficits, increased seizure severity and in vitro hyperexcitability in the hippocampus and entorhinal cortex. *Neuroscience* 1999; 93: 1491-506.
 337. Conti AM, Fischer SJ, Windebank AJ. Inhibition of axonal growth from sensory neurons by excess nerve growth factor. *Ann Neurol* 1997; 42: 838-46.
 338. Zapf-Colby A, Eichhorn J, Webster NJ, Olefsky JM. Inhibition of PLC-gamma1 activity converts nerve growth factor from an anti-mitogenic to a mitogenic signal in CHO cells. *Oncogene* 1999; 18: 4908-19.
 339. Liu Y, Kim D, Himes BT, Chow SY, Schallert T, Murray M, et al. Transplants of fibroblasts genetically modified to express BDNF promote regeneration of adult rat rubrospinal axons and recovery of forelimb function. *J Neurosci* 1999; 19: 4370-87.
 340. Marconi P, Simonato M, Zucchini S, Bregola G, Argnani R, Krisky D, et al. Replication-defective herpes simplex virus vectors for neurotrophic factor gene transfer in vitro and in vivo. *Gene Ther* 1999; 6: 904-12.
 341. Martínez-Serrano A, Björklund A. Protection of the neostriatum against excitotoxic damage by neurotrophin-producing, genetically modified neural stem cells. *J Neurosci* 1996; 16: 4604-16.
 342. Wyman T, Rohrer D, Kirigiti P, Nichols H, Pilcher K, Nilaver G, et al. Promoter-activated expression of nerve growth factor for treatment of neurodegenerative diseases. *Gene Ther* 1999; 6: 1648-60.
 343. Yoshimoto Y, Lin Q, Collier TJ, Frim DM, Breakefield XO, Bohn MC. Astrocytes retrovirally transduced with BDNF elicit behavioral improvement in a rat model of Parkinson's disease. *Brain Res* 1995; 691: 25-36.
 344. Poduslo JF, Curran GL. Permeability at the blood-brain and blood-nerve barriers of the neurotrophic factors: NGF, CNTF, NT-3, BDNF. *Brain Res Mol Brain Res* 1996; 36: 280-6.
 345. Salim KN, McEwen BS, Chao HM. Ginsenoside Rb1 regulates ChAT, NGF and trkA mRNA expression in the rat brain. *Brain Res Mol Brain Res* 1997; 47: 177-82.
 346. Fukumitsu H, Sometani A, Ohmiya M, Nitta A, Nomoto H, Furukawa Y, et al. Induction of a physiologically active brain-derived neurotrophic factor in the infant rat brain by peripheral administration of 4-methylcatechol. *Neurosci Lett* 1999; 274: 115-8.
 347. Nitta A, Ogihara Y, Onishi J, Hasegawa T, Furukawa S, Nabeshima T. Oral administration of propentofylline, a stimulator of nerve growth factor (NGF) synthesis, recovers cholinergic neuronal dysfunction in-

- duced by the infusion of anti-NGF antibody into the rat septum. *Behav Brain Res* 1997; 83: 201-4.
348. Yamada K, Nitta A, Hasegawa T, Fuji K, Hiramatsu M, Kameyama T, et al. Orally active NGF synthesis stimulators: potential therapeutic agents in Alzheimer's disease. *Behav Brain Res* 1997; 83: 117-22.
 349. Hughes PE, Alexi T, Walton M, Fuji K, Hiramatsu M, Kameyama T, et al. Activity and injury-dependent expression of inducible transcription factors, growth factors and apoptosis-related genes within the central nervous system. *Prog Neurobiol* 1999; 57: 421-50.
 350. Smith MA. Hippocampal vulnerability to stress and aging: possible role of neurotrophic factors. *Behav Brain Res* 1996; 78: 25-36.
 351. Anonymous. A controlled trial of recombinant methionyl human BDNF in ALS: the BDNF Study Group (Phase III). *Neurology* 1999; 52: 1427-3.
 352. Mandel P, Edel S, Poirel G. Free nucleotides in rabbit and guinea pig brain. *J Neurochem* 1966; 12: 885-6.
 353. Wenzel M, Wenzel J, Grosse G, Lindner G, Kirsche W, Matthies H. [The effect of orotic acid and orotic acid derivatives on the in vitro differentiation of hippocampus neurons-morphometric and electron microscopic studies of ribosomes]. *Z Mikrosk Anat Forsch* 1976; 90: 834-51.
 354. Wenzel M, Wenzel J, Grosse G, Lindner G, Matthies H. [Morphometric studies on the in vivo differentiation of hippocampus neurons under pharmacological conditions]. *Verh Anat Ges* 1977; (71 Pt 1): 121-6.
 355. Lindner G, Grosse G. [The effect of active substances on nerve fiber regeneration in nerve tissue cultures]. *Z Mikrosk Anat Forsch* 1980; 94: 1105-13.
 356. Grosse G, Lindner G, Matthies HJ. [Effect of orotic acid on the in vitro cultured nerve tissue]. *Z Mikrosk Anat Forsch* 1976; 90: 499-506.
 357. Grosse G, Lindner G, Matthies HJ. [The effect of orotic acid and sodium orotate on organ cultures of the hippocampus of embryonic rats]. *Z Mikrosk Anat Forsch* 1980; 94: 283-91.
 358. Götz D, Ziesch C, Wenzel M, Grosse G, Schuster T, Wenzel J. [Electron microscopic and morphometric studies on the in vitro differentiation of mitochondria in neurons of hippocampus explant cultures as affected by orotic acid and sodium orotate]. *Z Mikrosk Anat Forsch* 1982; 96: 613-32.
 359. Bergado Rosado JA, Rüethrich H, Matthies H. La estimulación eléctrica de la vía perforante como estímulo condicionado en shuttle box. Efecto del orotato de metilglucamina sobre la retención. *Rev Cubana Invest Biomed* 1987; 6: 347-59.
 360. Rüthrich HL, Wetzel W, Matthies H. Memory retention in old rats: improvement by orotic acid. *Psychopharmacology (Berlin)* 1983; 79: 348-51.
 361. Bergado JA, Krug M, Rüethrich H, Matthies H. Orotate improves memory and enhances synaptic long-term potentiation in active avoidance behaviour in rats with perforant path stimulation as the conditioned stimulus. *Eur J Pharmacol* 1988; 157: 155-63.
 362. Pohle W, Matthies H. Incorporation of RNA-precursors into neuronal and glial cells of rat brain during a learning experiment. *Brain Res* 1974; 65: 231-7.
 363. Krug M, Rüethrich H, Bergado J. The nootropic substance methylglucamine orotate prolongs both, postconditioning potentiation and post-tetanic LTP in the dentate gyrus of freely moving rats. *Activ Nerv Sup (Prague)* 1988; 30: 232-3.
 364. Krug M, Rüethrich H, Wagner M, Matthies H, Ott T. Einfluß von systemisch und intraventricular applizierter Orotsäure auf monosynaptisch evozierte Potentiale im Gyrus dentatus der freibeweglichen Ratte. *Biomed Biochim Acta* 1985; 44: 623-31.
 365. Krug M, Koch M, Schoof E, Wagner M, Matthies H. Methylglucamine orotate, a memory-improving drug, prolongs hippocampal long-term potentiation. *Eur J Pharmacol* 1989; 173: 223-6.
 366. Akiho H, Iwai A, Katoh-Soudh M, Tsukamoto S, Koshiya K, Yamaguchi T. Post-ischaemic treatment with orotic acid prevents neuronal injury in gerbil brain ischaemia. *Neuroreport* 1998; 8: 607-10.
 367. Sonnewald U, Akiho H, Koshiya K, Iwai A. Effect of orotic acid on the metabolism of cerebral cortical astrocytes during hypoxia and reoxygenation: an NMR spectroscopy study. *J Neurosci Res* 1998; 51: 103-8.
 368. Geiss KR, Stergiou N, Jester, Neuenfeld HU, Jester HG. Effects of magnesium orotate on exercise tolerance in patients with coronary heart disease. *Cardiovasc Drugs Ther* 1998; 12: 153-6.
 369. Zimmer HG. Effects of magnesium orotate on rat heart function. *Cardioscience* 1994; 5: 55-61.
 370. Jellinek H, Takacs E. Morphological aspects of the effects of orotic acid and magnesium orotate on hypercholesterolaemia in rabbits. *Arzneimittelforschung* 1995; 45: 836-42.
 371. Rahmann H. Brain gangliosides and memory formation. *Behav Brain Res* 1995; 66: 105-16.
 372. Skaper SD, Leon A, Facci L. Ganglioside GM1 prevents death induced by excessive excitatory neurotransmission in cultured hippocampal pyramidal neurons. *Neurosci Lett* 1991; 126: 98-101.
 373. Ledeen RW. Biology of gangliosides: neurotogenic and neuronotrophic properties. *J Neurosci Res* 1984; 12: 147-59.
 374. Masco D, Seifert W. Gangliosides in lesion-induced synaptogenesis: studies in the hippocampus of the rat brain. *Brain Res* 1990; 514: 84-92.
 375. Popov N, Toffano G, Riechert U, Matthies H. Effects of intraventricularly applied gangliosides and N-acetylneuraminic acid on acquisition and retention performance of a brightness discrimination task in rats. *Pharmacol Biochem Behav* 1989; 34: 209-12.
 376. Ramírez OA, Gómez RA, Carrer HF. Gangliosides improve synaptic transmission in dentate gyrus of hippocampal rat slices. *Brain Res* 1990; 506: 291-3.
 377. Okada Y, Rahmann H, Hirai H, Terashima A. Exogenously applied gangliosides (GM₁, GC_{1a} and G_{m1x}) fail to facilitate the induction of long-term potentiation (LTP) in the slices of hippocampus and superior colliculus of the guinea pig. *Neurosci Lett* 1994; 170: 269-72.
 378. Hadjiconstantinou M, Neff NH. GM1 ganglioside: in vivo and in vitro trophic actions on central neurotransmitter systems. *J Neurochem* 1998; 70: 1335-45.
 379. Cannella MS, Oderfeld-Nowak B, Gradkowska M, Skup M, Garofalo L, Cuello AC, et al. Derivatives of ganglioside GM1 as neuronotrophic agents: comparison of in vivo and in vitro effects. *Brain Res* 1990; 513: 286-94.
 380. Nagai Y. Functional roles of gangliosides in bio-signaling. *Behav Brain Res* 1995; 66: 99-104.
 381. McEwen BS. Gonadal and adrenal steroids regulate neurochemical and structural plasticity of the hippocampus via cellular mechanisms involving NMDA receptors. *Cell Mol Neurobiol* 1996; 16: 103-16.
 382. Bettini E, Pollio G, Santagati S, Maggi A. Estrogen receptor in rat brain: presence in the hippocampal formation. *Neuroendocrinology* 1992; 56: 502-8.
 383. Azcoitia I, Sierra A, García-Segura LM. Neuroprotective effects of estradiol in the adult rat hippocampus: interaction with insulin-like growth factor-I signalling. *J Neurosci Res* 1999; 58: 815-22.
 384. Woolley CS, McEwen BS. Estradiol regulates hippocampal dendritic spine density via an N-methyl-D-aspartate receptor-dependent mechanism. *J Neurosci* 1994; 14: 7680-7.
 385. Blanco G, Díaz H, Carrer HF, Beaugé L. Differentiation of rat hippocampal neurons induced by estrogen in vitro: effects on neuritogenesis and Na, K-ATPase activity. *J Neurosci Res* 1990; 27: 47-54.
 386. Woolley CS, Weiland NG, McEwen BS, Schwartzkroin PA. Estradiol increases the sensitivity of hippocampal CA1 pyramidal cells to NMDA receptor-mediated synaptic input: correlation with dendritic spine density. *J Neurosci* 1997; 17: 1848-59.
 387. Woolley CS, Wenzel HJ, Schwartzkroin PA. Estradiol increases the frequency of multiple synapse boutons in the hippocampal CA1 region of the adult female rat. *J Comp Neurol* 1996; 373: 108-17.
 388. Naftolin F, Leranthy C, Pérez J, García-Segura LM. Estrogen induces synaptic plasticity in adult primate neurons. *Neuroendocrinology* 1993; 57: 935-9.
 389. Murphy DD, Segal M. Morphological plasticity of dendritic spines in central neurons is mediated by activation of cAMP response element binding protein. *Proc Natl Acad Sci U S A* 1997; 94: 1482-7.
 390. Klintsova A, Levy WB, Desmond NL. Astrocytic volume fluctuates in the hippocampal CA1 region across the estrous cycle. *Brain Res* 1995; 690: 269-74.
 391. Warren SG, Humphreys AG, Juraska JM, Greenough WT. LTP varies across the estrous cycle: enhanced synaptic plasticity in proestrus rats. *Brain Res* 1995; 703: 26-30.
 392. Morse JK, DeKosky ST, Scheff SW. Neurotrophic effects of steroids on lesion-induced growth in the hippocampus. II. Hormone replacement. *Exp Neurol* 1992; 118: 47-52.
 393. Poirier J, Dea D, Baccichet A, Gauthier S. Modulation of gamma-actin and α -tubulin expression by corticosterone during neuronal plasticity in the hippocampus. *Mol Brain Res* 1992; 15: 263-8.
 394. O'Donnell D, Baccichet A, Seckl JR, Meaney MJ, Poirier J. Entorhinal cortex lesions transiently alter glucocorticoid but not mineralocorticoid receptor gene expression in the rat hippocampus. *J Neurochem* 1993; 61: 356-9.
 395. Bodnoff SR, Humphreys AG, Lehman JC, Diamond DM, Rose GM, Meaney MJ. Enduring effects of chronic corticosterone treatment on spatial learning, synaptic plasticity, and hippocampal neuropathology in young and mid-aged rats. *J Neurosci* 1995; 15: 61-9.
 396. Joseph R. Environmental influences on neural plasticity, the limbic system, emotional development and attachment: a review. *Child Psychiatry Hum Dev* 1998; 29: 189-208.
 397. Rosenzweig MR, Bennett EL. Psychobiology of plasticity: effects of training and experience on brain and behavior. *Behav Brain Res* 1996; 78: 57-65.
 398. Kolb B, Forgie M, Gibb R, Gorny G, Rowntree S. Age, experience and the changing brain. *Neurosci Biobehav Rev* 1998; 22: 143-59.

399. Paylor R, Morrison SK, Rudy JW, Waltrip LT, Wehner JM. Brief exposure to an enriched environment improves performance on the Morris water task and increases hippocampal cytosolic protein kinase C activity in young rats. *Behav Brain Res* 1992; 52: 49-59.
400. Johansson BB, Zhao L, Mattsson B. Environmental influence on gene expression and recovery from cerebral ischemia. *Acta Neurochir Suppl (Wien)* 1999; 73: 51-5.
401. Saito S, Kobayashi S, Ohashi Y, Igarashi M, Komiya Y, Ando S. Decreased synaptic density in aged brains and its prevention by rearing under enriched environment as revealed by synaptophysin contents. *J Neurosci Res* 1994; 39: 57-62.
402. Mohammed AH, Henriksson BG, Söderström S, Ebendal T, Olsson T, Seckl JR. Environmental influences on the central nervous system and their implications for the aging rat. *Behav Brain Res* 1993; 57: 183-91.
403. Chaillan FA, Roman FS, Soumireu-Mourat B. Modulation of synaptic plasticity in the hippocampus and piriform cortex by physiologically meaningful olfactory cues in an olfactory task. *J Physiol (Paris)* 1996; 90: 343-7.
404. Joublin F, Spengler F, Wacquant S, Dinse HR. A columnar model of somatosensory reorganizational plasticity based on Hebbian and non-Hebbian learning rules. *Biol Cybern* 1996; 74: 275-86.
405. Kirkwood A, Rioult MG, Bear MF. Experience-dependent modification of synaptic plasticity in visual cortex. *Nature* 1996; 381: 526-8.
406. Kaczmarek L, Chaudhuri A. Sensory regulation of immediate-early gene expression in mammalian visual cortex: implications for functional mapping and neural plasticity. *Brain Res Rev* 1997; 23: 237-56.
407. Kleim JA, Lussnig E, Schwarz ER, Comery TA, Greenough WT. Synaptogenesis and FOS expression in the motor cortex of the adult rat after motor skill learning. *J Neurosci* 1996; 16: 4529-35.
408. Liepert J, Miltner WH, Bauder H, Sommer M, Dettmers C, Taub E, et al. Motor cortex plasticity during constraint-induced movement therapy in stroke patients. *Neurosci Lett* 1998; 250: 5-8.
409. Petit TL, LeBoutillier JC, Markus EJ, Milgram NW. Synaptic structural plasticity following repetitive activation in the rat hippocampus. *Exp Neurol* 1989; 105: 72-9.
410. Lindholm D, Castren E, Berzaghi M, Blochl A, Thoenen H. Activity-dependent and hormonal regulation of neurotrophin mRNA levels in the brain: implications for neuronal plasticity. *J Neurobiol* 1994; 25: 1362-72.
411. Croll SD, Sharp PE, Bostock E. Evidence for NMDA receptor involvement in environmentally induced dentate gyrus plasticity. *Hippocampus* 1992; 2: 23-8.
412. Kilgard MP, Merzenich MM. Plasticity of temporal information processing in the primary auditory cortex. *Nat Neurosci* 1998; 1: 727-31.
413. Polley DB, Chen-Bee CH, Frostig RD. Two directions of plasticity in the sensory-deprived adult cortex. *Neuron* 1999; 24: 623-37.
414. Fordyce DE, Wehner JM. Physical activity enhances spatial learning performance with an associated alteration in hippocampal protein kinase C activity in C57BL/6 and DBA/2 mice. *Brain Res* 1993; 619: 111-9.
415. Russo-Neustadt A, Beard RC, Cotman CW. Exercise, antidepressant medications, and enhanced brain derived neurotrophic factor expression. *Neuropsychopharmacology* 1999; 21: 679-82.
416. Widenfalk J, Olson L, Thoren P. Deprived of habitual running, rats downregulate BDNF and TrkB messages in the brain. *Neurosci Res* 1999; 34: 125-32.
417. Van PH, Christie BR, Sejnowski TJ, Gage FH. Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc Natl Acad Sci U S A* 1999; 96: 13427-31.
418. Anderson BJ, Li X, Alcántara AA, Isaacs KR, Black JE, Greenough WT. Glial hypertrophy is associated with synaptogenesis following motor-skill learning, but not with angiogenesis following exercise. *Glia* 1994; 11: 73-80.
419. Kleim JA, Vij K, Ballard DH, Greenough WT. Learning-dependent synaptic modifications in the cerebellar cortex of the adult rat persist for at least four weeks. *J Neurosci* 1997; 17: 717-21.
420. Merzenich M, Wright B, Jenkins W, Xerri C, Byl N, Miller S, et al. Cortical plasticity underlying perceptual, motor, and cognitive skill development: implications for neurorehabilitation. *Cold Spring Harbor Symp Quant Biol* 1996; 61: 1-8.

MECANISMOS CELULARES DE LA NEUROPLASTICIDAD

Resumen. *Objetivo. Presentar, de manera unificada, una visión de los principales mecanismos de neuroplasticidad conocidos, destacando su universalidad. Desarrollo. La concepción del sistema nervioso como una entidad inmutable ha sufrido modificaciones sustanciales durante la segunda mitad del siglo XX. La neuroplasticidad, es decir, la capacidad de cambio y reparación del cerebro, se expresa de formas diversas, desde modificaciones funcionales de estructuras ya existentes, hasta la formación por crecimiento y proliferación de nuevas estructuras y neuronas. El presente trabajo aborda los mecanismos celulares y moleculares de los fenómenos neuroplásticos y los clasifica en dos grandes grupos: plasticidad por crecimiento, donde se incluyen los mecanismos de regeneración axonal, colateralización y sinaptogénesis reactiva; y plasticidad funcional, que abarca cambios en la eficacia de la transmisión sináptica como la potenciación a largo plazo y la activación de sinapsis silentes. Se presentan además algunas relaciones de fenómenos neuroplásticos con enfermedades del sistema nervioso, así como ejemplos de factores fisiológicos, físicos y farmacológicos que pueden, en el futuro, convertirse en herramientas terapéuticas para estimular y modular la neuroplasticidad. Conclusiones. Los mecanismos neuroplásticos muestran un alto grado de conservación filogenética y ontogenética, y son importantes tanto en la génesis de trastornos y enfermedades del sistema nervioso, como en su reparación tras sufrir traumatismos y daños muy diversos. La modulación de los mecanismos neuroplásticos por agentes físicos y químicos se vislumbra como una de las más potentes herramientas terapéuticas de la neurología restaurativa. [REV NEUROL 2000; 31: 1074-95] [http://www.revneurolog.com/3111/j111074.pdf]*

Palabras clave. Ácido orótico. Ambiente complejo. Colateralización. Ejercicio físico. Esteroides. Factores neurotróficos. Gangliósidos. Neurogénesis. Neuroplasticidad. Patología. Plasticidad cortical. Potenciación a largo plazo. Regeneración. Sinaptogénesis.

MECANISMOS DA NEUROPLASTICIDADE

Resumo. *Objetivo. Apresentar, de forma unificada, uma visão dos mecanismos principais de neuroplasticidade conhecidos, destacando sua universalidade. Desenvolvimento. A concepção do sistema nervoso, como uma entidade imutável, sofreu modificações significativas, durante a segunda metade do século XX. A neuroplasticidade, ou seja, a capacidade de mudança e reparação do cérebro, é expressada de modos diversos, desde modificações funcionais de estruturas já existentes, até a formação por crescimento e proliferação de novas estruturas e neurônios. O presente trabalho aborda os mecanismos celulares e moleculares dos fenômenos neuroplásticos e os classifica em dois grandes grupos: plasticidade por crescimento, onde são incluídos os mecanismos de regeneração axonal, colateralização e sinaptogénesis reativa. E plasticidade funcional que abarca mudanças na efetividade da transmissão sináptica como a potenciação a longo prazo e a ativação de sinapsis silentes. Apresentam-se, também, algumas relações de fenômenos neuroplásticos com enfermidades do sistema nervoso, como também exemplos de fatores fisiológicos, físicos e farmacológicos que podem, no futuro, tornar-se ferramentas terapêuticas para estimular e modular a neuroplasticidade. Conclusões. Os mecanismos neuroplásticos mostram um alto grau de conservação filogenética e ontogenética e são tão importantes na gênese de transtornos e enfermidades do sistema nervoso como em sua reparação, depois de sofrer traumatismos e danos muito diversos. A modulação dos mecanismos neuroplásticos por agentes físicos e químicos desponta como uma das mais potentes ferramentas terapêutica na neurologia restaurativa. [REV NEUROL 2000; 31: 1074-95] [http://www.revneurolog.com/3111/j111074.pdf]*

Palavras chave. Ácido orótico. Ambiente complexo. Colateralização. Esteróides. Exercício físico. Fatores neurotróficos. Gangliósidos. Neurogénesis. Neuroplasticidade. Patologia. Plasticidade cortical. Potenciação a longo prazo. Regeneração. Sinaptogénesis.

Aging and Synaptic Plasticity: A Review

Jorge A. Bergado and William Almaguer

International Center for Neurological Restoration. Havana, Cuba

SUMMARY

Aging affects all systems, but the brain seems to be particularly vulnerable to the action of negative, age-dependent factors. A gradual loss of memory functions is one of the earliest and most widespread consequences of brain aging. The causes for such impairment are still unclear. Long-term potentiation (LTP) is one form of neural plasticity, which has been proposed as the cellular correlate for memory. LTP is affected by aging, and such alteration might be causally related to memory dysfunction. In the present paper, we review the evidence sustaining the existence of a causal link between cognitive and LTP impairments, as well as the possible mechanisms involved. New results indicate a possible involvement of a deficient reinforcement of LTP by affective influences.

INTRODUCTION

Aging, in biological or cosmological dimension, is the most evident consequence of the passing of time. Time is one of the coordinates (along with space) that form the referential frame in which we exist. It is difficult to assess whether time itself has any influence on living organisms. More probable are thermodynamically irreversible processes that

occur in a timely organized sequence, those responsible for aging and for giving us the sense of the passing of time. In biological terms, the aging process is accompanied by a progressive multi-systemic deterioration which, in the absence of other factors (like accidents or disease), inevitably leads to death.

Although there appears to be little hope at present of reaching immortality, major improvements in living conditions and medical care during the last century have brought a steady and significant lengthening of human life (at least for a part of mankind). Consequently, one of the main goals of modern science is to provide a better quality to those years added to life by retarding, reducing, or eliminating (when possible) the negative consequences of aging. Any progress in this direction will be based on a better understanding of the mechanisms involved in the progressive impairments accompanying aging.

The preservation of memory abilities, in both senses, namely, the ability to learn new information and to retrieve old contents, is among the most desired because it is precisely one of the more severely affected by aging. A widely accepted view at present attributes memory formation and consolidation to functional modifications at synapses in different regions of the brain. Whereas the brain system serving particular forms of memory (declarative, motor skills, emotional, classical conditioning, and others) might be different, the basic mechanism modifying synaptic efficacy seems to be common (Matthies, 1998; Medina et al., 2002; Milner et al., 1998).

Prof. Jorge A. Bergado, International Center for Neurological Restoration (CIREN) Ave. 25 # 15805 11300 Playa Ciudad de La Habana, Cuba; e-mail: bergado@neubas.sld.cu

Long-term potentiation (LTP) of synaptic efficacy (Bliss & Gardner-Medwin, 1973; Bliss & Lomo, 1973) is a long-lasting increase in synaptic efficacy after high frequency (tetanic) stimulus, and appears as the best candidate mechanism for explaining the learning-induced changes in synaptic connectivity. A growing body of evidence links LTP and memory. This evidence is based, not only on the suggestive and numerous analogies between both phenomena but also on direct experiments demonstrating a relation between changes in synaptic efficacy and memory in different brain structures as well (Bergado et al., 1988; Matthies et al., 1986; Rioult-Pedotti et al., 1998; Rogan et al., 1997) (for a recent review on the subject see Martin & Morris, 2002). It is plausible, therefore, that age-related disorders in synaptic plasticity might be functionally linked to memory impairment (Shapiro, 2001). These concepts are beginning to gain consideration for the effective prevention or treatment of such impairments (Rosenzweig, 1996).

In the present review we will focus our attention on describing how LTP is affected by aging, stressing new results linking memory and emotional disorders with the modulation of LTP in aged animals.

THE GENERAL PROBLEM OF AGING

Several hypotheses have been forwarded to explain the causes of aging. General theories emphasize the role of alterations in biological membranes, the attack of free radicals, calcium dysregulation, and (particularly in mammals) the negative effect of glucocorticoids and stress (Lynch, 1998b). Obviously, all the mentioned factors strongly interact with each other and might potentiate reciprocally, obscuring the possibility to develop a testable hypothesis regarding which might be the primary events in aging.

What appears clear is that aging is a process, and that aging is caused by the action of multiple factors. Aging affects all systems: Muscles weaken, bones become fragile, skin shrinks, arteries harden, hormones (particularly sex hormones) decay, immunity fades and memory fails.

Aging and the nervous system

The alterations in nervous system function as a consequence of aging are diverse. Aged persons (and animals) move different, sleep different, and show alterations in mood and memory. Neuron death and loss of afferents have been proposed as primary events leading to brain dysfunction (Morrison & Hof, 1997; Ward et al., 1999). Although neuron death in specific brain regions is common in neurodegenerative diseases (Barili et al., 1998; Gerlach & Riederer, 1996; Graeber et al., 1998; Landfield et al., 1992; Morrison & Hof, 1997; West, 1993), the relative contribution of this factor to impairment in normal aging is a matter of debate (Rapp & Gallagher, 1996; West, 1993). Non-neuronal alterations like an impaired blood flow (Ajmani et al., 2000) or glial dysfunction (Sykova, 2001) should also contribute to a generalized brain malfunction.

Aging and memory impairment

The decline in cognitive capacities in humans is one of the earliest, most dramatic, and generalized consequences of aging. Even in normal aged persons, memory dysfunctions limit intellectual abilities. In several pathological, age-related conditions, whether vascular or neuro-degenerative like Alzheimer's disease and other dementias, the impact of age on cognition is devastating. The literature showing memory alterations as a consequence of aging in rodents (Ando & Ohashi, 1991; Kadar et al., 1990; Luparini et al., 2000; Ward et al., 1999; Ward et al., 1999), as well as in humans (Albert, 1997; Langley & Madden, 2000; Soininen

et al., 1994) and non-human primates (Bachevalier, Landis et al., 1991) is extensive. The cholinergic hypothesis of geriatric memory dysfunction (Bartus, 2000; Bartus et al., 1982) stresses the importance of the cholinergic afferents arising from the basal forebrain in memory functions (Baxter et al., 1999; Ikegami, 1994; Ikegami et al., 1992; Pedigo Jr, 1994; Russell, 1996; Shen et al., 1996; Smith et al., 1995) and has inspired some promising efforts in the search for effective treatments to overcome the age-related cognitive impairments (Fernández et al., 1994; Fischer, 1994; Fischer et al., 1994; Flood et al., 1996; Garrone et al., 1998; Levin & Torry, 1996; Scali et al., 1994; Sirvio, 1999; Smith et al., 1999; Vannucchi et al., 1997). Despite the intensive effort and literature supporting the cholinergic hypothesis, no clear picture has emerged on how acetylcholine might act to support memory processing.

When the methodological basis of animal learning models was established in the early years of the 20th Century, it was immediately recognized that for an animal to learn something a strong motivation must be present. Much less attention has been paid, however, both clinically and experimentally, to the possible impact of dysfunctional emotional-motivational reactions on cognitive performance. Such neglect might appear surprising considering that aged related alterations in mood are widespread (Blazer, 1987) and often associated with neurodegenerative diseases (Harwood et al., 2000).

AGING AND SYNAPTIC PLASTICITY

Mechanisms of plasticity

Decades of intensive research have contributed to clarify the mechanisms involved in the induction, development, and maintenance of LTP. Although different forms of LTP seem to exist—according to the induction mechanisms involved—the most common form requires the activation of

NMDA-type glutamate receptors (Bashir et al., 1991; Bashir et al., 1990; Collingridge, 1987; 1992). The NMDA ionophore allows the entrance of calcium to the postsynaptic region, an event that seems decisive to the strength and direction of the plastic modification. When the increase in Ca^{++} is above certain values, an increase in synaptic efficacy (i.e., LTP) develops, but lower values could lead to the opposite, a reduction in synaptic efficacy (the so called Long-Term Depression, LTD) (Cormier et al., 2001; Foster & Norris, 1997; Teyler et al., 1994). In LTP, Ca^{++} activates protein kinases; whereas phosphatases are activated in LTD.

Several kinases have been proved to participate in the event cascade involved in LTP; among them, the Ca^{++} -calmodulin dependent protein kinase II (PKII), the Ca^{++} -phospholipid dependent protein kinase C (PKC), and the cyclic AMP dependent protein kinase A (PKA) (Abel et al., 1997; Colley et al., 1990; Fukunaga et al., 1996; Huang & Kandel, 1994; Matthies & Reymann, 1993; Reymann et al., 1988a; 1988b; Silva et al., 1992; Stevens et al., 1994). These enzymes can modify synaptic efficacy by acting pre- and post-synaptically. For example, PKII can increase the conductance of the AMPA-glutamate receptors (Derkach et al., 1999), and consequently, increase the level of post-synaptic depolarization. But they can also act at distant regions, like the nucleus, regulating gene expression and protein synthesis (Nguyen & Kandel, 1996; 1997; Pontzer et al., 1990).

The dependency of late phases of LTP (L-LTP, >4 h) on protein synthesis is well established (Frey et al., 1988; Krug et al., 1984; Otani & Abraham, 1989). Such dependency has led to a multi-phase model of LTP, similar to that proposed for memory (Matthies et al., 1990). Yet, the identification of specific proteins required for LTP late maintenance has proven difficult. It seems that in a first step, proto-oncogenes are activated (Abraham et al., 1991; Bishop et al., 1994; Roberts et al., 1996), which can, in turn, activate structural genes; like

those coding subunits of AMPA receptors (Desmond & Weinberg, 1998).

The mechanism to guarantee LTP specificity after massive, non-specific protein synthesis requires the setting of a tag at the activated synapses, so that arriving proteins can be 'captured' only by those synapses that have been previously activated above a certain limit (Frey & Morris, 1997; 1998).

Although the incorporation of new AMPA receptors might well subserve an increased synaptic efficacy in potentiated synapses, one can't help but wonder why so many other proteins are needed. One likely explanation is that really long, long-lasting changes in synaptic efficacy are accompanied by morphological changes at the existing synapses—a possibility raised in early studies (Desmond & Levy, 1986a; 1986b; 1988; Fifkova, 1985)—or the growth and establishment of new functional synaptic contacts. Electron microscope studies reveal signs of dendritic spine division and axonal sprouting days after induction of LTP by strong stimuli (Geinisman, 1993; Geinisman et al., 1989; Yuste & Bonhoeffer, 2001).

Recent developments have shown that the plastic processes initiated by the activation of specific glutamatergic synapses can be modulated by synapses different from those previously involved in LTP induction (i.e., hetero-synaptically). This point is conceptually important because LTP has been considered a homosynaptic phenomenon, despite evidence indicating the need for hetero-synaptic cooperation. An impaired LTP has been reported in animals bearing lesions of the fimbria-fornix, the fiber system providing the hippocampus with cholinergic and aminergic fibers of subcortical origin (Bergado et al., 1996; Buzsáki & Gage, 1989). Cooperation between the perforant pathway (mainly glutamatergic) and the septum (mainly non-glutamatergic) and the locus coeruleus (mainly noradrenergic) has also been documented (Harley & Sara, 1992; Kitchigina et al., 1997;

Robinson & Racine, 1982; 1986).

More recently, a group of studies has shown that the activation of the amygdala—a limbic structure related to emotion (Quirk et al., 1996)—can influence the induction of LTP at the dentate gyrus (Akirav & Richter-Levin, 1999; Ikegaya et al., 1995; Ikegaya et al., 1996). We have recently shown that the electrical stimulation of the amygdala also contributes to the maintenance of LTP, converting a short lasting early-LTP (E-LTP <4h) into a late-LTP (L-LTP >4h) (Frey et al., 2001). Interestingly, the effect is protein-synthesis dependent and requires the activity of noradrenergic and cholinergic inputs. Lesioning of the fimbria-fornix, the main source of subcortical aminergic and cholinergic input to the hippocampal formation, impairs the reinforcing effect of the amygdala stimulation on L-LTP (Abe et al., 1998; Jas et al., 2000). The activity of the amygdala also seems crucial for the so-called behavioral reinforcement of LTP. According to this paradigm, E-LTP can also be converted into an L-LTP by giving the animals a behavioral stimulus with a strong motivational content (i.e. drinking after 24-h water deprivation) shortly before, or after, inducing LTP using a weak tetanus (3x15 impulses at 200 Hz) (Seidenbecher et al., 1995; 1997). We have recently obtained evidence that lesioning, or blocking the amygdala with lidocaine, abolishes the behavioral reinforcement of LTP (Almaguer et al., in press).

AGE-RELATED CHANGES OF SYNAPTIC PLASTICITY

Developmental changes

Like many other functions, synaptic plasticity presumably shows developmental changes during the early stages of postnatal life, followed by a period of relative stability in adulthood and then deteriorating slowly at older ages.

LTP is difficult to induce before postnatal day 5. At P15 a level similar to adults can be reached (Teyler et al., 1989), but outlasting in time the duration of LTP in adults under similar induction paradigms (Bronzino et al., 1994; 1995). Younger animals, on the other hand, seemed to be more prone to develop LTD (Dudek & Bear, 1993; Wasling et al., 2002).

Older animals show impaired synaptic plasticity

Although earlier reports pointed to a reduced synaptic plasticity accompanying age-dependent memory impairment (see for example (Landfield et al., 1972), a demonstration of LTP deterioration with aging came from a series of elegant studies from Barnes and McNaughton (1985) (according to the references available to us). These authors demonstrated an impressive parallelism between the slower rate of development and the faster decay of memory and LTP among aged rats, compared with young controls (Barnes, 1979; Barnes & McNaughton, 1985). This result was later confirmed by others (Bergado et al., 1997; de Toledo-Morrell & Morrell, 1991; de Toledo-Morrell et al., 1988; Diana, De Carolis et al., 1994; Geinisman, de Toledo-Morrell et al., 1995) showing that behavioral, electrophysiological, and histological alterations were more profound in old rats with memory impairments than in similarly old animals without cognitive impairments. Later studies showed that aged rats were also more prone to develop LTD or to reverse LTP (depotentialization) by low frequency stimulation (Norris et al., 1996).

The former doesn't mean that aged rats are unable to develop LTP. When adequately stimulated, aged rats can reach an increase in synaptic efficacy, similar to that of young animals (Diana et al., 1994; Moore et al., 1993)—even those showing memory impairment—but the former seem to require stronger stimulation, and the decay rate is faster. This phenomenon has been attributed to an

impaired "ability of aged rats' synapses to provide the sustained depolarization necessary to activate the LTP-induction cascade" (Rosenzweig et al., 1997).

POSSIBLE MECHANISMS OF IMPAIRMENT

Impaired induction?

Considering the importance of NMDA receptor activation for LTP induction, it seems plausible that any impairment in glutamatergic systems would have an impact on synaptic plasticity. In patients with Alzheimer's disease, neuropathology studies have shown that even in the early stages, the number of neurons in the entorhinal cortex is reduced (Braak & Braak, 1992; Palmer, 2002). The evidence from aged animals is not so clear, at least regarding the total number of cells originating the perforant pathway (Gazzaley et al., 1997; Merrill et al., 2000; 2001). In the rat, however, there are indications of a reduced glutamatergic influence on the EPSP (Barnes et al., 1997; Billard et al., 1997). This decline might be related to a reduction in glutamate receptors in the cortex and in the hippocampus—particularly of the NMDA type—that have been repeatedly reported in rats (Kito et al., 1990; Liang & Lu, 1992; Magnusson, 1998; Magnusson & Cotman, 1993; Tamaru et al., 1991; Wenk et al., 1991) and probably associated with a change in the subunit composition (Clayton & Browning, 2001; Clayton et al., 2002; Magnusson et al., 2002) and subtle subregional variations within hippocampal subfields (Wenk & Barnes, 2000). The activity-dependent redistribution of NMDA receptors is also affected among aged rats (Clayton et al., 2002). Nevertheless, the basic mechanisms of NMDA-receptor induction mechanism for LTP seemed to be preserved (Barnes et al., 1996).

Aging seems to provoke a shift from NMDA-

receptor calcium influx to Voltage Dependent Calcium Channels (VDCC) (Shankar et al., 1998), with the probable consequence that the increase in intracellular calcium will not reach the level required to activate Ca^{++} -dependent kinases, but sufficient to activate phosphatases (Foster, 1999; Foster & Norris, 1997). This situation would lead to the prediction that LTD and depotentiation are facilitated in aged rats, for which there is some experimental support (Norris et al., 1996). That blockade of VDCC attenuates this altered plasticity is in line with this hypothesis (Norris et al., 1998).

Other indications point to impaired regulation of protein kinases in aged animals. For PKII, a dysfunctional regulation of the alpha subunit has been reported in aged animals showing a decremental LTP (Davis et al., 2000), in contrast to those maintaining LTP beyond 3 h. An age-related decrease of cortical plasticity was found in mutant mice lacking the αCaMKII subunit (Kirkwood et al., 1997). Similarly, a reduced PKC activity and its substrates has been experimentally documented in aged animals (Casoli et al., 1996; Chang et al., 1997; Mons et al., 2001; Okuma et al., 2000).

Impaired maintenance?

The temporal course of LTP in aged animals, particularly its faster decay, suggests an absent protein-synthesis dependent L-LTP. Several reports show alteration in protein synthesis and expression pattern or early genes among aged rats after LTP induction (Lanahan et al., 1997; Mullany & Lynch, 1997). An impaired gene activation and protein synthesis might, in turn, affect the ability of axons and dendritic spines to divide, grow, and form new contacts that apparently are required for very long-lasting LTP. In aged rats, electron micro-scopic studies have evidenced that this capacity might be quantitatively altered (Chang et al., 1991; Geinisman et al., 1992). In line with this evidence are findings showing that the expression of cell

adhesion molecules required for axonal growth is impaired in aged rats (Ronn et al., 2000), indicating a deficient maintenance of LTP (Lynch & Voss, 1994).

Impaired heterosynaptic reinforcement?

As mentioned previously, the activation of heterosynaptic afferents (i.e. behavioral or amygdala-induced reinforcement) to a neuronal population, in which an E-LTP has been induced through a mild tetanus, can prolong the duration of the plastic process, converting it into an L-LTP that is protein-synthesis dependent. This effect suggests that the activation of such afferents is able to induce the activity of molecular cascades, which complement the insufficiently activated cascade by the previous glutamatergic tetanus. Norepinephrine and acetylcholine have been implicated in those effects (Frey et al., 2001). Both transmitters have been shown to induce a slowly developing potentiation in *in vitro* studies, which might correlate with their effects on L-LTP. Norepinephrine and acetylcholine, when applied to slices obtained from young animals, induce a slowly developing potentiation (Segal & Auerbach, 1997; Stanton & Sarvey, 1987), which might represent an L-LTP. The effect of NE is protein synthesis dependent (Stanton & Sarvey, 1985). Acetylcholine is responsible for theta rhythm in the hippocampus. The magnitude of LTP induced by tetani delivered at the positive theta peak was reduced in aged animals rats (Orr et al., 2001).

Recent results from our group provided the first evidence that such heterosynaptic reinforcing mechanisms might be deficient in aged rats, particularly among those animals showing memory deficits in a spatial task. Both behavioral- and amygdala-induced reinforcement are impaired, not only in cognitively deficient aged rats but also among aged rats showing no memory impairment in the Morris water maze (Almaguer et al., 2002;

Bergado et al., 2001). Both noradrenergic and cholinergic systems in the brain are affected by aging (Baxter et al., 1999; Chouinard et al., 1995; Friedman & Duckles, 1994; Isacson et al., 2002; Luine et al., 1990; Powers et al., 1988; Sherman & Friedman, 1990; Stemmelin et al., 2000) and both can activate kinase systems involved in regulating protein synthesis (Bevilaqua et al., 1997; Stratton et al., 1988).

We assume that the deficient reinforcement of LTP by amygdala or by behavioral stimulation is caused by a reduction in noradrenergic and/or cholinergic afferents to the dentate gyrus, the brain area in which both studies were carried out.

The results of these studies provide a rationale, at the cellular level, for a possible functional link between two of the major consequences of aging: memory impairment and affective dystonia.

Additional factors might increase the negative impact of aging on synaptic plasticity. Chronic stress has been shown to lead to a disruption in LTP (Sapolsky, 1999) by the excessive production of glucocorticoids (Bodnoff et al., 1995; McEwen, 1994), to which aged rats seemed particularly sensitive. Also, inflammatory cytokines (Lynch, 1998a) have been shown to affect LTP in an age-dependent manner.

Whereas a general agreement seems to have been reached on the relation between age-related impairments in plasticity and memory, some debate remains about which of the factors mentioned plays the major role in the degenerative process. We sustain the view that, most likely, all factors can be involved and interact cooperatively, leading to a continuously deteriorating plastic ability that causes, in turn, the decay in mnemonic capacities.

LINKS TO PATHOLOGY?

Another important aspect to consider is whether alterations in synaptic plasticity might be

also involved in memory losses in pathological forms of aging like Alzheimer's disease. New developments in animal models have provided indications that such a relation might exist.

The RNA for the amyloid precursor protein (APP), one of the key molecules in Alzheimer's pathogenesis, is upregulated after LTP induction in young rats, but not among aged rats regardless of whether they were cognitively impaired or not (Stephan et al., 2002). On the other hand, mutant mice lacking the APP gene, and in mice expressing the carboxy terminus of APP, released after proteolytic cleavage of the protein, showed an altered LTP maintenance (Nalbantoglu et al., 1997; Seabrook, Smith et al., 1999). Moreover, mice overexpressing mutant forms of human APP developed not only the typical histopathology of Alzheimer's but also an altered plasticity (Chapman et al., 1999; Larson et al., 1999) (see however (Fitzjohn et al., 2001).

AN IMPACT ON THERAPEUTICS?

The studies on the alterations of synaptic plasticity in aging have also provided some results of potential therapeutic implications.

General factors like caloric restriction (Eckles-Smith et al., 2000) or dietary supplement with antioxidants (vitamin E or lipoic acid) (Murray & Lynch, 1998; McGahon et al., 1999) reverse age-dependent alterations in LTP, and improve glutamatergic transmission in the dentate gyrus. The use of calcium regulators like nimodipine or nifedipine has shown beneficial effects on LTP among aged rats (De Jong et al., 1992; Norris et al., 1998), and cholinergic agonists have improved declining LTP in aged rats (Fujii & Sumikawa, 2001).

Trophic factors are among the greatest hopes of restorative neurology in the last decades. Some have shown a direct effect on synaptic plasticity. Early reports suggested an action for epidermal

growth factor (EGF) and fibroblast growth factor (FGF), but not nerve growth factors (NGF), promoting LTP in normal and lesioned rats (Abe et al., 1992; Ishiyama et al., 1991; Terlau & Seifert, 1990). More recently, the results of *in vitro* and *in vivo* studies have shown that brain-derived neurotrophic factor (BDNF) or neurotrophin-3 (NT-3), but again not NGF, are able to induce an LTP-like plastic development in the hippocampus (Chen et al., 1999; Kang & Schuman, 1995a; 1995b; 1996; Kovalchuk et al., 2002; Lu & Chow, 1999), thus re-opening the issue for a potential use of BDNF for the treatment of age-related memory impairments. Although NGF cannot by itself induce LTP, chronic treatment with NGF restores the ability of aged, memory-deficient rats to develop LTP (Bergado et al., 1997; Bergado et al., 1998), a result that correlates with behavioral studies showing an improvement in memory function after similar treatment (Fischer et al., 1994). We interpret this effect as mediated by the protective and restorative function of NGF on the basal forebrain cholinergic projection to the hippocampus and cortex. Similar results have been reported after the transplantation of septal fetal neurons to aged rats, or in rats with lesion of the septo-hippocampal projection (Bergado et al., 1997; Björklund & Stenevi, 1977; Fernández et al., 1994; Leanza et al., 1998). Although neurotrophic administration to the brain is technically difficult, an intensive search for new strategies could bring some promising results to this important field.

CONCLUSIONS

The accumulated evidence seems reasonably sufficient to postulate the existence of a functional link between the age-dependent impairment in synaptic plasticity and the deterioration of memory function. The knowledge about the mechanisms of LTP can lead not only to a better understanding of

the causality of memory decline with age but also to the development of new therapeutic strategies for treating it. Several lines of evidence also point to an impaired relation, at the cellular level, between affective and cognitive processes. Any progress in these important topics can represent a substantial contribution to reach the goal of successful aging, so that the years gained for life can also be an enjoyable period of existence.

REFERENCES

- Abe K, Ishiyama J, Saito H. 1992. Effects of epidermal growth factor and basic fibroblast growth factor on generation of long-term potentiation in the dentate gyrus of fimbria-fornix-lesioned rats. *Brain Res* 593: 335–338.
- Abe K, Noguchi K, Saito H. 1998. Medial amygdala-induced spike potentiation in the rat dentate gyrus is dependent on N-methyl-D-aspartate receptors and subcortical afferents. *Neurosci Lett* 246: 85–88.
- Abel T, Nguyen PV, Barad M, Deuel TAS, Kandel ER. 1997. Genetic demonstration of a role for PKA in the late phase of LTP and in hippocampus-based long-term memory. *Cell* 88: 615–626.
- Abraham WC, Dragunow M, Tate WP. 1991. The role of immediate early genes in the stabilization of long-term potentiation. *Mol Neurobiol* 5: 297–314.
- Ajmani RS, Metter EJ, Jaykumar R, Ingram DK, Spangler EL, et al. 2000. Hemodynamic changes during aging associated with cerebral blood flow and impaired cognitive function. *Neurobiol Aging* 21: 257–269.
- Akirav I, Richter-Levin G. 1999. Priming stimulation in the basolateral amygdala modulates synaptic plasticity in the rat dentate gyrus. *Neurosci Lett* 270: 83–86.
- Albert MS. The ageing brain: normal and abnormal memory. 1997. *Philos Trans R Soc Lond B Biol Sci* 352: 1703–1709.
- Almaguer W, Estupiñán B, Frey JU, Bergado JA. 2002. Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP. *Neurobiol*

- Aging 23: 319–324.
- Almaguer-Melian W, Martinez-Marti L, Frey JU, Bergado JA. In press. The amygdala is part of the behavioral reinforcement system required for LTP in rat hippocampus. *Neuroscience*.
- Ando S, Ohashi Y. 1991. Longitudinal study on age-related changes of working and reference memory in the rat. *Neurosci Lett* 128: 17–20.
- Bachevalier J, Landis LS, Walker LC, Brickson M, Mishkin M, Price DL, Cork LC. 1991. Aged monkeys exhibit behavioral deficits indicative of widespread cerebral dysfunction. *Neurobiol Aging* 12: 99–111.
- Barili P, De Carolis G, Zaccheo D, Amenta F. 1998. Sensitivity to ageing of the limbic dopaminergic system: a review. *Mech Ageing Dev* 106: 57–92.
- Barnes CA. 1979. Memory deficits associated with senescence: A neurophysiological and behavioural study in the rat. *J Comp Physiol Psychol* 93: 74–104.
- Barnes CA, McNaughton BL. 1985. An age comparison of the rates of acquisition and forgetting. *Behav Neurosci* 99: 1040–1048.
- Barnes CA, Rao G, McNaughton BL. 1996. Functional integrity of NMDA-dependent LTP induction mechanisms across the lifespan of F-344 rats. *Learn Mem* 3: 124–137.
- Barnes CA, Rao G, Shen J. 1997. Age-related decrease in the N-methyl-D-aspartate_R-mediated excitatory postsynaptic potential in hippocampal region CA1. *Neurobiol Aging* 18: 445–452.
- Bartus RT. 2000. On neurodegenerative diseases, models, and treatment strategies: lessons learned and lessons forgotten a generation following the cholinergic hypothesis. *Exp Neurol* 163: 495–529.
- Bartus RT, Dean III RL, Beer B, Lippa AS. 1982. The cholinergic hypothesis geriatric memory dysfunction. *Science* 217: 408–417.
- Bashir ZI, Alford S, Davies SN, Randall AD, Collingridge GL. 1991. Long-term potentiation of NMDA receptor-mediated synaptic transmission in the hippocampus. *Nature* 349: 156–158.
- Bashir ZI, Tam B, Collingridge GL. 1990. Activation of the glycine site in the NMDA receptor is necessary for the induction of LTP. *Neurosci Lett* 108: 261–266.
- Baxter MG, Frick KM, Price DL, Breckler SJ, Markowska AL, Gorman LK. 1999. Presynaptic markers of cholinergic function in the rat brain: relationship with age and cognitive status. *Neuroscience* 89: 771–779.
- Bergado JA, Almaguer W, Ravelo J, Rosillo JC, Frey JU. 2001. Behavioral reinforcement of long-term potentiation is impaired in aged rats with cognitive deficiencies. *Neuroscience* 108: 1–5.
- Bergado JA, Fernández CI, Gómez-Soria A, González O. 1997. Chronic intraventricular infusion with NGF improves LTP in old cognitively-impaired rats. *Brain Res* 770: 1–9.
- Bergado JA, Gómez-Soria A, Cruz R, Fernandez CI. 1998. Nerve growth factor improves evoked potentials and long-term potentiation in the dentate gyrus of presenile rats. *Eur J Pharmacol* 345: 181–184.
- Bergado JA, Krug M, Rutherich H, Matthies H. 1988. Orotate improves memory and enhances synaptic long-term potentiation in active avoidance behaviour in rats with perforant path stimulation as the conditioned stimulus. *Eur J Pharmacol* 157: 155–163.
- Bergado JA, Moreno H, Nuñez N. 1996. Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biol Res* 29: 197–202.
- Bergado JA, Moreno H, Soto J, Castellano O, Castillo L. 1997. Septal fetal tissue transplants restore long-term potentiation in the dentate gyrus of fimbria-fornix-lesioned rats. *J Neural Transplant Plast* 6: 31–40.
- Bevilaqua L, Ardenghi P, Schröder N, Bromberg E, Schmitz PK, Schaeffer E, et al. 1997. Drugs acting upon the cyclic adenosine monophosphate protein kinase A signalling pathway modulate memory consolidation when given late after training into rat hippocampus but not amygdala. *Behav Pharmacol* 8: 331–338.
- Billard JM, Jouvenceau A, Lamour Y, Dutar P. 1997. NMDA receptor activation in the aged rat: electrophysiological investigations in the CA1 area of the hippocampal slice ex vivo. *Neurobiol Aging* 18: 535–542.
- Bishop JM, Capobianco AJ, Doyle HJ, Finney RE, McMahon M, Robbins SM, et al. 1994. Proto-oncogenes and plasticity in cell signaling. *Cold Spring Harbor Symp Quant Biol* 59: 165–172.
- Björklund A, Stenevi U. 1977. Reformation of the severed septohippocampal cholinergic pathway in the adult rat by transplanted septal neurons. *Cell Tissue Res* 185: 289–302.
- Blazer D. 1989. Depression in the elderly. *N Engl J Med* 320: 164–166.

- Bliss TV, Gardner-Medwin AR. 1973. Long-lasting potentiation of synaptic transmission in the dentate area of the unanaesthetized rabbit following stimulation of the perforant path. *J Physiol (Lond)* 232: 357–374.
- Bliss TV, Lomo T. 1973. Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol (Lond)* 232: 331–356.
- Bodnoff SR, Humphreys AG, Lehman JC, Diamond DM, Rose GM, Meaney MJ. 1995. Enduring effects of chronic corticosterone treatment on spatial learning, synaptic plasticity, and hippocampal neuropathology in young and mid-aged rats. *J Neurosci* 15: 61–69.
- Braak H, Braak E. 1992. The human entorhinal cortex: normal morphology and lamina-specific pathology in various diseases. *Neurosci Res* 15: 6–31.
- Bronzino JD, Abu-Hasaballah K, Austin-LaFrance RJ, Morgane PJ. 1994. Maturation of long-term potentiation in the hippocampal dentate gyrus of the freely moving rat. *Hippocampus* 4: 439–446.
- Bronzino JD, Abu-Hasaballah K, Austin-LaFrance RJ, Morgane PJ. 1995. Quantitative analysis of long-term potentiation in the hippocampal dentate gyrus of the freely-moving 15-day-old rat. *Brain Res Bull* 36: 321–324.
- Buzsáki G, Gage FH. 1989. Absence of long-term potentiation in the subcortically deafferented dentate gyrus. *Brain Res* 484: 94–101.
- Casoli T, Spagna C, Fattoretti P, Gesuita R, Bertoni-Freddari C. 1996. Neuronal plasticity in aging: a quantitative immunohistochemical study of GAP-43 distribution in discrete regions of the rat brain. *Brain Res* 714: 111–117.
- Chang JW, Schumacher E, Coulter PM, Vinters HV, Watson JB. 1997. Dendritic translocation of RC3/neurogranin mRNA in normal aging, Alzheimer disease and fronto-temporal dementia. *J Neuropathol Exp Neurol* 56: 1105–1118.
- Chang PL, Isaacs KR, Greenough WT. 1991. Synapse formation occurs in association with the induction of long-term potentiation in two-year-old rat hippocampus in vitro. *Neurobiol Aging* 12: 517–522.
- Chapman PF, White GL, Jones MW, Cooper-Blacketer D, Marshall VJ, Irizarry M, et al. 1999. Impaired synaptic plasticity and learning in aged amyloid precursor protein transgenic mice. *Nat Neurosci* 2: 271–276.
- Chen G, Kolbeck R, Barde YA, Bonhoeffer T, Kossel A. 1999. Relative contribution of endogenous neurotrophins in hippocampal long-term potentiation. *J Neurosci* 19: 7983–7990.
- Chouinard ML, Gallagher M, Yasuda RP, Wolfe BB, McKinney M. 1995. Hippocampal muscarinic receptor function in spatial learning- impaired aged rats. *Neurobiol Aging* 16: 955–963.
- Clayton DA, Browning MD. 2001. Deficits in the expression of the NR2B subunit in the hippocampus of aged Fisher 344 rats. *Neurobiol Aging* 22: 165–168.
- Clayton DA, Grosshans DR, Browning MD. 2002. Aging and surface expression of hippocampal NMDA receptors. *J Biol Chem* 277: 14367–14369.
- Clayton DA, Mesches MH, Alvarez E, Bickford PC, Browning MD. 2002. A hippocampal NR2B deficit can mimic age-related changes in long-term potentiation and spatial learning in the Fischer 344 rat. *J Neurosci* 22: 3628–3637.
- Colley PA, Sheu FS, Routtenberg A. 1990. Inhibition of protein kinase C blocks two components of LTP persistence, leaving initial potentiation intact. *J Neurosci* 10: 3353–3360.
- Collingridge G. 1987. The role of NMDA receptors in learning and memory. *Nature* 330: 604–605.
- Collingridge GL. 1992. The mechanism of induction of NMDA receptor-dependent long-term potentiation in the hippocampus. *Exp Physiol* 77: 771–797.
- Cormier RJ, Greenwood AC, Connor JA. 2001. Bidirectional synaptic plasticity correlated with the magnitude of dendritic calcium transients above a threshold. *J Neurophysiol* 85: 399–406.
- Davis S, Salin H, Helme-Guizon A, Dumas S, Stephan A, et al. 2000. Dysfunctional regulation of alphaCaMKII and syntaxin 1B transcription after induction of LTP in the aged rat. *Eur J Neurosci* 12: 3276–3282.
- De Jong GI, Buwalda B, Schuurman T, Luiten PG. 1992. Synaptic plasticity in the dentate gyrus of aged rats is altered after chronic nimodipine application. *Brain Res* 596: 345–348.
- Derkach V, Barria A, Soderling TR. 1999. Ca^{2+} /calmodulin-kinase II enhances channel conductance of alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionate type glutamate receptors. *Proc Natl Acad Sci USA* 96: 3269–3274.
- Desmond NL, Levy WB. 1986a. Changes in the numerical density of synaptic contacts with long-

- term potentiation in the hippocampal dentate gyrus. *J Comp Neurol* 253: 466–475.
- Desmond NL, Levy WB. 1986b. Changes in the postsynaptic density with long-term potentiation in the dentate gyrus. *J Comp Neurol* 253: 476–482.
- Desmond NL, Levy WB. 1988. Synaptic interface surface area increases with long-term potentiation in the hippocampal dentate gyrus. *Brain Res* 453: 308–314.
- Desmond NL, Weinberg RJ. 1998. Enhanced expression of AMPA receptor protein at perforated axospinous synapses. *Neuroreport* 9: 857–860.
- deToledo-Morrell L, Geinisman Y, Morrell F. 1988. Age-dependent alterations in hippocampal synaptic plasticity: relation to memory disorders. *Neurobiol Aging* 9: 581–590.
- deToledo-Morrell L, Morrell F. 1991. Age-related alterations in long-term potentiation and susceptibility to kindling. In: Morrell F, ed, *Kindling and synaptic plasticity: the legacy of Graham Goddard*. Boston, Massachusetts, USA: Birkhäuser; 160–175.
- Diana G, Scotti de Carolis A, Frank C, Domenici MR, Sagratella S. 1994. Selective reduction of hippocampal dentate frequency potentiation in aged rats with impaired place learning. *Brain Res Bull* 35: 107–111.
- Diana G, Domenici MR, Loizzo A, Scotti de Carolis A, Sagratella S. 1994. Age and strain differences in rat place learning and hippocampal dentate gyrus frequency-potentiation. *Neurosci Lett* 171: 113–116.
- Dudek SM, Bear MF. 1993. Bidirectional long-term modification of synaptic effectiveness in the adult and immature hippocampus. *J Neurosci* 13: 2910–2918.
- Eckles-Smith K, Clayton D, Bickford P, Browning MD. 2000. Caloric restriction prevents age-related deficits in LTP and in NMDA receptor expression. *Brain Res Mol Brain Res* 78: 154–162.
- Fernandez CI, Bergado J, De La Quetara K, Nunez N, Castellanos O, Gonzalez O, et al. 1994. Brain aging and neurotransplantation. II. Effects of septal cell suspension grafts to hippocampus of aged rodents on learning and memory impairments. *Arch Gerontol Geriatr* 4 (Suppl): 59–65.
- Fifkova E. 1985. A possible mechanism of morphometric changes in dendritic spines induced by stimulation. *Cell Mol Neurobiol* 5: 47–63.
- Fischer W. 1994. Nerve growth factor reverses spatial memory impairments in aged rats. *Neurochem Int* 25: 47–52.
- Fischer W, Sirevaag A, Wiegand SJ, Lindsay RM, Bjorklund A. 1994. Reversal of spatial memory impairments in aged rats by nerve growth factor and neurotrophins 3 and 4/5 but not by brain-derived neurotrophic factor. *Proc Natl Acad Sci USA* 91: 8607–8611.
- Fitzjohn SM, Morton RA, Kuenzi F, Rosahl TW, Shearman M, Lewis H, et al. 2001. Age-related impairment of synaptic transmission but normal long-term potentiation in transgenic mice that overexpress the human APP695SWE mutant form of amyloid precursor protein. *J Neurosci* 21: 4691–4698.
- Flood JF, Harris FJ, Morley JE. 1996. Age-related changes in hippocampal drug facilitation of memory processing in SAMP8 mice. *Neurobiol Aging* 17: 15–24.
- Foster TC. 1999. Involvement of hippocampal synaptic plasticity in age-related memory decline. *Brain Res Brain Res Rev* 30: 236–249.
- Foster TC, Norris CM. 1997. Age-associated changes in Ca²⁺-dependent processes: relation to hippocampal synaptic plasticity. *Hippocampus* 7: 602–612.
- Frey S, Bergado JA, Seidenbecher T, Pape HC, Frey JU. 2001. Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Hetero-synaptic induction of late-LTP. *J Neurosci* 21: 3697–3603.
- Frey U, Krug M, Reymann KG, Matthies H. 1988. Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Res* 452: 57–65.
- Frey U, Morris RG. 1997. Synaptic tagging and long-term potentiation. *Nature* 385(6616): 533–536.
- Frey U, Morris RG. 1998. Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 21: 181–188.
- Friedman DJ, Duckles SP. 1994. Influence of age on control of norepinephrine release: Ca²⁺ channels and dopamine D2 receptors. *Eur J Pharmacol* 252: 1–9.
- Fujii S, Sumikawa K. 2001. Acute and chronic nicotine exposure reverse age-related declines in the induction of long-term potentiation in the rat

- hippocampus. *Brain Res* 894: 347–353.
- Fukunaga K, Muller D, Miyamoto E. 1996. CaM kinase II in long-term potentiation. *Neurochem Int* 28: 343–358.
- Garrone B, Luparini MR, Tolu L, Landolfi C, Milanese C. 1998. Effect of the subchronic treatment with the acetylcholinesterase inhibitor heptastigmine on central cholinergic transmission and memory impairment in aged rats. *Neurosci Lett* 245: 53–57.
- Gazzaley AH, Thakker MM, Hof PR, Morrison JH. 1997. Preserved number of entorhinal cortex layer II neurons in aged macaque monkeys. *Neurobiol Aging* 18: 549–553.
- Geinisman Y. 1993. Perforated axospinous synapses with multiple, completely partitioned transmission zones: Probable structural intermediates in synaptic plasticity. *Hippocampus* 3: 417–434.
- Geinisman Y, deToledo-Morrell L, Morrell F, Heller RE. 1995. Hippocampal markers of age-related memory dysfunction: behavioral, electrophysiological and morphological perspectives. *Prog Neurobiol* 45: 223–252.
- Geinisman Y, Morrell F, de Toledo-Morrell L. 1989. Perforated synapses on double-headed dendritic spines: a possible structural substrate of synaptic plasticity. *Brain Res* 480: 326–329.
- Geinisman Y, Toledo-Morrell L, Morrell F, Persina IS, Rossi M. 1992. Structural synaptic plasticity associated with the induction of long-term potentiation is preserved in the dentate gyrus of aged rats. *Hippocampus* 2: 445–456.
- Gerlach M, Riederer P. 1996. Animal models of Parkinson's disease: An empirical comparison with the phenomenology of the disease in man. *J Neural Transm* 103: 987–1041.
- Graeber MB, Grasbon-Frodl E, Eitzen UV, Kösel S. 1998. Neurodegeneration and aging: Role of the second genome. *J Neurosci Res* 52: 1–6.
- Harley CW, Sara SJ. 1992. Locus coeruleus bursts induced by glutamate trigger delayed perforant path spike amplitude potentiation in the dentate gyrus. *Exp Brain Res* 89: 581–587.
- Harwood DG, Barker WW, Ownby RL, Duara R. 2000. Relationship of behavioral and psychological symptoms to cognitive impairment and functional status in Alzheimer's disease. *Int J Geriatr Psychiatry* 15: 393–400.
- Huang YY, Kandel ER. 1994. Recruitment of long-lasting and protein kinase A-dependent long-term potentiation in the CA1 region of hippocampus requires repeated tetanization. *Learn Mem* 1: 74–82.
- Ikegami S. 1994. Behavioral impairment in radial-arm maze learning and acetylcholine content of the hippocampus and cerebral cortex in aged mice. *Behav Brain Res* 65: 103–111.
- Ikegami S, Shumiya S, Kawamura H. 1992. Age-related changes in radial-arm maze learning and basal forebrain cholinergic systems in senescence accelerated mice (SAM). *Behav Brain Res* 51: 15–22.
- Ikegaya Y, Saito H, Abe K. 1995. High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 22: 203–207.
- Ikegaya Y, Saito H, Abe K. 1996. The basomedial and basolateral amygdaloid nuclei contribute to the induction of long-term potentiation in the dentate gyrus in vivo. *Eur J Neurosci* 8: 1833–1839.
- Isacson O, Seo H, Lin L, Albeck D, Granholm AC. 2002. Alzheimer's disease and Down's syndrome: roles of APP, trophic factors and ACh. *Trends Neurosci* 25: 79–84.
- Ishiyama J, Saito H, Abe K. 1991. Epidermal growth factor and basic fibroblast growth factor promote the generation of long-term potentiation in the dentate gyrus of anaesthetized rats. *Neurosci Res* 12: 403–411.
- Jas J, Almaguer W, Frey JU, Bergado J. 2000. Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res* 861: 186–189.
- Kadar T, Silbermann M, Brandeis R, Levy A. 1990. Age-related structural changes in the rat hippocampus: correlation with working memory deficiency. *Brain Res* 512: 113–120.
- Kang H, Schuman EM. 1995a. Long-lasting neurotrophin-induced enhancement of synaptic transmission in the adult hippocampus. *Science* 267 (5204): 1658–1662.
- Kang HJ, Schuman EM. 1995b. Neurotrophin-induced modulation of synaptic transmission in the adult hippocampus. *J Physiol Paris* 89: 11–22.
- Kang HJ, Schuman EM. 1996. A requirement for local protein synthesis in neurotrophin-induced hippocampal synaptic plasticity. *Science* 273(5280): 1402–1406.
- Kirkwood A, Silva A, Bear MF. 1997. Age-dependent decrease of synaptic plasticity in the neocortex of

- alphaCaMKII mutant mice. *Proc Natl Acad Sci USA*. 94: 3380–3383.
- Kitchigina V, Vankov A, Harley C, Sara SJ. 1997. Novelty-elicited, noradrenaline-dependent enhancement of excitability in the dentate gyrus. *Eur J Neurosci* 9: 41–47.
- Kito S, Miyoshi R, Nomoto T. 1990. Influence of age on NMDA receptor complex in rat brain studied by in vitro autoradiography. *J Histochem Cytochem* 38: 1725–1731.
- Kovalchuk Y, Hanse E, Kafitz KW, Konnerth A. 2002. Postsynaptic induction of BDNF-mediated long-term potentiation. *Science* 295(5560): 1729–1734.
- Krug M, Lossner B, Ott T. 1984. Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 13: 39–42.
- Lanahan A, Lyford G, Stevenson GS, Worley PF, Barnes CA. 1997. Selective alteration of long-term potentiation-induced transcriptional response in hippocampus of aged, memory-impaired rats. *J Neurosci* 17: 2876–2885.
- Landfield PW, McGaugh JL, Tusa RJ. 1972. Theta rhythm: a temporal correlate of memory storage processes in the rat. *Science* 175: 87–89.
- Landfield PW, Thibault O, Mazzanti ML, Porter NM, Kerr DS. 1992. Mechanisms of neuronal death in brain aging and Alzheimer's disease: Role of endocrine-mediated calcium dyshomeostasis. *J Neurobiol* 23: 1247–1260.
- Langley LK, Madden DJ. 2000. Functional neuroimaging of memory: implications for cognitive aging. *Microsc Res Tech* 51: 75–84.
- Larson J, Lynch G, Games D, Seubert P. 1999. Alterations in synaptic transmission and long-term potentiation in hippocampal slices from young and aged PDAPP mice. *Brain Res* 840: 23–35.
- Leanza G, Martinez-Serrano A, Bjorklund A. 1998. Amelioration of spatial navigation and short-term memory deficits by grafts of foetal basal forebrain tissue placed into the hippocampus and cortex of rats with selective cholinergic lesions. *Eur J Neurosci* 10: 2353–2370.
- Levin ED, Torry D. 1996. Acute and chronic nicotine effects on working memory in aged rats. *Psychopharmacology (Berl)* 123: 88–97.
- Liang F, Lu B. 1992. Changes in excitatory amino acids and N-methyl-D-aspartate receptors in the brain in aged mice. *Zhonghua Yi Xue Za Zhi* 72: 33–35, 64. Chinese.
- Lu B, Chow A. 1999. Neurotrophins and hippocampal synaptic transmission and plasticity. *J Neurosci Res* 58: 76–87.
- Luine V, Bowling D, Hearn M. 1990. Spatial memory deficits in aged rats: contributions of monoaminergic systems. *Brain Res* 537: 271–278.
- Luparini MR, Del Vecchio A, Barillari G, Magnani M, Prosdocimi M. 2000. Cognitive impairment in old rats: a comparison of object displacement, object recognition and water maze. *Aging (Milano)* 12: 264–273.
- Lynch MA. 1998a. Age-related impairment in long-term potentiation in hippocampus: a role for the cytokine, interleukin-1 beta?. *Prog Neurobiol* 56: 571–589.
- Lynch MA. 1998b. Analysis of the mechanisms underlying the age-related impairment in long-term potentiation in the rat. *Rev Neurosci* 1998b. 9: 169–201.
- Lynch MA, Voss KL. 1994. Membrane arachidonic acid concentration correlates with age and induction of long-term potentiation in the dentate gyrus in the rat. *Eur J Neurosci* 6: 1008–1014.
- Magnusson KR. 1998. Aging of glutamate receptors: correlations between binding and spatial memory performance in mice. *Mech Ageing Dev* 104: 227–248.
- Magnusson KR, Cotman CW. 1993. Effects of aging on NMDA and MK801 binding sites in mice. *Brain Res* 604: 334–37.
- Magnusson KR, Nelson SE, Young AB. 2002. Age-related changes in the protein expression of subunits of the NMDA receptor. *Brain Res Mol Brain Res* 99: 40–45.
- Martin SJ, Morris RG. 2002. New life in an old idea: the synaptic plasticity and memory hypothesis revisited. *Hippocampus* 12: 609–636.
- Matthies H. 1998. Neuronale Grundlagen der Gedächtnisbildung. *Enzyklopädie der Psychologie* 15–42.
- Matthies H, Frey U, Reymann K, Krug M, Jork R, Schroeder H. Different mechanisms and multiple stages of LTP. *Adv Exp Med Biol* 1990. 268: 359–368.
- Matthies H, Reymann KG. 1993. Protein kinase A inhibitors prevent the maintenance of hippocampal long-term potentiation. *Neuroreport* 4: 712–714.
- Matthies H, Ruethrich H, Ott T, Matthies HK, Matthies R. 1986. Low frequency perforant path stimulation as a conditioned stimulus demonstrates

- correlations between long-term synaptic potentiation and learning. *Physiol Behav* 36: 811–821.
- McEwen BS. 1994. Corticosteroids and hippocampal plasticity. *Ann NY Acad Sci* 746: 134–142; discussion 142–144, 178–179.
- McGahon BM, Martin DS, Horrobin DF, Lynch MA. 1999. Age-related changes in LTP and antioxidant defenses are reversed by an alpha-lipoic acid-enriched diet. *Neurobiol Aging* 20: 655–664.
- Medina JF, Christopher Repa J, Mauk MD, LeDoux JE. 2002. Parallels between cerebellum- and amygdala-dependent conditioning. *Nat Rev Neurosci* 3: 122–131.
- Merrill DA, Chiba AA, Tuszynski MH. 2001. Conservation of neuronal number and size in the entorhinal cortex of behaviorally characterized aged rats. *J Comp Neurol* 438: 445–456.
- Merrill DA, Roberts JA, Tuszynski MH. 2000. Conservation of neuron number and size in entorhinal cortex layers II, III, and V/VI of aged primates. *J Comp Neurol* 422: 396–401.
- Milner B, Squire LR, Kandel ER. 1998. Cognitive neuroscience and the study of memory. *Neuron* 20: 445–468.
- Mons N, Enderlin V, Jaffard R, Huguieret P. 2001. Selective age-related changes in the PKC-sensitive, calmodulin-binding protein, neurogranin, in the mouse brain. *J Neurochem* 79: 859–867.
- Moore CI, Browning MD, Rose GM. 1993. Hippocampal plasticity induced by primed burst, but not long-term potentiation, stimulation is impaired in area CA1 of aged Fischer 344 rats. *Hippocampus* 3: 57–66.
- Morrison JH, Hof PR. 1997. Life and death of neurons in the aging brain. *Science* 278(5337): 412–419.
- Mullany P, Lynch MA. 1997. Changes in protein synthesis and synthesis of the synaptic vesicle protein, synaptophysin, in entorhinal cortex following induction of long-term potentiation in dentate gyrus: an age-related study in the rat. *Neuropharmacology* 36: 973–980.
- Murray CA, Lynch MA. 1998. Dietary supplementation with vitamin E reverses the age-related deficit in long term potentiation in dentate gyrus. *J Biol Chem* 273: 12161–12168.
- Nalbantoglu J, Tirado-Santiago G, Lahsaïni A, Poirier J, Goncalves O, Verge A, et al. 1997. Impaired learning and LTP in mice expressing the carboxy terminus of the Alzheimer amyloid precursor protein. *Nature* 387(6632): 500–505.
- Nguyen PV, Kandel ER. 1996. A macromolecular synthesis-dependent late phase of long-term potentiation requiring cAMP in the medial perforant pathway of rat hippocampal slices. *J Neurosci* 16: 3189–3198.
- Nguyen PV, Kandel ER. 1997. Brief theta-burst stimulation induces a transcription-dependent late phase of LTP requiring cAMP in area CA1 of the mouse hippocampus. *Learn Mem* 4: 230–243.
- Norris CM, Halpain S, Foster TC. 1998. Reversal of age-related alterations in synaptic plasticity by blockade of L-type Ca²⁺ channels. *J Neurosci* 18: 3171–3179.
- Norris CM, Korol DL, Foster TC. 1996. Increased susceptibility to induction of long-term depression and long-term potentiation reversal during aging. *J Neurosci* 16: 5382–5392.
- Okuma Y, Murayama T, Tha KK, Yamada C, Hosokawa M, Ishikawa A, et al. 2000. Learning deficiency and alterations in acetylcholine receptors and protein kinase C in the brain of senescence-accelerated mouse (SAM)-P10. *Mech Ageing Dev* 114: 191–199.
- Orr G, Rao G, Houston FP, McNaughton BL, Barnes CA. Hippocampal synaptic plasticity is modulated by theta rhythm in the fascia dentata of adult and aged freely behaving rats. *Hippocampus* 2001. 11: 647–654.
- Otani S, Abraham WC. 1989. Inhibition of protein synthesis in the dentate gyrus, but not the entorhinal cortex, blocks maintenance of long-term potentiation in rats. *Neurosci Lett* 106: 175–180.
- Palmer AM. 2002. Pharmacotherapy for Alzheimer's disease: progress and prospects. *Trends Pharmacol Sci* 23: 426–433.
- Pedigo NW Jr. 1994. Neurotransmitter receptor plasticity in aging. *Life Sci* 55: 1985–1991.
- Pontzer NJ, Chandler LJ, Stevens BR, Crews FT. 1990. Receptors, phosphoinositol hydrolysis and plasticity of nerve cells. *Prog Brain Res* 86: 221–225.
- Powers RE, Struble RG, Casanova MF, O'Connor DT, Kitt CA, Price DL. 1988. Innervation of human hippocampus by noradrenergic systems: normal anatomy and structural abnormalities in aging and in Alzheimer's disease. *Neuroscience* 25: 401–417.
- Quirk GJ, Armony JL, Repa JC, Li XF, LeDoux JE.

1996. Emotional memory: A search for sites of plasticity. *Cold Spring Harbor Symp Quant Biol* 61: 247–257.
- Rapp PR, Gallagher M. 1996. Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. *Proc Natl Acad Sci USA* 93: 9926–9930.
- Reymann KG, Brödemann R, Kase H, Matthies H. 1988. Inhibitors of calmodulin and protein kinase C block different phases of hippocampal long-term potentiation. *Brain Res* 461: 388–392.
- Reymann KG, Frey U, Jork R, Matthies H. 1988. Polymyxin B, an inhibitor of protein kinase C, prevents the maintenance of synaptic long-term potentiation in hippocampal CA1 neurons. *Brain Res* 440: 305–314.
- Rioult-Pedotti MS, Friedman D, Hess G, Donoghue JP. 1998. Strengthening of horizontal cortical connections following skill learning. *Nat Neurosci* 1: 230–234.
- Roberts LA, Higgins MJ, O'Shaughnessy CT, Stone TW, Morris BJ. 1996. Changes in hippocampal gene expression associated with the induction of long-term potentiation. *Mol Brain Res* 42: 123–127.
- Robinson GB, Racine RJ. 1982. Heterosynaptic interactions between septal and entorhinal inputs to the dentate gyrus: long-term potentiation effects. *Brain Res* 249: 162–166.
- Robinson GB, Racine RJ. 1986. Interactions between septal and entorhinal inputs to the rat dentate gyrus facilitation effects. *Brain Res* 379: 63–67.
- Rogan MT, Staubli UV, LeDoux JE. 1997. Fear conditioning induces associative long-term potentiation in the amygdala. *Nature* 390: 604–607.
- Ronn LC, Berezin V, Bock E. 2000. The neural cell adhesion molecule in synaptic plasticity and ageing. *Int J Dev Neurosci* 18: 193–199.
- Rosenzweig ES, Rao G, McNaughton BL, Barnes CA. 1997. Role of temporal summation in age-related long-term potentiation-induction deficits. *Hippocampus* 7: 549–558.
- Rosenzweig MR. 1996. Aspects of the search for neural mechanisms of memory. *Annu Rev Psychol* 47: 1–32.
- Russell RW. 1996. Continuing the search for cholinergic factors in cognitive dysfunction. *Life Sci* 58: 1965–1970.
- Sapolsky RM. 1999. Glucocorticoids, stress, and their adverse neurological effects: relevance to aging. *Exp Gerontol* 34: 721–732.
- Scali C, Casamenti F, Pazzagli M, Bartolini L, Pepeu G. 1994. Nerve growth factor increases extracellular acetylcholine levels in the parietal cortex and hippocampus of aged rats and restores object recognition. *Neurosci Lett* 170: 117–120.
- Seabrook GR, Smith DW, Bowery BJ, Easter A, Reynolds T, Fitzjohn SM, et al. 1999. Mechanisms contributing to the deficits in hippocampal synaptic plasticity in mice lacking amyloid precursor protein. *Neuropharmacology* 38: 349–359.
- Segal M, Auerbach JM. 1997. Muscarinic receptors involved in hippocampal plasticity. *Life Sci* 60: 1085–1091.
- Seidenbecher T, Balschun D, Reymann KG. 1995. Drinking after water deprivation prolongs "unsaturated" LTP in the dentate gyrus of rats. *Physiol Behav* 57: 1001–1004.
- Seidenbecher T, Reymann KG, Balschun D. 1997. A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc Natl Acad Sci USA* 94: 1494–1499.
- Shankar S, Teyler TJ, Robbins N. 1998. Aging differentially alters forms of long-term potentiation in rat hippocampal area CA1. *J Neurophysiol* 79: 334–341.
- Shapiro M. 2001. Plasticity, hippocampal place cells, and cognitive maps. *Arch Neurol* 58: 874–881.
- Shen J, Barnes CA, Wenk GL, McNaughton BL. 1996. Differential effects of selective immunotoxic lesions of medial septal cholinergic cells on spatial working and reference memory. *Behav Neurosci* 110: 1181–1186.
- Sherman KA, Friedman E. 1990. Pre- and post-synaptic cholinergic dysfunction in aged rodent brain regions: new findings and an interpretative review. *Int J Dev Neurosci* 8: 689–708.
- Silva AJ, Stevens CF, Tonegawa S, Wang Y. 1992. Deficient hippocampal long-term potentiation in alpha-calcium-calmodulin kinase II mutant mice. *Science* 257(5067): 201–206.
- Sirvio J. 1999. Strategies that support declining cholinergic neurotransmission in Alzheimer's disease patients. *Gerontology* 45 Suppl 1: 3–14.
- Smith DE, Roberts J, Gage FH, Tuszynski MH. 1999. Age-associated neuronal atrophy occurs in the primate brain and is reversible by growth factor gene therapy. *Proc Natl Acad Sci USA* 96: 10893–10898.

- Smith TD, Gallagher M, Leslie FM. 1995. Cholinergic binding sites in rat brain: Analysis by age and cognitive status. *Neurobiol Aging* 16: 161–173.
- Soininen HS, Partanen K, Pitkänen A, Vainio P, Hänninen T, Hallikainen M et al. 1994. Volumetric MRI analysis of the amygdala and the hippocampus in subjects with age-associated memory impairment: correlation to visual and verbal memory. *Neurology* 44: 1660–1668.
- Stanton PK, Sarvey JM. 1985. Blockade of norepinephrine-induced long-lasting potentiation in the hippocampal dentate gyrus by an inhibitor of protein synthesis. *Brain Res* 361: 276–283.
- Stanton PK, Sarvey JM. 1987. Norepinephrine regulates long-term potentiation of both the population spike and dendritic EPSP in hippocampal dentate gyrus. *Brain Res Bull* 18: 115–119.
- Stemmelin J, Lazarus C, Cassel S, Kelche C, Cassel J. 2000. Immunohistochemical and neurochemical correlates of learning deficits in aged rats. *Neuroscience* 96: 275–289.
- Stephan A, Davis S, Salin H, Dumas S, Mallet J, Laroche S. 2002. Age-dependent differential regulation of genes encoding APP and alpha-synuclein in hippocampal synaptic plasticity. *Hippocampus* 12: 55–62.
- Stevens CF, Tonegawa S, Wang Y. 1994. The role of calcium-calmodulin kinase II in three forms of synaptic plasticity. *Curr Biol* 4: 687–693.
- Stratton KR, Baraban JM, Worley PF. 1988. Norepinephrine stimulation of adenylate cyclase potentiates protein kinase C action: electrophysiological studies in the dentate gyrus. *Synapse* 2: 614–618.
- Sykova E. 2001. Glial diffusion barriers during aging and pathological states. *Prog Brain Res* 132:339–363.
- Tamaru M, Yoneda Y, Ogita K, Shimizu J, Nagata Y. 1991. Age-related decreases of the N-methyl-D-aspartate receptor complex in the rat cerebral cortex and hippocampus. *Brain Res* 542: 83–90.
- Terlau H, Seifert W. 1990. Fibroblast growth factor enhances long-term potentiation in the hippocampal slice. *Eur J Neurosci* 2: 973–977.
- Teyler TJ, Cavus I, Coussens C, DiScenna P, Grover L, Lee YP, Little Z. 1994. Multideterminant role of calcium in hippocampal synaptic plasticity. *Hippocampus* 4: 623–634.
- Teyler TJ, Perkins AT, Harris KM. 1989. The development of long-term potentiation in hippocampus and neocortex. *Neuropsychologia* 27: 31–39.
- Vannucchi MG, Scali C, Kopf SR, Pepeu G, Casamenti F. 1997. Selective muscarinic antagonists differentially affect in vivo acetylcholine release and memory performances of young and aged rats. *Neuroscience* 79: 837–846.
- Ward MT, Oler JA, Markus EJ. 1999. Hippocampal dysfunction during aging I: deficits in memory consolidation. *Neurobiol Aging* 20: 363–372.
- Ward MT, Stoelzel CR, Markus EJ. 1999. Hippocampal dysfunction during aging II: deficits on the radial-arm maze. *Neurobiol Aging* 20: 373–380.
- Wasling P, Hanse E, Gustafsson B. 2002. Long-term depression in the developing hippocampus: low induction threshold and synapse nonspecificity. *J Neurosci* 22: 1823–1830.
- Wenk GL, Barnes CA. 2000. Regional changes in the hippocampal density of AMPA and NMDA receptors across the lifespan of the rat. *Brain Res* 885: 1–5.
- Wenk GL, Walker LC, Price DL, Cork LC. 1991. Loss of NMDA, but not GABA-A, binding in the brains of aged rats and monkeys. *Neurobiol Aging* 12: 93–98.
- West MJ. 1993. Regionally specific loss of neurons in the aging human hippocampus. *Neurobiol Aging* 14: 287–293.
- Yuste R, Bonhoeffer T. 2001. Morphological changes in dendritic spines associated with long-term synaptic plasticity. *Annu Rev Neurosci* 24:1071–1089.

RESULTADOS

Los resultados que conforman esta tesis están recogidos en una serie de artículos científicos que aparecen a continuación.

A la copia de cada artículo antecede una breve descripción del contenido fundamental del mismo que facilitará al lector seguir el orden de pensamiento que guió este trabajo. Con ese mismo propósito los artículos aparecen en orden lógico y no cronológico ya que algunos, por imperativos materiales y de oportunidad, tuvieron que ser realizados antes y otros, por razones de igual índole, tuvieron que ser diferidos.

Capítulo 1: El reforzamiento conductual de la potenciación sináptica duradera es dependiente de la síntesis de proteínas

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Los procesos de plasticidad sináptica, incluida la LTP, dependen del metabolismo macromolecular. La activación de los procesos de síntesis de proteínas que se requieren para la consolidación del fenómeno neuroplástico puede ser provocada por el mismo agente inductor, o por algún evento modulador adicional.

En el modelo de reforzamiento conductual que empleamos, la tetanización de la vía perforante (inductor) no parece tener intensidad suficiente como para activar los mecanismos de síntesis de proteínas. De acuerdo con esa concepción formulamos la hipótesis que dio origen a este trabajo:

El reforzamiento conductual actúa como evento modulador que, por vías posiblemente heterosinápticas, activa la maquinaria biosintética celular y proporciona a las sinapsis potenciadas las proteínas necesarias para consolidar (prolongar) el estado de potenciación.

De aquí puede derivarse fácilmente la hipótesis que se explora directamente en este trabajo: Si el reforzamiento conductual actúa como evento modulador activador de la síntesis de proteínas, el bloqueo de la síntesis de proteínas debe abolir el efecto reforzador del evento conductual.

Para comprobar esta hipótesis aplicamos el modelo de reforzamiento conductual a un grupo de animales a los que se inyectó intraventricularmente un bloqueador de la síntesis de proteínas (anisomicina) una hora antes de inducir la LTP y permitir el acceso al agua

(reforzamiento conductual). Un grupo similar de animales, también privado de agua durante 24 horas, al que se inyectó solución salina sirvió como control.

El resultado fundamental fue que el bloqueo de la síntesis de proteínas abolió totalmente el efecto de reforzamiento conductual, como muestra la figura 1b, donde se comparan ambos grupos.

Este resultado confirma la hipótesis planteada y permite asumir que el reforzamiento conductual actúa como evento modulador de la plasticidad sináptica mediante la activación de los mecanismos celulares de biosíntesis que aportan las proteínas requeridas por el proceso neuroplástico para su consolidación.

Behavioral reinforcement of long-term potentiation in rat dentate gyrus in vivo is protein synthesis-dependent

Jorge A. Bergado^a, W. Almaguer-Melian^a, Sergiy Kostenko^b, Sabine Frey^b, Julietta U. Frey^{b,*}

^aInternational Center for Neurological Restoration, Avenue 25 # 15805, Playa, Havana, Cuba

^bLeibniz-Institute for Neurobiology, Brennekestrasse 6, 39118 Magdeburg, Germany

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Abstract

A transient, protein synthesis-independent long-term potentiation (early-LTP, <4 h) can be reinforced into a maintained protein synthesis-dependent late-LTP (>4 h) by specific electrical stimulation of limbic structures (J. Neurosci. 21 (2001) 3697). Similarly, LTP-modulation can be obtained by behavioral stimuli with strong motivational content. However, the requirement of protein synthesis during behavioral reinforcement has not been shown so far. Thus, we have studied here this specific question using a behavioral reinforcement protocol, i.e. allowing water-deprived animals to drink 15 min after induction of early-LTP. This procedure transformed early-LTP into late-LTP. Anisomycin, a reversible protein synthesis inhibitor, abolished behavioral LTP-reinforcement. These results demonstrate that behavioral reinforcement depends on protein synthesis.

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Keywords: Behavioral reinforcement; Long-term potentiation; Late-long-term potentiation; Protein synthesis; Anisomycin; Dentate gyrus; Hippocampus

Long-term potentiation (LTP) is a long-lasting increase of synaptic efficacy which is considered to be a cellular correlate of memory formation [15]. LTP can be separated in at least two temporally distinct phases by the use of protein synthesis inhibitors during its induction: an early phase (early-LTP) lasting less than 4 h and a protein synthesis-dependent late-LTP [8,13]. Early-LTP in the dentate gyrus (DG), induced by a weak tetanus to the perforant pathway (PP), can be converted into late-LTP by heterosynaptic, modulating signals arising from other limbic regions like the basolateral amygdala (BLA) [6] or the septum [5]. Both effects are protein synthesis-dependent as they are abolished by the intraventricular administration of anisomycin. A reinforcement of LTP can also be observed when animals receive a behavioral stimulus with a strong motivational content, an effect we called behavioral reinforcement (BR) [19]. It remains unclear whether BR depends on protein synthesis as well. BR and amygdala-induced reinforcement have similar effective time windows

and we have recently suggested that the BLA is an important structure of the neural system involved in BR [2]. Our findings led to the hypothesis that processes of BR on plastic events, such as prolonging LTP, require the synthesis of proteins in general. We have now tested this hypothesis. Male adult WISTAR rats (obtained from CENPALAB and weighing 250 g during electrode implantation) were housed individually after surgery in plastic, translucent cages. Water and food (Rat Chow, Cenpalab, Cuba) remained ad libitum throughout the experiment except for the series investigating the effect of BR of LTP in rats which were water-deprived for 24 h. Animals were handled according to the German Regulations for Using Laboratory Animals.

Surgery was performed under chloral hydrate narcosis (420 mg/kg, intraperitoneally) according to the procedure described elsewhere [1]. Electrodes for recording were implanted in the hilus of DG and for stimulation in the angular bundle of the PP. A guide cannulae was implanted to reach the right lateral ventricle [1]. One week after surgery the animals were habituated to the test chamber for at least 4 h before obtaining an input–output curve on which the stimulus intensity was determined for each animal (40% maximal population spike). Animals were then randomly

* Corresponding author. Leibniz-Institute for Neurobiology, Department of Neurophysiology, Brennekestrasse 6, 39118 Magdeburg, Germany. Tel.: +49-391-6263-422; fax: +49-391-6263-421.

E-mail address: frey@ifn-magdeburg.de (J.U. Frey).

assigned to one of the experimental groups. In the control-LTP group a weak tetanus (3×15 impulses at 200 Hz) was used to induce early-LTP (Fig. 1a, open circles). The BR group was water-deprived for 24 h before inducing early-LTP by the above protocol; 15 min later water was made available until the end of the recording session (Fig. 1a, filled circles). The same protocol was followed using a NaCl-treated BR group (NaCl-BR; NaCl used as the vehicle; Fig. 1b, open circles) and an anisomycin (ANI)-treated BR group (ANI-BR; Fig. 1b, filled circles). In the latter series $5 \mu\text{l}$ of the saline or anisomycin solution (0.905 mol , Sigma, St. Louis, MO) was injected, respectively, into the lateral ventricle (i.c.v.) 1 h before induction of the early-LTP and subsequent BR. Control experiments performed earlier in our laboratory revealed no effects of water deprivation on test potentials recorded in a control group

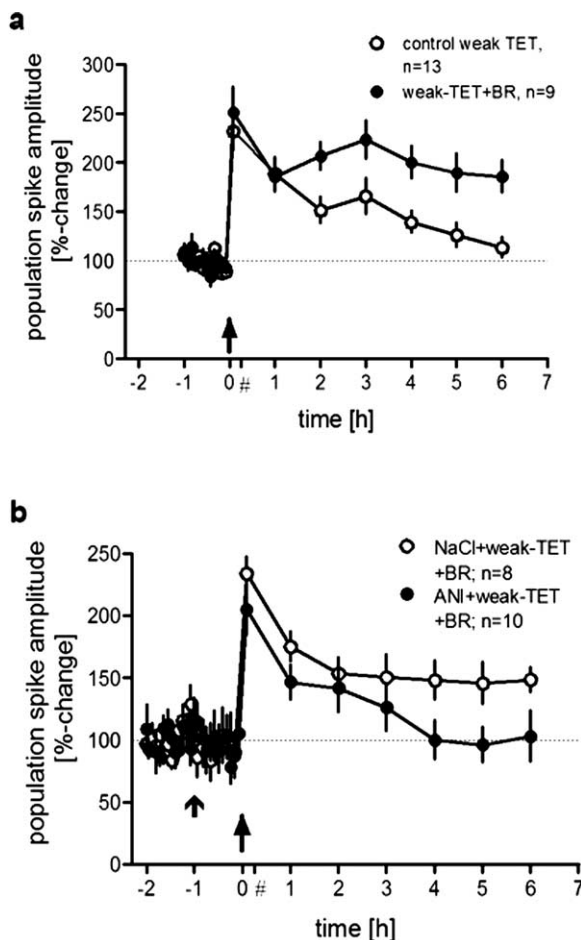


Fig. 1. The role of protein synthesis during behaviorally reinforced LTP. (a) Weak tetanization (strong arrow) resulted in a transient protein synthesis-independent early-LTP (open circles). Early-LTP was transferred into late-LTP if water-deprived animals were allowed to drink water 15 min (double cross) after tetanization (filled circles). (b) Early-LTP can also be reinforced into late-LTP by water access in water-deprived animals which received NaCl i.c.v. 1 h before tetanization (small arrow) and when drinking was allowed 15 min after tetanization (open circles). This reinforcement of early-LTP beyond 4 h was blocked when the reversible protein synthesis inhibitor anisomycin was applied i.c.v. instead of NaCl before tetanization (filled circles). Statistics: see text.

without tetanization [19]. Test recordings were made every 15 min after early-LTP induction. Each data-point presented in Fig. 1 represents an averaged value of five consecutive potentials recorded at a frequency of 0.1 Hz. All animals involved in BR protocols, even those receiving anisomycin, drank abundantly within 1 min after water was made available. No group differences were found in the total volume of water consumed at the end of the recording period (BR-control group: $15.8 \pm 3.81 \text{ ml}$, NaCl-weak-TET-BR control group: $16.5 \pm 3.53 \text{ ml}$, ANI-weak-TET-BR group: $17.0 \pm 4.84 \text{ ml}$; the values represent the mean volume of water consumption $\pm$ standard deviation). After finishing the experiments the animals were perfused under narcosis and the correct location of the cannulae as well as electrodes was proven histologically.

Administration of anisomycin showed no effect on the population spike amplitude (PSA, Fig. 1b: baseline recordings, filled circles) during control baseline recordings taken before the induction of LTP. The two-way ANOVA showed no treatment ($F(1,21) = 0.125418$, $P > 0.05$) or time ($F(23,483) = 0.736305$, $P > 0.05$; data not shown) effect.

Tetanic stimulation resulted in a significant increase of the PSA for a duration of 4 h, when the data of the control LTP group were compared with its own pre-tetanus baseline values (Fig. 1a, open circles, control early-LTP; Student's *t*-test for paired samples). Other groups (BR, NaCl-BR and ANI-BR) showed a similar initial level of potentiation measured 5 min after tetanization (no differences detected by using the one-way ANOVA: $F(3,39) = 0.776967$, $P > 0.05$). However, the time course of LTP differed significantly among the latter two groups. The two-way ANOVA with treatment and time (repeated measures) as factors showed significant differences for both factors and their interaction (treatment, $F(3,39) = 3.15204$; time, $F(5,195) = 15.83220$; interaction, $F(15,195) = 3.14201$; $P < 0.05$ in every case). A post-hoc analysis (Duncan's test) proved statistically significant differences between the control BR-LTP group and the ANI-BR group (Fig. 1b, filled circles), respectively, but not the NaCl-BR group (Fig. 1, open circles). Further analysis of the NaCl-BR and ANI-BR groups (Duncan's test) revealed significant differences between groups only at later phases, i.e. after 4 h, corresponding to late-LTP (two-way ANOVA, $F(1,17) = 6.028451$, $P < 0.05$).

Our results demonstrate that also BR of LTP depends on protein synthesis. The requirement of protein synthesis has been consistently reported for long-term memory formation in several species and brain structures [3,11,18]. Also processes of functional plasticity like LTP [8,13,16], LTD [14,17] and other forms [4,10,12] are characterized by a similar requirement: a preserved induction but an impaired maintenance if protein synthesis is blocked during initial phases. Although one can easily conceive a general role for proteins in long-term plasticity mechanisms the identification of specific proteins involved has been shown to be difficult, perhaps because of the big number of proteins

synthesized in apparent correlation with processes of functional plasticity. Two related problems refer to the mechanisms of induction and synapse-specific distribution of newly synthesized plasticity-related proteins. Data from our group and others suggest that modulatory, non-glutamatergic transmitters like norepinephrine, dopamine and acetylcholine mediate the reinforcing effect of heterosynaptic afferents during LTP [5–7] by activating intracellular cascades resulting in the regulation of the synthesis of plasticity-related proteins within the neuron [7]. Regarding their distribution, the synaptic tagging hypothesis [9] offers a plausible explanation to maintain synapse-specificity during protein synthesis-dependent events. According to the hypothesis of synaptic tagging our results of protein synthesis-dependent BR of early- into late-LTP can be explained as follows: a weak tetanus used to induce early-LTP may tag specifically the activated synapse population. Modulatory, behaviorally reinforcing inputs which were activated by drinking 15 min after tetanization led to synapse non-specific synthesis of plasticity-related proteins. These proteins were captured by synapses with a tag set, thus transforming early- into late-LTP by activation of the rewarding system, probably involving the amygdala. When protein synthesis was blocked at the time of water presentation, the reinforcing effect was abolished. Our data strongly suggest the physiological significance of heterosynaptic reinforcement processes of plastic events in general.

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References

- [1] W. Almaguer, B. Estupiñán, J.U. Frey, J.A. Bergado, Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP, *Neurobiol. Aging* 23 (2002) 319–324.
- [2] W. Almaguer-Melian, L. Martinez-Marti, J.U. Frey, J.A. Bergado, The amygdala is part of the behavioural reinforcement system modulating long-term potentiation in rat hippocampus, *Neuroscience* 119 (2) (2003) 319–322.
- [3] V.F. Castellucci, W.N. Frost, P. Golet, P.G. Montarolo, S. Schacher, J.A. Morgan, H. Blumenfeld, E.R. Kandel, Cell and molecular analysis of long-term sensitization in *Aplysia*, *J. Physiol. (Paris)* 81 (1986) 349–357.
- [4] N. del-Olmo, A. Handler, L. Alvarez, J. Bustamante, d.R. Martin, J.M. Solis, Taurine-induced synaptic potentiation and the late phase of long-term potentiation are related mechanistically, *Neuropharmacology* 44 (2003) 26–39.
- [5] S. Frey, J.A. Bergado, J.U. Frey, Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum, *Neuroscience* 118 (2003) 1055–1062.
- [6] S. Frey, J.A. Bergado, T. Seidenbecher, H.C. Pape, J.U. Frey, Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: heterosynaptic induction of late-LTP, *J. Neurosci.* 21 (2001) 3697–3703.
- [7] U. Frey, Y.-Y. Huang, E.R. Kandel, Effects of cAMP simulate a late stage of LTP in hippocampal CA1 neurons, *Science* 260 (1993) 1661–1664.
- [8] U. Frey, M. Krug, K.G. Reymann, H. Matthies, Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro, *Brain Res.* 452 (1988) 57–65.
- [9] U. Frey, R.G.M. Morris, Synaptic tagging and long-term potentiation, *Nature* 385 (1997) 533–536.
- [10] W.A. Gottschalk, H. Jiang, N. Tartaglia, L. Feng, A. Figurov, B. Lu, Signaling mechanisms mediating BDNF modulation of synaptic plasticity in the hippocampus, *Learn. Mem.* 6 (1999) 243–256.
- [11] G. Grecksch, H. Matthies, Two sensitive periods for the amnesic effect of anisomycin, *Pharmacol. Biochem. Behav.* 12 (1980) 663–665.
- [12] L.S. Jones, S.Y. Grooms, D.M. Lapadula, D.V. Lewis, Protein synthesis inhibition blocks maintenance but not induction of epileptogenesis in hippocampal slice, *Brain Res.* 599 (1992) 338–344.
- [13] M. Krug, B. Lössner, T. Ott, Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats, *Brain Res. Bull.* 13 (1984) 39–42.
- [14] D.J. Linden, A protein synthesis-dependent late phase of cerebellar long-term depression, *Neuron* 17 (1996) 483–490.
- [15] H. Matthies, Neurobiological aspects of learning and memory, *Annu. Rev. Psychol.* 40 (1989) 381–404.
- [16] S. Otani, W.C. Abraham, Inhibition of protein synthesis in the dentate gyrus, but not the entorhinal cortex, blocks maintenance of long-term potentiation in rats, *Neurosci. Lett.* 106 (1989) 175–180.
- [17] S. Sajikumar, J.U. Frey, Anisomycin inhibits the late maintenance of long-term depression in rat hippocampal slices in vitro, *Neurosci. Lett.* 338 (2003) 147–150.
- [18] G.E. Schafe, N.V. Nadel, G.M. Sullivan, A. Harris, J.E. LeDoux, Memory consolidation for contextual and auditory fear conditioning is dependent on protein synthesis, PKA, and MAP kinase, *Learn. Mem.* 6 (1999) 97–110.
- [19] T. Seidenbecher, K.G. Reymann, D. Balschun, A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli, *Proc. Natl. Acad. Sci. USA* 94 (1997) 1494–1499.

Capítulo 2: La amígdala es parte del sistema de reforzamiento conductual que modula la plasticidad sináptica duradera

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Además del problema del mecanismo celular implicado en el efecto de reforzamiento del proceso neuroplástico por factores afectivos, es necesario conocer cuáles son las estructuras nerviosas que median dicho efecto.

Conocido el papel de la amígdala como elemento central del Sistema Límbico en la determinación del estado afectivo, así como su acción reforzadora de procesos de memoria, resulta obvio suponer que:

La amígdala es parte del sistema neural implicado en el reforzamiento de procesos neuroplásticos por factores afectivos.

Para evaluar esta hipótesis empleamos nuevamente el modelo de reforzamiento conductual de la LTP en ratas. La participación de la amígdala en el fenómeno de reforzamiento conductual fue evaluado comparando los resultados de grupos de animales en los que se evaluó el reforzamiento conductual en condiciones de bloqueo temporal de (por inyección tópica de un anestésico local) o permanente de la amígdala (por lesión electrolítica). En ambos casos, el resultado fue similar, la inactivación temporal (fig. 1b) o permanente (fig. 2a) de la amígdala bloquea completamente el efecto reforzador del estímulo conductual aplicado después de la inducción de la LTP.

Este resultado confirma la hipótesis planteada y demuestra que la amígdala es parte del sistema neural implicado en el reforzamiento de procesos neuroplásticos por factores afectivos.

Lo anterior no significa, por supuesto, que otras estructuras cerebrales no tengan un papel en estos procesos. Más que un enunciado de cautela, tal reconocimiento es

particularmente importante en este caso, considerando que no existen proyecciones anatómicas directas desde la región bloqueada de la amígdala (amígdala basolateral), hasta la región donde se indujo y estudió el fenómeno de LTP (giro dentado) lo que necesariamente implica la participación de otros mediadores neurales.

LETTER TO NEUROSCIENCE

THE AMYGDALA IS PART OF THE BEHAVIOURAL REINFORCEMENT SYSTEM MODULATING LONG-TERM POTENTIATION IN RAT HIPPOCAMPUS

W. ALMAGUER-MELIAN,^a L. MARTÍNEZ-MARTÍ,^a
J. U. FREY^b AND J. A. BERGADO^{a*}

^aInternational Center for Neurological Restoration (CIREN), Avenue 25 Number 15805, Playa 11300, Havana, Cuba

^bLeibniz-Institute for Neurobiology, Brennekestrasse 6, PF 1860, D-39008 Magdeburg, Germany

Abstract—Long-term potentiation (LTP) in the dentate gyrus can be modulated and prolonged by emotional/motivational influences when concurrently activated. A similar effect on LTP can be obtained by stimulating the amygdala, suggesting that this limbic structure might be part of the neural system involved in behavioural reinforcement. To confirm this we have performed a series of experiments in which the basolateral amygdala was either temporary inactivated by injection of lidocaine or permanently lesioned electrolytically. Both manipulations completely blocked the reinforcing effect of a motivational stimulus (drinking after 24-h deprivation) on LTP at the perforant pathway-dentate gyrus synapses, whilst leaving intact the non-reinforced potentiation. These results demonstrate that the basolateral amygdala is a key structure within the system involved in the modulatory interaction between the affective status of the animal and the mechanisms of functional plasticity. © 2003 IBRO. Published by Elsevier Science Ltd. All rights reserved.

Key words: lidocaine, electrolytic lesions, synaptic plasticity, emotion, motivation, memory.

Long-term potentiation (LTP) has been considered as a cellular mechanism of learning and memory (Matthies, 1989, 1998). It is well known that learning and memory are influenced by the emotional/motivational status of the animal. A similar dependence has been recently demonstrated on LTP (Seidenbecher et al., 1997). Behavioural stimuli with a strong motivational content (behavioural reinforcement), are able to prolong a short-lasting LTP (2–4 h) into a long-lasting LTP (with a duration beyond 4 h). We have obtained coincident results by stimulating the basolateral amygdala (BLA) shortly before or after the induction of protein synthesis-independent early-LTP (E-LTP) (Frey et al., 2001). We were able to show that this E-LTP can be

transformed into a protein synthesis-dependent late-LTP (L-LTP). This suggests that the BLA might be part of the neural system involved in behavioural reinforcement of LTP.

To prove this hypothesis we have performed a series of experiments based on the behavioural reinforcement protocol described by Seidenbecher and co-workers (Seidenbecher et al., 1997) combined with the temporal or permanent inactivation of the BLA by lidocaine injection or electrolytic lesion, respectively.

EXPERIMENTAL PROCEDURES

Male Sprague–Dawley rats, weighing 250–300 g at the time of surgery, were prepared stereotactically and implanted with a recording electrode at the dentate gyrus and a stimulating electrode at the perforant pathway (PP) to obtain monosynaptically evoked potentials. A guide cannula was also implanted in the BLA. Some animals received no cannulae; instead a monopolar electrode was lowered to the BLA in order to lesion this structure by passing 2.5-mA cathodal current during 10 s. All implant or lesion procedures were carried out during the same surgical session. Two weeks after surgery the animals were habituated to the recording chamber for at least 4 h and assigned randomly to the different experimental protocols. The electrophysiological studies included the induction of E-LTP using a weak tetanus (3×15 impulses at 200 Hz). Each impulse was a 0.1-ms square pulse adjusted in intensity to produce a population spike about 40% of its maximal value. In one series 1 μ l of a lidocaine solution (2%) was injected into the BLA within 1 min to temporarily inactivate this nucleus. Lidocaine, or saline, was applied to the BLA 5 min after the tetanus. Behavioural stimulation, which lead to behavioural reinforcement of E-LTP under control conditions (Fig. 1a, closed circles), was applied 15 min after E-LTP induction and consisted of access to water after a 24-h deprivation period. All deprived animals drank abundantly, starting 1–2 min after water was made available. There were no noticeable differences in the volume consumed or the number of drinking bouts during the first 10 min. Water remained *ad libitum* during the rest of the experiment.

Efforts were made to minimise pain or discomfort of the animals according to the Cuban Regulations for the Use of Laboratory Animals. At the end of the experiments the animals were perfused with formalin under chloral hydrate narcosis and the brains processed histologically to confirm the location of the cannulae or the success of electrolytic lesion, respectively. Only data from animals with a correct cannulae location or lesion were processed. The lesion of the BLA often compromised also the lateral or the central region of the amygdala.

Changes in the population spike amplitude (PSA) were measured and statistically evaluated. The relative value to baseline (% PSA) was calculated to each averaged recording of

*Corresponding author. Tel: +53-7-271-5379; fax: +53-7-33-2420.

E-mail address: bergado@neubas.sld.cu (J. A. Bergado).

Abbreviations: ANCOVA, analysis of covariance; BLA, basolateral amygdala; E-LTP, early-long-term potentiation; EPSPs, slope of the excitatory postsynaptic potential; LTP, long-term potentiation; L-LTP, late-long-term potentiation; PP, perforant pathway; PSA, population spike amplitude.

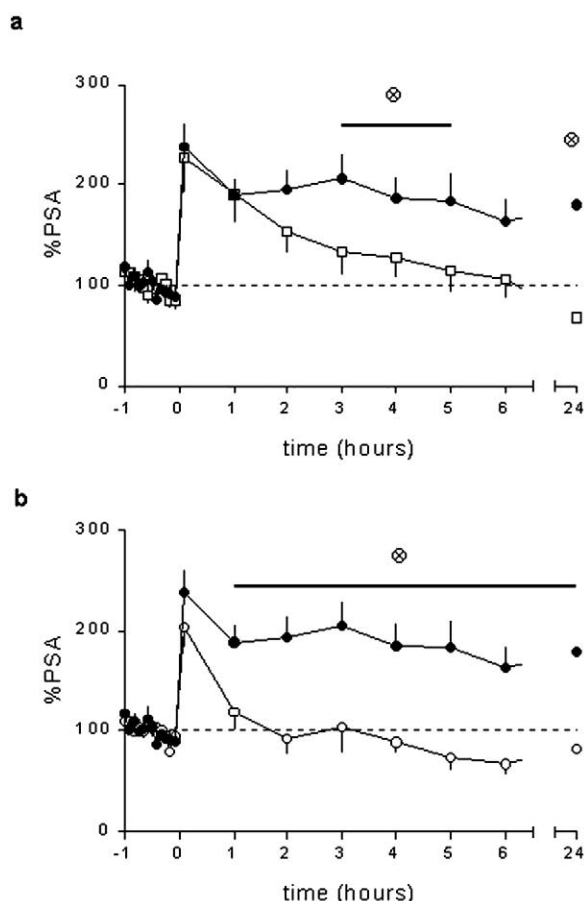


Fig. 1. (a) The weak tetanus used (3×15 impulses at 200 Hz) induced early-long-term potentiation (E-LTP) (control LTP, open squares, $n=8$) lasting about 2 h. When this stimulus is combined with access to water in deprived animals (LTP and behavioural stimulation, filled circles in a and b, $n=14$) LTP is prolonged up to the end of the follow-up period (Wilcoxon test). Statistically significant differences between curves were found at the time points under the symbol ($\otimes$), U test, $P<0.05$). (b) Injection of lidocaine to the basolateral amygdala (LTP induction, lidocaine and behavioural stimulation, open circles, $n=8$) reduces the time of the E-LTP to less than 1 h (Wilcoxon test) and abolishes the behavioural reinforcement effect. Symbol ($\otimes$) as in (a).

five consecutive potentials ($f=0.1$ Hz). The slope of the field excitatory postsynaptic potential (EPSP) showed a similar trend though the changes are smaller in value. The recording electrodes were positioned in the hilus, i.e. a location far away from EPSP generation; thus the PSA was normally used for analysis. Non-parametric tests were used because of the non-normal distribution of the values. The Wilcoxon paired measures test was used for within-group comparison of data obtained after tetanization with their own baseline values. The Kruskal-Wallis test was used for comparing whole curves obtained for two groups, whilst the Mann-Whitney's U test was used to compare isolated points between two curves. To compare the input-output curves between lesioned and non-lesioned animals linear regression curves were adjusted to the data in each group. The adjustment to linearity of each regression line was assessed with the runs test. To compare the slopes of both regression lines a lineal model of PSA versus EPSP was adjusted, introducing an interaction term (ANCOVA). In every case, $P<0.05$ was considered as statistically significant.

RESULTS

The weak tetanus to the PP induced an E-LTP which remained significantly different from baseline values for the first 2 h ($n=8$, Wilcoxon test; Fig. 1a, open squares). Behavioural stimulation reinforced, i.e. prolonged this potentiation to an L-LTP. Control experiments revealed that the injection of $1 \mu\text{l}$ saline into the BLA did not modify this effect (Kruskal-Wallis test; $n=6$, data not shown). Therefore, both groups were pooled together to form one behavioural reinforcement group ($n=14$, Fig. 1a, closed circles).

The injection of lidocaine into the BLA did not modify baseline potentials in the dentate gyrus evoked by stimulation of the PP ($n=5$, data not shown). Injection of lidocaine to the BLA 5 min after tetanization, did not change the time course of E-LTP ($n=8$, data not shown). However, when this protocol was combined with behavioural stimulation, i.e. drinking 15 min after induction of LTP, the inactivation of the BLA completely blocked the normally reinforcing effect of the appetitive stimulus ($n=8$, Fig. 1b, open circles). In fact, E-LTP even seemed to decay faster by inactivation of the BLA and behavioural reinforcement, and a small, but statistically significant depression was observed 5–6 h after tetanization (Wilcoxon test).

A second series of experiments investigated whether a complete electrical lesion of the BLA influenced the behavioural reinforcement of LTP in the dentate gyrus similarly to the temporal inactivation of the BLA by lidocaine. E-LTP in the dentate gyrus induced by the weak tetanus did not differ from that of non-lesioned animals ($n=9$, data not shown). However, when LTP induction was combined with behavioural stimulation ($n=6$) no reinforcement of E-LTP was detected (Fig. 2 filled squares), an effect identical to the one observed in the lidocaine treated animals (Fig. 2a, open circles). Both curves differed significantly from the one obtained in control animals after behavioural reinforcement (Fig. 1b, filled circles). Stimulation to obtain input-output curves before LTP-induction revealed that the PSA and the slope of the EPSPs were increased amongst lesioned animals. These changes were statistically significant when linear regression curves (EPSP versus PSA) were calculated using the EPSPs as independent and the PSA as dependent variables, respectively. Both curves adjust to linearity ($P>0.5$, runs test), but slopes differed significantly between lesioned and non-lesioned animals analysis of covariance (ANCOVA), $P<0.05$ (Fig. 2b). This could reflect an increased excitability within this group, though it does not seem to be an immediate consequence of the lesion because no such differences were observed in the control potentials recorded at the end of surgery.

DISCUSSION

The results obtained in intact, non-treated animals, confirm previous reports showing that behavioural stimuli with a strong motivational content were able to prolong LTP (Seidenbecher et al., 1995, 1997). The complete abolition of this effect after temporal or permanent inactivation of the BLA supports the hypothesis that this limbic structure is part of the neural system involved in the reinforcement of

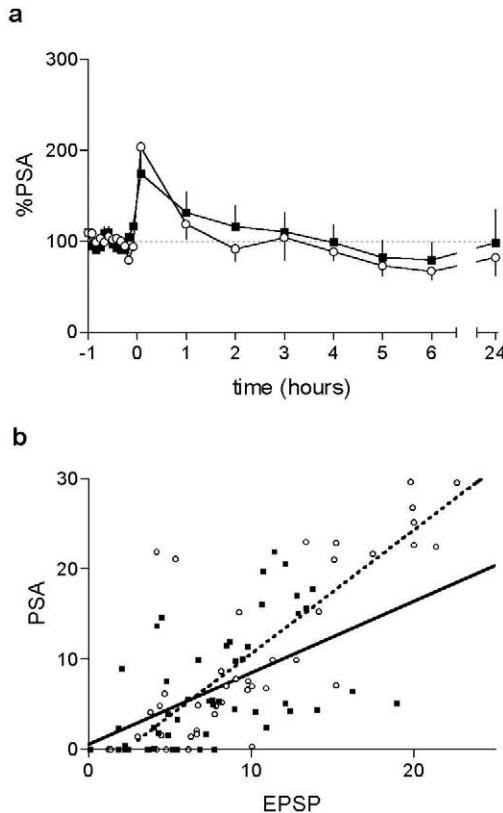


Fig. 2. (a) Lesioning the amygdala (basolateral amygdala lesioning, long-term potentiation [LTP] induction and behavioural stimulation, filled squares, $n=6$) or its temporary inhibition by lidocaine (LTP induction, lidocaine application and behavioural stimulation, open circles, $n=8$) have similar preventing effects on behavioural reinforcing of early-LTP. No differences between curves were found (Kruskal-Wallis test). (b) Lesioning the amygdala increases the slope of the input-output curve, when the slope of the field excitatory postsynaptic potential (EPSP) was plotted against the amplitude of the population spike (PSA). The adjusted regression lines (non-lesioned animals: continuous line; lesioned animals: interrupted line), and scatter plots of the data points (non-lesioned animals: filled squares; lesioned animals: open circles) are shown. The slopes of the respective lines (non-lesioned = 0.7901 ± 0.1869 ms; lesioned = 1.369 ± 0.1252 ms) differed significantly (ANCOVA).

functional plasticity in the hippocampus by motivational stimuli. However, it should be considered that both inactivation methods affect also the fibers crossing through the amygdala and therefore, a possible contribution of these fibers cannot be ruled out. On the other hand, as electrolytic lesions extended beyond the BLA to other nuclei of the amygdala, it could also be the case that the observed effects are not exclusively contributed to the BLA. As no differences were observed in the drinking or motor behaviour of the lesioned or lidocaine-injected animals with regard to controls, it is unlikely that the differences observed in the time course of LTP are caused by such unspecific factors.

The amygdala is part of a system related to emotion and motivation (Quirk et al., 1996) which modulates memory consolidation in other brain structures (Packard et al., 1994; McGaugh and Cahill, 1997). In earlier reports, we

have demonstrated that comparable effects on processes of functional plasticity in the hippocampus can be obtained by direct, electrical stimulation of the amygdala (Frey et al., 2001). We hypothesise that the transition from E-LTP into L-LTP may represent a cellular correlate of memory consolidation. If so, behavioural reinforcement of LTP can represent a cellular mechanism and can be used as a model to study the effect of emotional/motivational influences on memory formation. Interestingly, both processes are dependent on protein synthesis and the induction of L-LTP as well as the transformation from E- into L-LTP requires heterosynaptic mechanisms (Frey and Morris, 1997a,b). Thus, synaptic inputs activated to induce E-LTP are different from those responsible for the reinforcement. In addition, the synapses involved seem to carry different neurotransmitters. In earlier reports, we have shown that noradrenergic and muscarinic, but not glutamatergic receptor antagonists, prevented the reinforcing effect of direct electrical stimulation of the BLA on the granule cells of the dentate gyrus (Frey et al., 2001). In addition, we had shown that the pathway involved in the reinforcing BLA effects on dentate gyrus LTP requires the fimbria-fornix (Jas et al., 2000), a system which provides cholinergic and aminergic innervation to the hippocampus.

Furthermore, we have shown recently that aging affects both amygdala and behavioural reinforcement of LTP (Almaguer et al., 2002; Bergado et al., 2001). Interestingly the combination of E-LTP induction and behavioural reinforcement not only fails to prolong LTP amongst aged rats, but produced a depression of the PP-evoked potentials which was statistically significant beyond 3 h after tetanization. This result is similar in magnitude and the time course to the data obtained here after blocking the BLA with lidocaine. Furthermore, a depression also develops 5–6 h after BLA stimulation when the amygdala is stimulated in young animals without pairing with a PP-tetani-tization protocol. The mechanisms and significance of such effects remain to be investigated, but the long latency suggests the mediation of plastic mechanisms rather than a direct process of inhibition.

Finally, we found that the lesion of the amygdala increases excitability of the granule cells stimulated via the PP as shown by the changed input-output characteristics. Despite this, lesioned animals displayed normal E-LTP, which could not be reinforced by motivational stimulation. Such behaviour suggests that behavioural reinforcement and probably also that induced by the stimulation of the amygdala, is not dependent on changes in the general excitability of the target cells, but on adaptive modifications, likely at the synaptic level, mediated by intracellular signal cascades leading to the synthesis, distribution and processing of plasticity-related molecules (Frey and Morris, 1997a,b).

In summary, our findings demonstrate that the amygdala is part of a behavioural reinforcement system (Frey et al., 2001; Seidenbecher et al., 1997). This assumption is in line with the general concept of the function of this particular limbic structure as a neural substrate of emotions and motivation and their influence on learning and memory.

Our data may add further insights into the complex relationships between affective and cognitive processes, two of the most relevant abilities guiding behaviour.

REFERENCES

- Almaguer W, Estupiñán B, Frey JU, Bergado JA (2002) Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP. *Neurobiol Aging* 23:319–324.
- Bergado JA, Almaguer W, Ravelo J, Rosillo JC, Frey JU (2001) Behavioral reinforcement of long-term potentiation is impaired in aged rats with cognitive deficiencies. *Neuroscience* 108:1–5.
- Frey S, Bergado JA, Seidenbecher T, Pape HC, Frey JU (2001) Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP. *J Neurosci* 21:3697–3703.
- Frey U, Morris RGM (1997a) Synaptic tagging and long-term potentiation. *Nature* 385:533–536.
- Frey U, Morris RGM (1997b) Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 21:181–188.
- Jas J, Almaguer W, Frey JU, Bergado J (2000) Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res* 861:186–189.
- Matthies H (1989) Neurobiological aspects of learning and memory. *Annu Rev Psychol* 40:381–404.
- Matthies H (1998) Neuronale Grundlagen der Gedächtnisbildung. *Enzyklopädie Psychologie* 6:15–42.
- McGaugh JL, Cahill L (1997) Interaction of neuromodulatory systems in modulating memory storage. *Behav Brain Res* 83:31–38.
- Packard MG, Cahill L, McGaugh JL (1994) Amygdala modulation of hippocampal-dependent and caudate nucleus-dependent memory processes. *Proc Natl Acad Sci USA* 91:8477–8481.
- Quirk GJ, Armony JL, Repa JC, Li XF, LeDoux JE (1996) Emotional memory: a search for sites of plasticity. *Cold Spring Harbor Symp Quant Biol* 61:247–257.
- Seidenbecher T, Balschun D, Reymann KG (1995) Drinking after water deprivation prolongs “unsaturated” LTP in the dentate gyrus of rats. *Physiol Behav* 57:1001–1004.
- Seidenbecher T, Reymann KG, Balschun D (1997) A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc Natl Acad Sci USA* 94:1494–1499.

(Accepted 22 October 2002)

Capítulo 3: La estimulación de la amígdala refuerza los procesos de plasticidad sináptica duradera

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Este trabajo fue básicamente exploratorio. Su objetivo principal fue determinar si la estimulación eléctrica de la amígdala podía influir sobre el desarrollo de fenómenos neuroplásticos. La hipótesis que lo inspiró podría referirse como:

La amígdala es una estructura esencial en el procesamiento y establecimiento del estado afectivo(emoción) y el valor de las experiencias vividas (motivación). Sabiendo que factores afectivos pueden modular el curso temporal de la LTP y que la amígdala es parte esencial de este sistema de reforzamiento, debe ser posible lograr el mismo efecto estimulando adecuadamente la amígdala.

Los resultados mostraron que la estimulación de la amígdala en concurrencia temporal con la inducción de la LTP es capaz de prolongar el fenómeno neuroplástico, convirtiendo de hecho, una LTP temprana, con una duración inferior a 4 horas, en una LTP tardía. Esto fue cierto tanto para estímulos de alta frecuencia (200 Hz) como de baja frecuencia (1Hz) como muestran las figuras 2a, b y c, 3a y b. El efecto solo es posible dentro de una ventana temporal de ± 30 min. También el reforzamiento conductual se comporta de manera similar aunque la ventana se extiende hasta una hora.

Otra similitud de la estimulación de la amígdala y el reforzamiento conductual es su dependencia de la síntesis de proteínas, como se demostró al observar el bloqueo del reforzamiento inducido por la amígdala en animales tratados con anisomicina intraventricular (fig. 6).

Se exploró además cuales neurotransmisores podrían estar involucrados en este efecto, mediante el empleo de bloqueadores específicos. El bloqueo de receptores de tipo

NMDA (usando AP-5, ver figura 4) o dopaminérgicos tipo D1 (SCH23390) no impidió el reforzamiento de la LTP por estimulación de la amígdala (fig. 5b). Esto sugiere que los sistemas glutamatérgicos y dopaminérgicos no intervienen en el mecanismo de ese reforzamiento en el giro dentado. Sin embargo el bloqueo de receptores noradrenérgicos (propranolol, fig. 5a) o colinérgicos de tipo muscarínico (atropina, fig. 5c) abolió completamente el efecto de reforzamiento lo cual sugiere que ambos neurotransmisores forman parte del mecanismo neural implicado en el efecto de reforzamiento descrito en este artículo.

Reinforcement of Early Long-Term Potentiation (Early-LTP) in Dentate Gyrus by Stimulation of the Basolateral Amygdala: Heterosynaptic Induction Mechanisms of Late-LTP

Sabine Frey,¹ Jorge Bergado-Rosado,² Thomas Seidenbecher,³ Hans-Christian Pape,³ and J. Uwe Frey¹

¹Leibniz-Institute for Neurobiology, Department of Neurophysiology, PF 1860, D-39008 Magdeburg, Germany, ²Centro Internacional de Restauración Neurológica CIREN, Laboratory of Experimental Electrophysiology, CH21, Havana, Cuba, and ³Institute for Physiology, Medical Faculty of the Otto-von-Guericke-University, 39120 Magdeburg, Germany

The basolateral amygdala (BLA) can influence distinct learning and memory formation. Hippocampal long-term potentiation (LTP), the most prominent cellular model of memory formation, can be modulated by stimulation of the BLA in its induction and early maintenance. However, it is not known how the late maintenance of LTP beyond its initial phases might be affected. Behavioral stimuli have been shown to result in a reinforcement of a transient early-LTP into a lasting potentiation. Here we show that BLA stimulation mimics the behavioral effects on early-LTP in freely moving rats when the BLA is activated within

a time window of 30 min before or after tetanization of the perforant path. The reinforcement of LTP was blocked by inhibitors of muscarinic and β -adrenergic but not dopaminergic receptors and was dependent on translation. Through these heterosynaptic associative interactions, hippocampal sensory information can be stabilized by amygdaloid influences.

Key words: long-term potentiation; reinforcement; heterosynaptic LTP; associative LTP; late-LTP; basolateral amygdala; hippocampus; dentate gyrus

The hippocampus is important for the formation of certain kinds of memory. Although the information presented to and sent from the hippocampal network remains uncertain, hippocampal neurons exhibit a number of intriguing biophysical properties that enable them to participate in aspects of memory formation. These include mechanisms of synaptic plasticity that can respond to incoming information by detecting associative interactions between presynaptic, postsynaptic, and heterosynaptic activity and register these conjunctions as an increase in synaptic weights (Frey and Morris, 1998a). The latter process was named long-term potentiation (LTP) (Bliss and Lomo, 1973), which has become the best-studied cellular model of memory formation. Interestingly, hippocampal LTP exhibits similar temporal stages as described for certain types of hippocampus-dependent memory. An early-LTP (with a duration of ~4–6 hr) can be dissociated from late-LTP (beyond 6 hr) by inhibition of specific protein kinases, by protein synthesis, and partially by mRNA synthesis (Krug et al., 1984; Frey et al., 1988, 1996; Frey, 1997; Frey and Morris, 1997, 1998a).

In recent studies, Seidenbecher et al. (1997) reported that early-LTP in the dentate gyrus (DG) was reinforced if an appetitive or aversive stimulation was presented within 30 min after LTP induction. This reinforcement was dependent on the activation of β -adrenergic receptors. It was speculated that the consol-

idation of a memory trace in the hippocampal formation, as part of a more complex memory processing system, is reinforced if a modifying, most likely extra-glutamatergic input is active within a distinct time interval. Because late-LTP in the hippocampus requires similar heterosynaptic processes (Frey, 1997; Frey and Morris, 1998a), we hypothesize that the synergistic action of different transmitter systems and therefore different brain structures are needed for the induction of cellular processes leading to the long-lasting consolidation of a memory trace. However, the brain structures involved in reinforcing LTP in the DG remain unclear.

Interestingly, stimulation of the BLA, a structure thought to be part of an emotional memory system, can influence the induction and early maintenance of DG LTP (Ikegaya et al., 1995a; Akirav and Richter-Levin, 1999) by aminergic mechanisms (Ikegaya et al., 1997), whereas lesion of the amygdala attenuates DG LTP (Ikegaya et al., 1994). These results led us to investigate whether the maintenance of DG LTP can also be modulated by BLA stimulation. Here, we have studied the effect of BLA stimulation on the early-LTP in DG. If LTP subserves cellular mechanisms during declarative learning, then a hypothetical heterosynaptic associative LTP reinforcement by BLA stimulation could be of special importance. Behavioral experiments by others (for review, see Cahill and McGaugh, 1998) have shown similar processes with respect to emotional arousal and the maintenance of declarative memory. Electrophysiological studies on the induction and early transient stages of LTP in DG (Ikegaya et al., 1995a,b, 1997; Akirav and Richter-Levin, 1999) revealed modulatory effects of the BLA on hippocampal function, but it is not known whether protein synthesis-dependent late LTP is also affected, a prerequisite for long-lasting memory traces to be formed. Interestingly, it has been shown previously that hippocampus-dependent long-term memory processes are influenced by the amygdala (Bevilacqua et al., 1997; Bianchin et al., 1999; Izquierdo et al., 1999). Here

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J.B.-R. and T.S. contributed equally to this work.

Correspondence should be addressed to Dr. J. U. Frey, Leibniz-Institute for Neurobiology, Department of Neurophysiology, Brenneckestrasse 6, PF 1860, D-39008 Magdeburg, Germany. E-mail: frey@ifn-magdeburg.de.

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we report that early-LTP in the DG *in vivo* can be influenced in its maintenance by stimulation of the BLA.

MATERIALS AND METHODS

Subjects and surgery. Experiments were performed on 8-week-old Wistar rats (200–250 gm). All experiments were performed in compliance with the relevant laws and institutional guidelines and have been approved by the Land Sachsen-Anhalt.

For chronic implantation of electrodes, the animals were anesthetized with pentobarbital [40 mg/kg, i.e., 40 mg was dissolved in 10 ml saline and 1 ml/100 gm was injected intraperitoneally as an initial dose, supplemented by 0.3 (1.2 mg)–0.5 ml (2 mg) intraperitoneal injections as necessary] and placed in a stereotaxic frame (David Kopf Instruments, Tujunga, CA). The scalp was incised and retracted, and head position was adjusted to place bregma 1 mm higher than lambda. Small holes were drilled in the skull for the placement of stimulating and recording electrodes. The electrodes consisted of insulated stainless steel wires 125 μ m in diameter. A monopolar recording electrode was placed in the DG–granular cell layer [coordinates -2.8 mm posterior to bregma (AP), 1.8 mm lateral to midline (L), 3.2 – 3.5 mm ventral from dura (V); coordinates from the atlas of Paxinos and Watson (1998)]; a bipolar stimulating electrode was implanted in the medial perforant pathway (AP -6.9 mm, L 4.1 mm, V 2.4 – 2.7 mm). For stimulating the amygdala, a monopolar electrode was placed into the basolateral amygdala (AP -3.5 mm, L 5.0 mm, V 7.6 mm) with an indifferent electrode consisting of silver bar wire lowered on dura anterior to the stimulating electrode. The electrodes were adjusted to optimize the population spike (PS) amplitude in the perforant path–DG system. All rats were allowed 8–10 d recovery after surgery before the electrophysiological experiments in freely moving animals. The positioning of electrodes was checked in each animal histologically after the end of the experiment, and only those animals with a correct positioning of the electrodes were included in further analyses [interestingly, no effects of DG LTP were detected in experiments in which the histological confirmation of electrode positioning afterward revealed a placement of the amygdala-stimulating electrode in the central instead of the basolateral nucleus (data not shown)].

Recording. All electrophysiological recordings were performed in special experimental boxes, where animals were connected by a flexible cable to a 10-channel swivel that allowed them to move freely with *ad libitum* access to water and food (Frey et al., 1996; Seidenbecher et al., 1997). Biphasic current pulses (0.1 msec per cycle, 150 – 250 μ A) were applied to the perforant path to evoke extracellular field potentials in the DG of $\sim 40\%$ of the maximal PS. PS recording and analysis were favored against the slope of field EPSPs because the latter is relatively unstable in the hilar region of the dentate gyrus in freely moving animals, especially if taken into consideration that the stimulation intensity was adjusted to obtain a population spike that influenced strongly the dipole of the field EPSP in the hilus. The spike, however, is required to induce LTP. A few experiments showed a reasonable, larger field EPSP that could be used for calculations, and we provide representative examples of the time course of the field EPSP for crucial experiments below. Analyses of these experiments revealed similar results as PS measurements, suggesting that the recorded and analyzed PS is not just a measure of changes in excitability but also represents adequate synaptic function. This is supported by the fact that, to our knowledge, LTP has never been reported to be associated with changes of excitability. The basolateral amygdala was stimulated by impulses with an intensity of standardized 300 μ A independent of the stimulation protocol (biphasic constant current pulses, 0.2 msec duration per polarity). This stimulation intensity evoked an average BLA DG potential as shown in Figure 1a.

After a stable baseline was recorded for at least 30 min (recordings every 5 min), an “unsaturated” LTP was induced by three bursts of 15 impulses, 200 Hz, 0.2 msec pulse width each stimulus, interburst interval 10 sec (“weak tetanus”), resulting in a potentiation that decayed within 4–7 hr to pretetanus value. In the series with late-LTP, tetanization consisted of 20 bursts of 15 impulses, 200 Hz, 0.2 msec pulse width each stimulus, interburst interval 10 sec (“strong tetanus”). This stimulation paradigm resulted in late-LTP with a duration of 8 hr, the longest time point we have investigated. Averaged responses were recorded every 15 min for up to 8 hr after tetanization.

For estimation of the time window for “reinforcement” of the unsaturated DG LTP by stimulation of the basolateral amygdala, the following stimulating protocols were used. At various time points (5, 15, and 30 min) before or after tetanization of the perforant path, the basolateral amygdala was stimulated by high frequency [three bursts of 15 impulses,

200 Hz, 0.2 msec pulse width each stimulus, interburst interval 10 sec (weak tetanus) at 300 μ A] or low frequency (45 impulses at 0.1 Hz, 0.2 msec pulse width, each stimulus at 300 μ A).

Pharmacology. Substances were applied intraventricularly through chronically implanted cannulas [anterior horn of the right lateral ventricle, for detail, see Seidenbecher et al. (1997)]. Propranolol (6.76 nmol), SCH 23390 (3.08 nmol), and atropine (1 nmol) (Sigma, St. Louis, MO) were injected 5 min after tetanization of the PP, that is, 10 min before BLA tetanization. The selected doses of propranolol and SCH 23390 have been shown previously to be effective (Balschun et al., 1997; Seidenbecher et al., 1997). The used concentration of atropine was able to inhibit oxotremorine-induced hippocampal theta EEG activity (Malisch and Ott, 1982) (intraperitoneal application of 0.2 mg/kg oxotremorine sesquifumarate; data not shown). Application of the drugs 5 min (propranolol, SCH 23390, atropine) or 10 min (AP-5, 100 nmol, from RBI) after perforant path (PP) tetanization (control experiments without stimulation of the BLA) did not influence the time course of early LTP [for propranolol, see Seidenbecher et al. (1997); other data not shown]. All of the above substances had no effect on baseline evoked potentials nor on early-LTP, with the exception of AP-5, which blocked early LTP when applied before PP tetanization (Fig. 4a). The latter result confirms earlier results that compounds with similar biophysical properties can diffuse within 5 min to their place of action, i.e., from the ventricle to the dentate gyrus. Anisomycin (0.905 mol; ICN Biochemicals, Costa Mesa, CA) was injected 2 hr before the PP was tetanized. To avoid possible nonspecific side effects of the presence of the reversible protein synthesis inhibitor anisomycin after intracerebroventricular injection on recorded control field potentials in the DG, the time point was first determined at which anisomycin was still effective in inhibiting protein synthesis when applied sufficiently before induction of LTP. This was achieved by measuring the incorporation of radioactive-labeled amino acids into hippocampal proteins. It was found that anisomycin inhibited the incorporation of amino acids into hippocampal proteins by $>90\%$ for at least 2 hr after its application (data not shown), the time at which LTP was induced. In the series with anisomycin and LTP induction, only those experiments with normal post-tetanic potentiation were used for statistical evaluation. In all cases the injection was performed at 1 μ l/min to a total volume of 5 μ l.

Statistics. Data analyzed here are from non-Gaussian populations but show near identical shapes of distributions. Therefore, nonparametric tests were performed. Within-group comparisons were made using the Wilcoxon test for paired samples. For comparisons between groups the Mann–Whitney *U* test was used after performing the Kruskal–Wallis test for the different groups. Differences were considered statistically significant only when $p < 0.05$ in Kruskal–Wallis and the post test. For clarity when comparing data, the mean of percentage change of the PS amplitude measured in millivolts $\pm$ SEM is shown.

RESULTS

Our studies revealed that low-frequency control stimulation of the perforant path or the BLA did not dramatically influence DG potentials (Fig. 1). Weak tetanization of the perforant path resulted only in early-LTP decaying to baseline values within 8 hr (Fig. 1b, ●). Strong tetanization, in contrast, revealed late-LTP with a duration of at least 8 hr, the latest time point we have investigated (Fig. 1b, ○).

BLA stimulation before and after tetanization of DG

We then investigated whether stimulation of the BLA before, simultaneously with, or after tetanization of the perforant pathway influences early-LTP in the DG. As shown in Figure 2a (●), the duration of LTP in the DG, induced by a tetanization protocol that normally would lead only to early-LTP, can be influenced by a subsequent high-frequency stimulation of the BLA applied 15 min after the tetanization of the perforant path. Although short-term potentiation (STP) (<1 hr) was not influenced, the maintenance of LTP was changed significantly. The transient time course of LTP induced by weak tetanization of the perforant path was transformed (or reinforced) to a long-lasting potentiation with a duration of at least 8 hr. As mentioned earlier, for technical reasons the DG LTP reinforcement by BLA stimulation

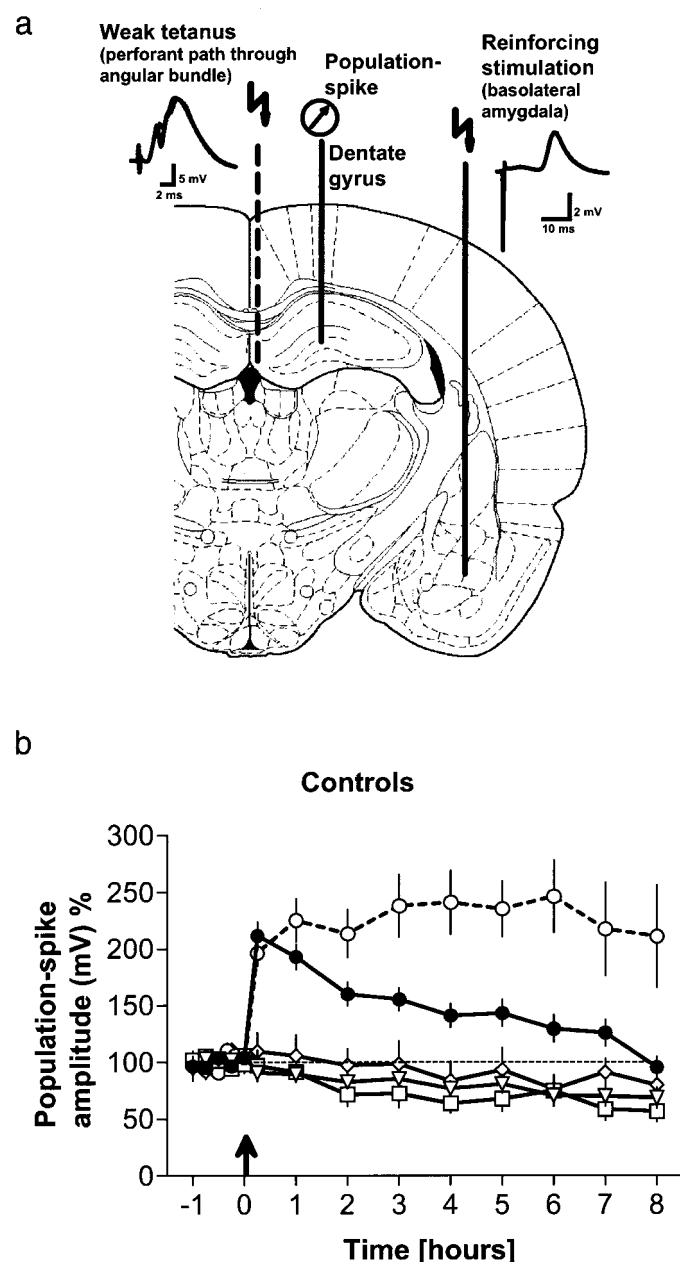


Figure 1. Reinforcement of hippocampal early-LTP by stimulation of the basolateral nucleus of the amygdala in freely moving rats. *a*, Schematic illustration of electrode localization. For clarity, in this section the DG stimulation electrode is shown activating the perforant pathway (broken line). Originally, this electrode was positioned in the angular bundle (see Materials and Methods). Insets show analog examples of recordings obtained before (dotted line) and after (filled line) LTP induction of the perforant path (top left) and after stimulation of the BLA (top right). *b*, Control recordings: induction of early-LTP (●, $n = 18$) or late-LTP (○, $n = 5$) by a weak or strong tetanus of the perforant path, respectively, or tetanization of the BLA (∇, $n = 11$), or after application of 45 impulses at an LFS in the BLA (□, $n = 11$), and finally after LFS of the perforant path alone (◇, $n = 6$). Low-frequency stimulation of the perforant path did not severely influence baseline potentials for the investigated 8 hr (Fig. 1*b*, ◇). A statistically significant difference was detected only at 6 hr after low-frequency stimulation when compared with prestimulation values (Wilcoxon test, $p < 0.05$). High-frequency (Fig. 1*b*, ∇) or low-frequency stimulation (Fig. 1*b*, □) of the BLA resulted in a slowly developing long-term depression at the DG synapses. At 2 hr a statistically significant depression of the initial baseline potentials was observed [$82.9 \pm 4.18\%$ (millivolts; percentage change $\pm$ SEM)] in the group with a high-frequency train to the amygdala and $71.6 \pm 10.24\%$ in the group with

was measured as percentage changes of the PS amplitude in general. However, the EPSP was similarly affected, as confirmed by the following example: the level of EPSP potentiation was 121.1% at 1 hr after tetanization and 84.7% at 8 hr after tetanization of the DG alone versus 119.2 and 124.6% after DG tetanization followed by BLA stimulation 15 min later.

Similar results were obtained when a low-frequency stimulation (LFS) pattern instead of a weak high-frequency train was applied to the BLA (Fig. 2*b*). Late-LTP induced by a strong tetanization protocol was not influenced by BLA stimulation 15 min after LTP induction (Fig. 2*c*, ■).

A series of experiments was then conducted exploring a possible time window in which the stimulation of the BLA and perforant pathway results in long-lasting associative plastic changes. Figure 3 summarizes the amount of potentiation obtained in the DG 15 min (Fig. 3*a*) and 8 hr (Fig. 3*b*) after tetanization of the perforant pathway when the BLA was stimulated either (1) before, (2) at the same time as, or (3) after tetanization of the perforant path. The time interval between stimulation of the two structures varied from 5 to 15 to 30 min. As shown in Figure 3*a*, the potentiation was transiently enhanced 15 min after LTP induction when either the BLA and perforant path stimulation were applied simultaneously or the BLA stimulation preceded perforant path tetanization.

Figure 3*b* shows the effect of BLA stimulation on the maintenance of LTP induced by a weak tetanus (after 8 hr). Simultaneous stimulation of the BLA and perforant path did not influence the time course of a weak potentiation in the DG. BLA tetanization 5 or 15 min after a weak tetanization of the perforant path resulted in a reinforced potentiation that was now long lasting (8 hr). Similar results were obtained when the BLA was tetanized at 5 and 15 but also at 30 min before perforant path tetanization, although with a decaying degree of reinforcement. The time course of the field EPSP paralleled the PS as shown by the following examples: the level of EPSP potentiation at the 15 min interval between BLA and DG tetanization ($n = 3$) was $116.7 \pm 1.33\%$ at 1 hr and $113.5 \pm 3.47\%$ at 8 hr after LTP induction; the 30 min interval ($n = 3$) results were $125.9 \pm 4.09\%$ at 1 hr versus $94.5 \pm 9.47\%$ at 8 hr after LTP induction. We do not know whether an increase in the time interval between BLA and perforant path tetanization beyond 30 min results in a reinforced long-lasting potentiation. Future experiments should be conducted to investigate this time window more thoroughly.

Involved transmitter systems

The next series of experiments was conducted to elaborate which transmitter systems are involved in the reinforcing effect of amygdala stimulation on early LTP in the DG. A number of transmitters, including dopamine, norepinephrine, acetylcholine, and opioids, are known to modulate LTP (Dunwiddie et al., 1982; Bliss et al., 1983; Krug et al., 1983; Stanton and Sarvey, 1985). We showed earlier (for review, see Frey, 1997; Frey and Morris, 1998a) that late-LTP requires the heterosynaptic activation of nonglutamatergic receptors during tetanization. In a control ex-

low-frequency stimulation, respectively] that remained at this level until the end of the experiment (Wilcoxon test, $p < 0.05$). However, statistical comparison of the BLA-stimulated control series revealed no significant difference (Mann–Whitney U test) with the exception at 7 hr (low-frequency perforant path control vs low-frequency BLA stimulation). The arrow indicates the time point of weak or strong tetanization or LFS, respectively.

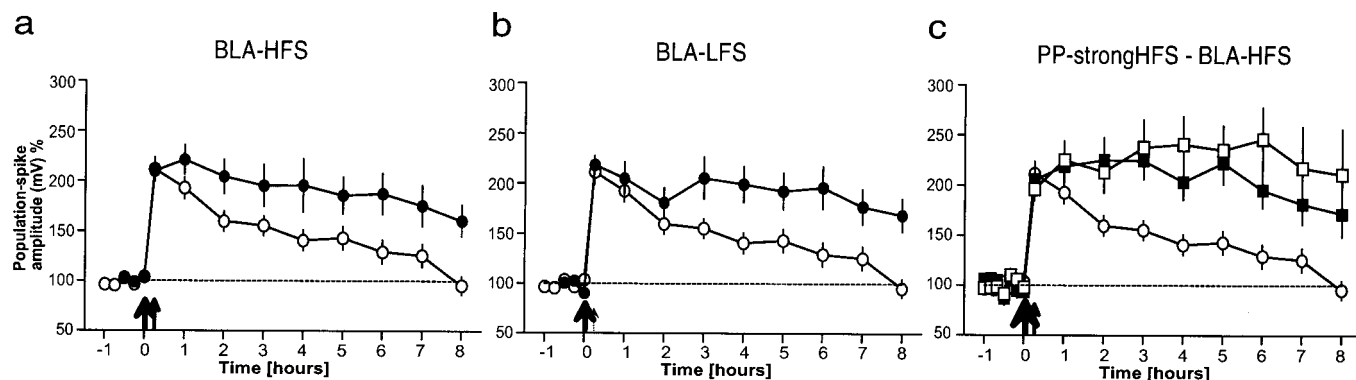


Figure 2. Reinforcement of early- but not late-LTP by high- and low-frequency BLA stimulation 15 min after tetanization of the DG. Reinforcement of early-LTP when the BLA was stimulated with either a high-frequency train (*a*, thin arrow) or a low-frequency stimulation (*b*, dotted arrow) 15 min after weak tetanization (thick arrow) of the DG (●, $n = 11$ for each series) is shown. Open circles indicate the time course of early-LTP when induced in the DG without BLA pairing ($n = 18$). LTP in the reinforced group was statistically significantly different from the nonpaired group from the second hour onward after tetanization of the DG (Mann–Whitney U test). *c* represents no influence on late-LTP when the BLA was tetanized 15 min after LTP induction in the perforant path DG by a strong tetanization protocol [□ represents a control late-LTP series of the DG alone; $n = 5$; ■ shows strong tetanization of DG (larger arrow) followed by BLA tetanization (thin arrow) 15 min after its induction; $n = 5$]. Strong DG LTP (□) and strong DG BLA LTP (■) were statistically significantly different from weak LTP (○) from the second hour onward (Mann–Whitney U test).

periment it was now investigated whether a direct, additional glutamatergic activation initiated by BLA stimulation was sufficient and responsible for the reinforcement of early LTP in the DG or whether additional transmitter systems are required. The possibility that subsequent tetanization of glutamatergic pathways can result in a greater level of potentiation attributable to a stronger depolarization and subsequent activation of NMDA receptors leads one to assume that the described reinforcement was triggered by such a mechanism (i.e., saturation of LTP by repeated tetanization of glutamatergic afferents). The NMDA receptor antagonist AP-5 was therefore applied intracerebroventricularly after tetanization of the perforant path. If AP-5 was injected 5 min after PP tetanus, the initial level of potentiation (late time points of post-tetanic potentiation) was influenced (data not shown). Therefore, AP-5 was injected 10 min after PP tetanus, i.e., 5 min before BLA stimulation, which was sufficient for the drug to perfuse to the DG. Control experiments revealed that AP-5 blocked LTP induction in the DG when applied intracerebroventricularly 5 min before tetanization (Fig. 4*a*). As shown in Figure 4*b*, the NMDA receptor antagonist did not influence the time course of reinforced LTP.

Figure 5 illustrates the action of the other tested receptor blockers on the reinforced potentiation. As shown in Figure 5, *a* and *c*, only the β -adrenergic receptor antagonist propranolol and the muscarinic antagonist atropine, but not the dopaminergic D1 receptor antagonist SCH 23390 (Fig. 5*b*), were effective in blocking the reinforced potentiation. Similar time courses were obtained in experiments with measurable field EPSPs [e.g., propranolol (single experiment): 117.1% at 1 hr vs 90.7% at 8 hr after LTP induction; and atropine ($n = 2$): 116.5 $\pm$ 4.40% vs 91.1 $\pm$ 5.55%]. When one of the two blockers was delivered into the ventricle after LTP induction in the DG, but 10 min before tetanization of the BLA, only early-LTP was seen, as was the case in the control experiments with weak tetanization of the perforant path alone.

Protein synthesis dependence of reinforcement

A last series of experiments investigated whether the reinforcement of early-LTP was accompanied by protein synthesis (Fig. 6). The protein synthesis inhibitor anisomycin was applied 2 hr

before tetanization to avoid possible effects on LTP induction (Fig. 6*a*). Under these conditions, early-LTP by weak tetanization of the perforant path could be induced, although the duration was shorter when compared with control early-LTP in nontreated groups (indicating a distinct requirement of protein synthesis during early-LTP). The reinforcing effect on early-LTP by subsequent BLA stimulation was prevented by anisomycin, suggesting a requirement of protein synthesis for the transformation from early- to late-LTP.

DISCUSSION

In summary, it was shown that only a weak transient form of LTP is affected by BLA stimulation in the intact animal. Induction of late-LTP by a strong tetanus in the DG is not influenced in either its induction or its maintenance, at least during the 8 hr after tetanization that we have investigated (Fig. 2*c*). Heterosynaptic, late associative effects on early-LTP occur when the amygdala is stimulated within a distinct time interval, before or after induction of LTP in the DG. Our pharmacological experiments using the NMDA receptor inhibitor AP-5 revealed that BLA stimulation does not interfere with the reinforcing effects in DG via a direct glutamatergic innervation, as would be expected during saturation experiments in which the same glutamatergic inputs are tetanized subsequently until an asymptotic level of potentiation is achieved (Barnes et al., 1994; Moser et al., 1998). However, direct activation of AMPA receptors in the DG by BLA stimulation cannot be excluded. The subsequent depolarization of granular cells and activation of voltage-dependent calcium channels therefore could be involved in the processes, resulting in a reinforced potentiation. However, the absence of morphological data describing a direct innervation of the DG by BLA makes this assumption unlikely (Pikkarainen et al., 1999).

We have proposed recently that consolidation of hippocampal LTP requires the synergistic activation of both glutamatergic inputs and an additional modulating transmitter system during the induction of LTP, during which coactivation of the latter is necessary to trigger the synthesis of plasticity-related proteins (for review, see Frey and Morris, 1998a). This is a prerequisite for the formation of late-LTP, i.e., its consolidation. Artificial

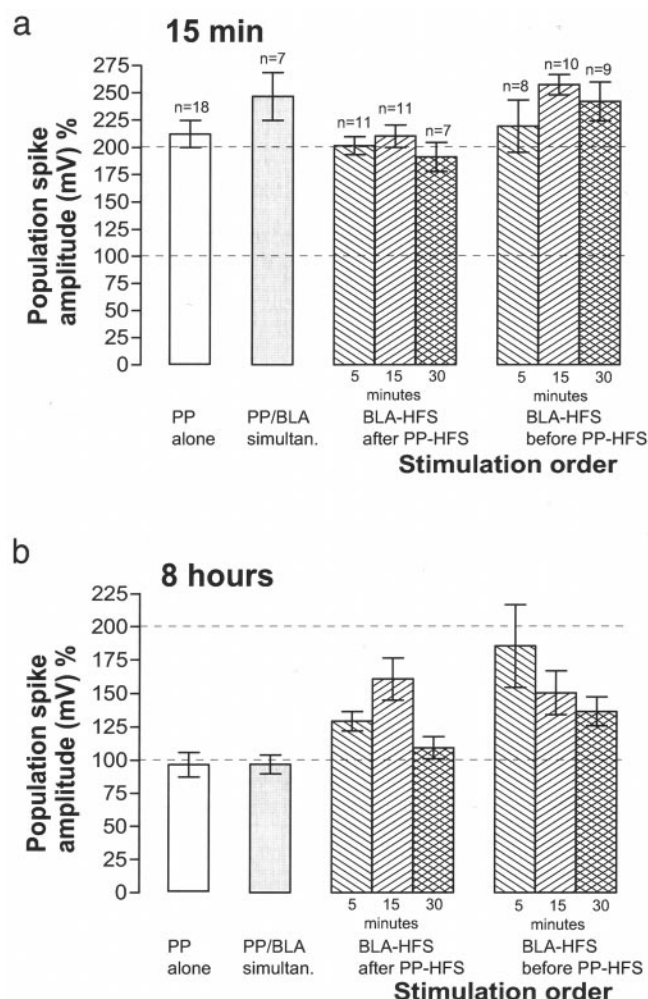


Figure 3. Time window of LTP reinforcement of DG LTP by BLA stimulation. Level of potentiation 15 min (*a*) and 8 hr (*b*) after induction of early-LTP in DG to illustrate the effect of BLA stimulation on the induction of DG LTP (*a*) and maintenance (*b*). Only the series of simultaneous stimulation of the BLA and DG and the series during which the BLA was stimulated 15 min before perforant path tetanization showed a statistically significant enhanced STP measured 15 min after LTP induction (*a*). The potentiation at 8 hr (*b*) returned to pretetanzation levels when the perforant pathway was tetanized alone or simultaneously with BLA, but not when the BLA was stimulated within 15 min before or after LTP induction in DG. In the series during which LTP was induced in the DG after BLA stimulation with a 30 min interval, a remaining statistically significant potentiation was still observed, which is probably attributable to the initial effect on STP by BLA stimulation.

electrical field stimulation in the brain may lead to late LTP if the stimulus is strong enough to reach sufficient heterosynaptic inputs, as could be the case for the series with strong tetanization that is shown here. More subtle stimulation may involve homosynaptic inputs that initiate only early-LTP if not preceded or followed by activation of an additional input of another transmitter system. However, the latter protocol seems to be the more physiological way of neuronal functioning.

Early-LTP in the DG in the intact animal can be transformed into protein synthesis-dependent late-LTP by heterosynaptic and associative mechanisms when both the perforant pathway and the basolateral nucleus of the amygdala are stimulated within a time window of ~30 min. The order of “paired” stimulation is not important, i.e., whether tetanization of the DG or electrical

stimulation of the BLA occurred first, and it is not important whether a high-frequency train or a low-frequency stimulation was delivered to the BLA. With respect to the order of stimulation, our data seem to be in contrast to earlier findings in which DG LTP was reinforced by behavioral stimuli only if the latter was presented after tetanization (Seidenbecher et al., 1997). A possible explanation could be that the behavioral and aversive stimuli used in these experiments involve the serial and parallel interaction of more complex structures at different times than the artificial direct stimulation of the BLA at a given time. However, regarding the order of stimulation, our results are in accordance with the properties of “synaptic tagging” (Frey and Morris, 1998b) (see below).

BLA action on DG LTP is neither direct (Pikkarainen et al., 1999) nor mediated by NMDA receptor stimulation but triggered through brain structures carrying norepinephrine or acetylcholine, or both, but not dopamine. However, because the receptor blockers were applied intraventricularly, a direct action of noradrenergic or muscarinic processes, or both, in the BLA cannot be excluded. Therefore, the reinforcement of DG LTP by BLA stimulation can also involve additional mechanisms such as the action of “modulated” BLA stimulation on different receptor systems and the BLA-dependent regulation of stress hormones required for normal hippocampal function (Brinton and McEwen, 1989; Cahill and McGaugh, 1998; Ferry et al., 1999). This would resemble findings in which hippocampus-associated learning is strongly influenced by BLA modulation (Packard et al., 1994; Cahill et al., 1995; Cahill and McGaugh, 1998).

Questions remain such as why simultaneous tetanization of the DG and BLA does not influence the maintenance of early-LTP (data not shown). It can be speculated that the effect of BLA stimulation on DG LTP is triggered by synergistic actions of glutamatergic and nonglutamatergic mechanisms that require a sequence of processes to be activated. It is not important which of the systems was activated first, but a simultaneous activation prevents the induction of events leading to the reinforcement of LTP. Heterosynaptic, nonglutamatergic receptor activation during tetanization may negatively influence the required level of depolarization for late-LTP to occur [e.g., BLA-stimulated norepinephrine release activates hippocampal interneurons (Bergles et al., 1996)]. Another possibility might be that simultaneous heterosynaptic, i.e., glutamatergic and β -adrenergic, receptor activation in the dentate gyrus cannot sufficiently shift processes such as intracellular calcium transients (Stanton and Heinemann, 1986; Gray and Johnston, 1987) required for late-LTP. The different regulation of calcium may then influence the balance between activated kinases and phosphatases in favor of short-lasting plastic events (Coussens and Teyler, 1996).

A 5 min delay of subsequent stimulation of the two brain structures, however, revealed reinforced LTP. This could have been achieved by heterosynaptic stimulation of the cAMP/PKA cascade, a prerequisite for late-LTP to occur (Frey et al., 1993), or by hormone-dependent regulation of plasticity-relevant proteins in the hippocampus through cAMP/PKA-dependent processes initiated either in the hippocampus (Bevilaqua et al., 1997) or indirectly in the BLA (Frey and Morris, 1998a). In addition, it cannot be ruled out that other brain structures directly interact with the granular cells in the DG because the BLA does not directly innervate the DG granular cells (Pikkarainen et al., 1999). BLA activation requires the additional stimulation of as yet unidentified structures directly connected with the DG by adrenergic or muscarinic fiber systems or influences the level

Figure 4. NMDA receptor activation for BLA-reinforced DG LTP is not required. The protocol with a 15 min time interval between perfortant path and BLA tetanization resulted in a stable reinforced potentiation of DG LTP and therefore was chosen for the pharmacological studies (Figs. 4b, 5, ○). This figure illustrates that the NMDA receptor antagonist AP-5 is effective in blocking early-LTP if applied intracerebroventricularly 5 min (thin arrow above) before tetanization of the DG (●, drug-treated group, $n = 6$; ○, induction of control early-LTP, $n = 15$) (a) and ineffective in influencing the BLA-reinforced DG LTP [●, $n = 7$; the drug (thin arrow above) was applied intracerebroventricularly 5 min before BLA stimulation (thin arrow below)] (b). As a control, BLA-reinforced DG LTP, a series was performed in which physiological NaCl was applied instead of AP-5 (○, $n = 9$). Other symbols are denoted as in Figure 2.

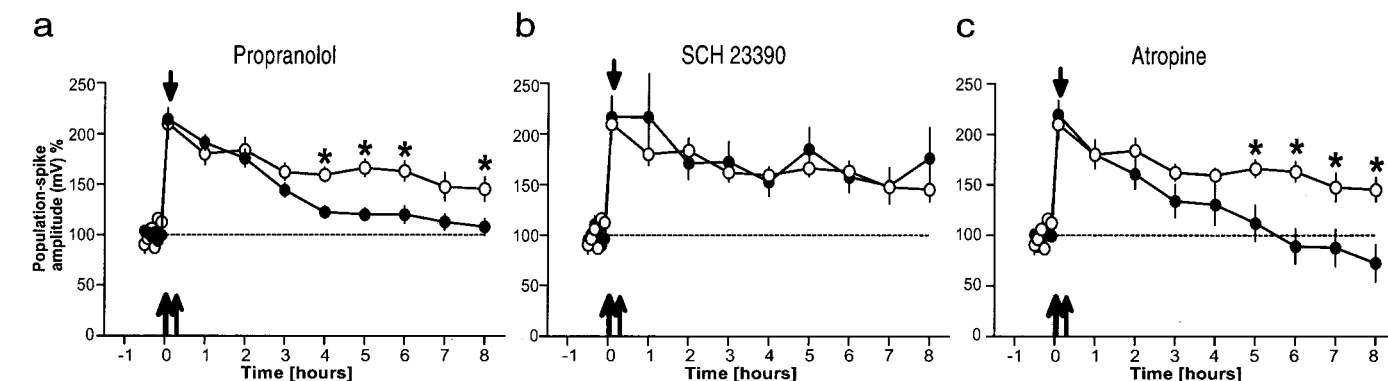
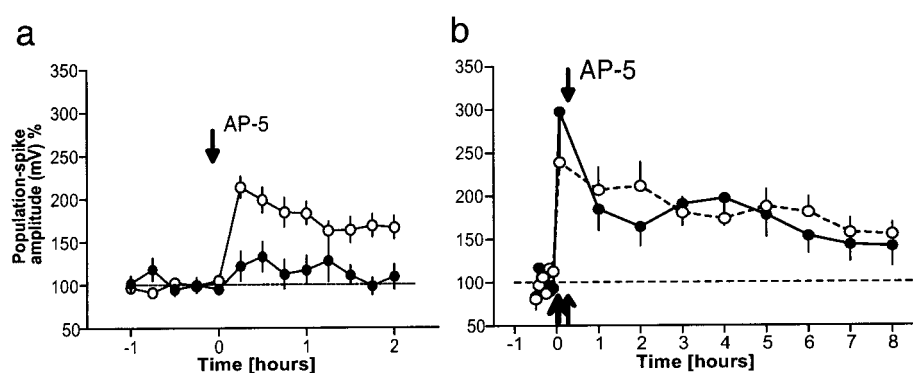
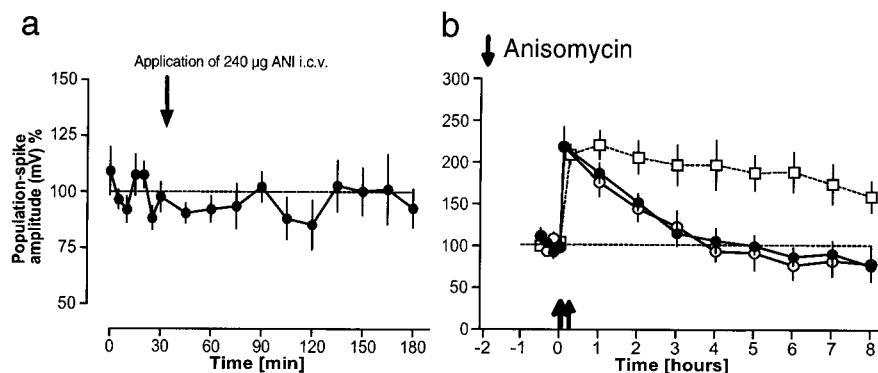


Figure 5. Pharmacology of BLA-reinforced DG LTP. Shown is the effect of the β -adrenergic-receptor antagonist propranolol (●, $n = 13$) (a), the dopaminergic D1 receptor antagonist SCH23390 ($n = 6$) (b), and the muscarinic receptor antagonist atropine ($n = 8$) (c) on BLA-reinforced DG LTP. Asterisks illustrate a statistically significant difference of the drug-treated group when compared with NaCl controls (○, $n = 9$; Mann–Whitney U test). Other symbols are denoted as in Figure 4.

Figure 6. Protein synthesis and BLA-reinforced DG LTP. a, The effects of intracerebroventricular application of 240 μ g of anisomycin on DG potentials are shown ($n = 7$). b, The reversible protein synthesis inhibitor anisomycin prevents the BLA reinforcement of DG LTP when applied intracerebroventricularly 2 hr before LTP induction (●, $n = 7$). The drug did not affect a control early-LTP that was not paired with BLA stimulation (○, $n = 6$) illustrating a specific effect of anisomycin on the reinforcing effect. Squares indicate a control series with reinforced LTP ($n = 10$). Other symbols are denoted as in Figure 4.



of stress hormones, which may then finally modulate neuronal plasticity in the DG.

Interestingly, our studies revealed a model to study early and late associative components of long-lasting plastic changes with an interaction of heterosynaptic components within the minute or even hour range, respectively. Further studies will determine the key players and the locus of action of BLA-dependent reinforcement of DG LTP.

The described time window, the protein synthesis dependence of the reinforcement, and the heterosynaptic associative components of these processes lead us to speculate that the described effects might be related to a phenomenon that we have recently described as synaptic tagging (Frey and Morris, 1997, 1998a).

Synaptic tagging characterizes a late associative property of LTP, which requires the transient setting of a synaptic tag with the function of capturing and processing plasticity-related proteins, thus facilitating consolidation, from a short-term into a long-lasting synaptic plastic change. Interestingly, it has been shown that under distinct circumstances the setting of the tag is sufficient to result in late-LTP at that input, if a heterosynaptic input was stimulated within a specific time window.

Considering the proposed role of LTP in information processing and the described interaction of the hippocampus and amygdala during distinct learning tasks (LeDoux, 1993; Packard et al., 1994; Cahill et al., 1995; Izquierdo and Medina, 1997; Cahill and McGaugh, 1998; Roozendaal et al., 1999), the data presented

here may provide a hint for more detailed investigation of inter-structural, associative interactions at the cellular level. We have shown that BLA stimulation can modulate hippocampus-specific long-lasting plasticity beyond its induction and early maintenance. However, future studies will show whether cellular consolidation under these conditions exceeds the investigated 8 hr. Our data support the hypothesis that describes the amygdala as a structure involved in the formation/modulation of declarative memory in other brain structures that might be related to emotionally arousing events (for review, see Cahill and McGaugh, 1998). The investigation and description of structures and processes that are functionally correlated may illuminate interneuronal mechanisms required for long-lasting plastic changes and the formation of declarative memory.

REFERENCES

- Akirav I, Richter-Levin G (1999) Biphasic modulation of hippocampal plasticity by behavioral stress and basolateral amygdala stimulation in the rat. *J Neurosci* 19:10530–10535.
- Balschun D, Seidenbecher T, Reymann KG (1997) Behavior-reinforced LTP in the freely moving rat: a cellular model of associative memory. *Soc Neurosci Abstr* 23:1939.
- Barnes CA, Jung MW, McNaughton BL, Korol DL, Andreasson K, Worley PF (1994) LTP saturation and spatial learning disruption: effects of task variables and saturation levels. *J Neurosci* 14:5793–5806.
- Bergles DE, Doze VA, Madison DV, Smith SJ (1996) Excitatory actions of norepinephrine on multiple classes of hippocampal CA1 interneurons. *J Neurosci* 16:572–585.
- Bevilaqua L, Ardenghi P, Schröder N, Bromberg E, Schmitz PK, Schaeffer E, Quevedo J, Bianchin M, Walz R, Medina JH, Izquierdo I (1997) Drugs acting upon the cyclic adenosine monophosphate protein kinase A signalling pathway modulate memory consolidation when given late after training into rat hippocampus but not amygdala. *Behav Pharmacol* 8:331–338.
- Bianchin M, Souza TME, Medina JH, Izquierdo I (1999) The amygdala is involved in the modulation of long-term memory, but not in working or short-term memory. *Neurobiol Learn Mem* 71:127–131.
- Bliss TV, Lomo T (1973) Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol (Lond)* 232:331–356.
- Bliss TVP, Goddard GV, Riives M (1983) Reduction of long-term potentiation in the dentate gyrus of the rat following selective depletion of monoamines. *J Physiol (Lond)* 334:475–491.
- Brinton RE, McEwen BS (1989) Vasopressin neuromodulation in the hippocampus. *J Neurosci* 9:752–759.
- Cahill L, McGaugh JL (1998) Mechanisms of emotional arousal and lasting declarative memory. *Trends Neurosci* 21:294–299.
- Cahill L, Babinsky R, Markowitsch HJ, McGaugh JL (1995) The amygdala and emotional memory. *Nature* 377:295–296.
- Coussens CM, Teyler TJ (1996) Protein kinase and phosphatase activity regulate the form of synaptic plasticity expressed. *Synapse* 24:97–103.
- Dunwiddie TV, Roberson NL, Worth T (1982) Modulation of long-term potentiation: effects of adrenergic and neuroleptic drugs. *Pharmacol Biochem Behav* 17:1257–1264.
- Ferry B, Roozendaal B, McGaugh JL (1999) Role of norepinephrine in mediating stress hormone regulation of long-term memory storage: a critical involvement of the amygdala. *Biol Psychiatry* 46:1140–1152.
- Frey U (1997) Cellular mechanisms of long-term potentiation: late maintenance. In: *Neural network models of cognition: biobehavioral foundations* (Donahoe JW, Dorsel VP, eds), pp 105–128. Amsterdam: Elsevier.
- Frey U, Morris RGM (1997) Synaptic tagging and long-term potentiation. *Nature* 385:533–536.
- Frey U, Morris RGM (1998a) Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 21:181–188.
- Frey U, Morris RGM (1998b) Weak before strong: dissociating synaptic tagging and plasticity-factor accounts of late-LTP. *Neuropharmacology* 37:545–552.
- Frey U, Krug M, Reymann KG, Matthies H (1988) Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Res* 452:57–65.
- Frey U, Huang Y-Y, Kandel ER (1993) Effects of cAMP simulate a late stage of LTP in hippocampal CA1 neurons. *Science* 260:1661–1664.
- Frey U, Frey S, Schollmeier F, Krug M (1996) Influence of actinomycin D, a RNA synthesis inhibitor, on long-term potentiation in rat hippocampal neurons in vivo and in vitro. *J Physiol (Lond)* 490:703–711.
- Gray R, Johnston D (1987) Noradrenaline and beta-adrenoceptor agonists increase activity of voltage-dependent calcium channels in hippocampal neurons. *Nature* 327:620–622.
- Ikegaya Y, Saito H, Abe K (1994) Attenuated hippocampal long-term potentiation in basolateral amygdala-lesioned rats. *Brain Res* 656:157–164.
- Ikegaya Y, Saito H, Abe K (1995a) High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 22:203–207.
- Ikegaya Y, Saito H, Abe K (1995b) Requirement of basolateral amygdala neuron activity for the induction of long-term potentiation in the dentate gyrus in vivo. *Brain Res* 671:351–354.
- Ikegaya Y, Nakanishi K, Saito H, Abe K (1997) Amygdala b-noradrenergic influence on hippocampal long-term potentiation in vivo. *NeuroReport* 8:3143–3146.
- Izquierdo I, Medina JH (1997) Memory formation: the sequence of biochemical events in the hippocampus and its connection to activity in other brain structures. *Neurobiol Learn Mem* 68:285–316.
- Izquierdo I, Medina JH, Vianna MRM, Izquierdo LA, Barros DM (1999) Separate mechanisms for short- and long-term memory. *Behav Brain Res* 103:1–11.
- Krug M, Chepkova AN, Geyer C, Ott T (1983) Aminergic blockade modulates long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 11:1–6.
- Krug M, Lössner B, Ott T (1984) Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 13:39–42.
- LeDoux JE (1993) Emotional memory systems in the brain. *Behav Brain Res* 58:69–79.
- Malisch R, Ott T (1982) Rhythmical slow wave electroencephalographic activity elicited by hippocampal injection of muscarinic agents in the rat. *Neurosci Lett* 28:113–118.
- Moser EI, Krobort KA, Moser MB, Morris RGM (1998) Impaired spatial learning after saturation of long-term potentiation. *Science* 281:2038–2042.
- Packard MG, Cahill L, McGaugh JL (1994) Amygdala modulation of hippocampal-dependent and caudate nucleus-dependent memory processes. *Proc Natl Acad Sci USA* 91:8477–8481.
- Paxinos G, Watson C (1998) The rat brain in stereotaxic coordinates, Ed. 4. San Diego: Academic.
- Pikkarainen M, Rönkkö S, Savander V, Insausti R, Pitkänen A (1999) Projections from the lateral, basal, and accessory basal nuclei of the amygdala to the hippocampal formation in rat. *J Comp Neurol* 403:229–260.
- Roozendaal B, Nguyen BT, Power AE, McGaugh JL (1999) Basolateral amygdala noradrenergic influence enables enhancement of memory consolidation induced by hippocampal glucocorticoid receptor activation. *Proc Natl Acad Sci USA* 96:11642–11647.
- Seidenbecher T, Reymann KG, Balschun D (1997) A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc Natl Acad Sci USA* 94:1494–1499.
- Stanton PK, Heinemann U (1986) Norepinephrine enhances stimulus-evoked calcium and potassium concentration changes in dentate granule cell layer. *Neurosci Lett* 67:233–238.
- Stanton PK, Sarvey JM (1985) Depletion of norepinephrine, but not serotonin, reduces long-term potentiation in the dentate gyrus of rat hippocampal slices. *J Neurosci* 5:2169–2176.

Capítulo 4: La lesión de la fimbria-fornix afecta el reforzamiento de la plasticidad sináptica duradera inducido por estimulación de la amígdala

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La demostración del efecto reforzador de la estimulación de la amígdala basolateral sobre la LTP en el giro dentado plantea una interrogante acerca de cuáles estructuras pueden mediar dicho efecto, teniendo en cuenta que no se conocen proyecciones anatómicas directas entre ambas estructuras.

Existen dos posibles rutas principales: una a través de la proyección colinérgica del septo medial hacia el cuál la amígdala envía fibras excitadoras. Otra, por la vía de la corteza entorrinal y su proyección excitadora glutamatérgica a la formación del hipocampo.

El efecto farmacológico de bloqueo de la acción reforzadora de la amígdala por antagonistas de receptores muscarínicos, es un argumento a favor de una mediación por la vía septal. Nuestra hipótesis en este trabajo era la siguiente:

Si la proyección amígdala- septum-giro dentado es parte esencial del sistema neural implicado en el reforzamiento por estimulación, entonces la interrupción de esta vía por lesión de la fimbria-fornix debe bloquear dicho efecto.

Este trabajo a diferencia del resto, se realizó en animales anestesiados. La estimulación eléctrica de la amígdala basolateral se aplicó 15 minutos antes de inducir la LTP en la sinapsis vía perforante-giro dentado. En los animales con lesión de fimbria-fornix la inducción de la LTP no se vio alterada, pero si su mantenimiento, lo cuál apoya la hipótesis planteada.

Este resultado demuestra la importancia de la proyección fimbria-fornix como mediadora en el reforzamiento de procesos neuroplásticos por estimulación de la amígdala. Aunque

la proyección colinérgica septo-hipocampal alcanza el giro dentado por la fimbria-fornix, esto no significa que sea esa, y mucho menos solo esa, la proyección implicada en el efecto estudiado. A través de la fimbria-fornix llegan al giro dentado muchas otras fibras de origen subcortical, como la noradrenérgicas del locus coeruleus o serotoninérgicas del núcleo del rafe.

El resultado tampoco excluye que la vía alternativa amígdala-corteza entorrinal-hipocampo tenga alguna participación en los procesos estudiados.

Short communication

Lesioning the fimbria–fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus

J. Jas^a, W. Almaguer^a, J.U. Frey^b, J. Bergado^{a,*}^a *Laboratory of Experimental Electrophysiology, International Center for Neurological Restoration (CIREN), Ave. 25 #15805, Playa 12100, La Habana, Cuba*^b *Leibniz Institute for Neurobiology, Brenneckestr. 6, 39120 Magdeburg, Germany*

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Abstract

We have recently shown that early-long-term potentiation (LTP) in the dentate gyrus can be reinforced into late LTP by stimulation of the basolateral nucleus of the amygdala [Frey et al., submitted for publication]. The pathways and mechanisms for such interactions are unclear, considering that no direct projection from the amygdala to the dentate gyrus is known. To ascertain the possible mediation of the septo-hippocampal projection we have transected the fimbria–fornix (FF) fiber system in young adult (2 months) male rats. The electrophysiological evaluation a week later showed that the lesion does not modify the effects of pre-stimulation of the basolateral amygdala (BLA) on the induction of LTP at the perforant pathway (PP)-granule cells synapses, but impairs its maintenance 1 h later. This suggests that two different pathways might mediate different aspects of the amygdala–hippocampal interactions. One seemed to be anatomically independent from the FF and might influence LTP induction; while the second, probably through the septo-hippocampal fornical projection appeared important for LTP maintenance. © 2000 Elsevier Science B.V. All rights reserved.

Keywords: Synaptic plasticity; Memory; Hippocampus; Limbic system; Rat

The amygdala and the hippocampus are limbic structures, both involved in memory functions [15]. While the hippocampus is considered a key structure for cognitive processes [24], the amygdala appears primarily related to emotions [18]. A plausible hypothesis is that amygdala acts on memory mediating the reinforcing effects of motivational/emotional stimuli temporarily associated with learning [23]. However, the cellular mechanisms, which may sustain such interactions, are poorly understood.

Hippocampal long-term potentiation (LTP) has been considered a cellular correlate of memory at synaptic level [4,16,22], and there are some evidence that amygdala might contribute to the induction of LTP in the hippocampal formation [11,12,14,27]. Behavioral stimuli with a strong motivational/emotional component have shown able to improve the maintenance of LTP [25], suggesting that

the amygdala influence might also extend to later LTP phases.

Recently, we have shown that induction of an early-LTP, with a duration of about 4 h, can be converted into a protein-synthesis-dependent late-LTP if the basolateral amygdala (BLA) was stimulated within a distinct time window during LTP-induction (Frey et al., submitted for publication).

However, the anatomical pathways through which such functional interaction might occur, are unclear considering that no direct communication between the amygdala and the hippocampus [13,19] has yet been described. Two alternative pathways appear as candidates to mediate the proposed interaction: the entorhinal cortex–perforant pathway and the septum–fimbria–fornix system. There are both anatomical and electrophysiological evidence of a monosynaptic projection from the amygdala to each of these relay structures [6,7,19], but which of them could be responsible for mediating the BLA-induced reinforcement of LTP remains to be established.

* Corresponding author. Fax: +53-7-332420; e-mail: bergado@neubas.sld.cu

We have tried to approach this question using the fimbria–fornix (FF) lesion, a paradigm widely used as model for disconnecting the septal projections to the hippocampus [5], to determine whether the interruption of this pathway modifies the effects of BLA on hippocampal LTP.

Lesion of the FF was carried out on 2-month-old male Sprague–Dawley rats at the beginning of the experiments. Under chloral hydrate narcosis (420 mg/kg, i.p.) a stereotactically guided bilateral knife transection at the coordinates AP: -1.4 mm, ML: ± 0.8 to ± 3.1 mm and DV: 5.0 mm was carried out. Control animals received the same treatment but no knife was inserted and no transection was performed. This procedure significantly reduce spatial learning in the Morris water maze and eliminates completely the septal cholinergic innervation to the hippocampus, as shown by immunohistochemistry for choline esterase [2]. Though all the electrophysiological studies were done in the right hemisphere, lesions were made bilaterally to prevent recovery by sprouting of the contralateral fibers.

One week after surgery, the animals were anaesthetized again and fixed at the stereotactic frame (David Kopf, Tujunga). One monopolar recording electrode was placed in the hilar region of the dentate gyrus, while stimulating electrodes were placed ipsilaterally on the perforant pathway (PP) and the BLA. Signals were filtered between 1.5 and 3000 Hz using a VC-11 memory oscilloscope and averaged using a QC-111J Averager. Stimuli were produced by a SEN-3301 via a SS-104J isolation unit (all from Nihon-Kohden, Japan). Once a convenient location of the electrodes was achieved, input/output curves were determined for both inputs. On the PP curve, a stimulus intensity producing 40% of the maximal population spike was selected for test recording and tetanization. On the BLA curve, an intensity able to evoke a potential of about 1 mV was determined for tetanizing this input. Three control recordings were made before the first tetanization (baseline values); and test recordings were made at different time points after tetanization. Each record consisted of four consecutive responses to square pulses (0.1 ms) averaged at 0.1 Hz. On the test recordings, the amplitude of the population spike (P) was measured (Fig. 1A) and expressed as percent of the baseline values.

A tetanus consisted of three bursts of 15 impulses at 200 Hz with 10-s interburst interval. In the main experiment, a group of lesioned animals (LBLA, $n = 7$) and a group of control rats (CBLA, $n = 9$) were first tetanized in the BLA ($t = -15$) followed by tetanization of the PP ($t = 0$). Test recordings were made in the interval between the two tetanizations: 10 min and immediately before the second tetanization, as well as 2, 5, 15, 30 and 60 min after PP tetanization. In control experiments, performed in independent groups of rats, single or double PP tetanization were applied to control and lesioned animals. In the case of double PP tetanization, the time schedule for tetanization and recordings was the same as described for

the main experiment. When single tetanization of the PP was used, test recordings were made 2, 5, 15, 30 and 60 min after tetanization.

Transection of the FF does not alter the main features of the BLA evoked potentials in the dentate gyrus. A low amplitude (3 mV maximum) monophasic negative wave of about 20 ms latency was the response of the dentate granule cell population to single stimuli applied to the BLA, as it has been previously described [13,26]. Control experiments using single or double tetanization of the PP showed no difference in the amplitude or duration of LTP between lesioned and control animals (data not shown). This is consistent with previous findings showing that lesioning the FF does not impair the potentiation of the population spike [3].

In the groups in which, the BLA was tetanized before the PP, a small but significant (Wilcoxon T -test, $p < 0.05$) enhancement of the population spike develops after stimulating the amygdaloid nucleus (see Fig. 1B). The level of this increase in the population spike amplitude was similar in control and lesioned rats. The ability of the amygdala to induce LTP-like enhancements in the PP-granule cells synapses have been recently reported after stimulating the medial nuclear complex [1,11] though others failed to obtain such effect [13].

Tetanizing the PP 15 min later induces an initially similar LTP in both groups (Fig. 1C). However, a significant difference between lesioned and control groups appeared after 60 min. The level of potentiation at that time point was significantly reduced in lesioned rats when compared with control rats (Fig. 1D). Values in the lesioned rats were not significantly different from baseline values after 1 h. These data support recent findings [1] describing a similar involvement of the central amygdala on LTP induced in the PP-granule cells synapses and the role of FF mediating later stages.

Summarizing our results, FF transection does not affect normal synaptic transmission nor early synaptic plasticity (such as short-term potentiation) at the PP synapses in the dentate gyrus *in vivo*. However, the maintenance of LTP at PP synapses is significantly reduced and even blocked 1 h after LTP induction when the FF afferents have been previously interrupted.

LTP has been classically considered a homosynaptic phenomenon. Recent results [8,9,20], however, indicate that the maintenance of LTP requires the synergistic activation of non-glutamatergic pathways. The influence of heterosynaptic modulating inputs on later stages of hippocampal LTP, such as shown by the lesion of the septo-hippocampal pathway, may support this hypothesis. We do not yet know which transmitter system is involved in the effect reported here. The FF is the major input of subcortical afferents to the hippocampus. Noradrenergic, serotonergic and dopaminergic fibers arising from brain stem nuclei and acetylcholine from the septal nuclei are within the major components [5]. We have recently shown that

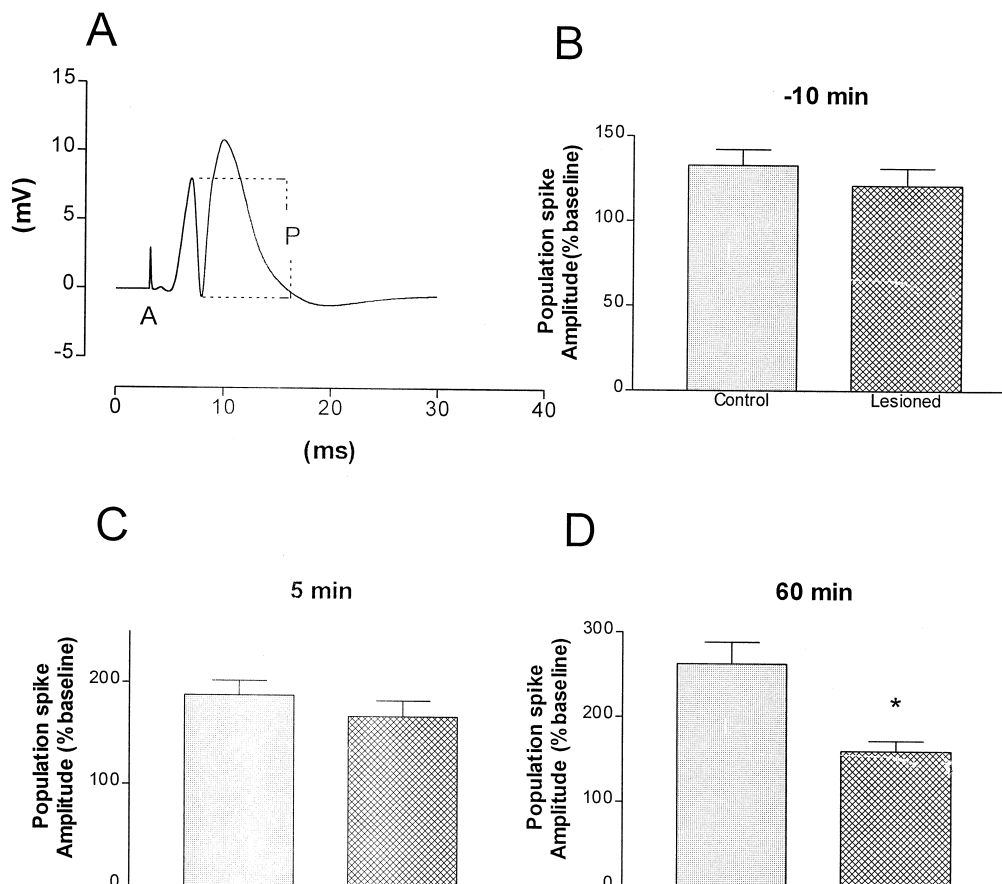


Fig. 1. (A) A test potential recorded in the dentate gyrus after stimulation of the PP. A: stimulus artefact; P: amplitude of the population spike. (B) Relative amplitude of the population spike after tetanizing the BLA to the baseline (%). The record was made 5 min after tetanizing the BLA, that is 10 min before tetanizing the PP. Control: non-lesioned animals. Lesioned: animals with bilateral transection of the FF. (C) Relative amplitude of the population spike 5 min after tetanizing the PP. (D) Relative amplitude of the population spike 60 min after tetanizing the PP, * $p < 0.05$ Mann–Whitney *U*-test.

the conversion of an early-LTP into a late-LTP by stimulation of the BLA is greatly influenced by noradrenergic and cholinergic, but not dopaminergic inputs (Frey et al., submitted for publication). This modulating inputs, we hypothesize, are required to regulate protein synthesis necessary for late-LTP to occur [17,21]. Glutamatergic inputs set in motion a series of 'fast-acting plasticity processors' whose function is to induce and maintain LTP during its early, protein-synthesis independent stages. The synergistic action of glutamatergic and modulatory inputs may activate the long-lasting reinforcement of the tagged synapses [10] only when such associative event takes place within a distinct time window.

The preservation of early effects of BLA stimulation in FF lesioned rats suggests that an alternative pathway may mediate this interaction. A likely candidate is the entorhinal cortex, considering the physiological [26] and pharmacological evidence [1]. The existence of two independent pathways linking the amygdala and the hippocampus may allow the former to influence effectively the different steps involved in hippocampal LTP. Future studies should

ascertain the specific function of the septo-hippocampal and entorhinal inputs on long-lasting neuronal plasticity.

The present results and the proposed interpretation stress the functional relevance of the amygdala influencing plastic mechanisms in the hippocampus and contribute to a better understanding of its role mediating emotional/motivational modulation of memory storage.

References

- [1] K. Abe, K. Noguchi, H. Saito, Medial amygdala-induced spike potentiation in the rat dentate gyrus is dependent on *N*-methyl-D-aspartate receptors and subcortical afferents, *Neurosci. Lett.* 246 (1998) 85–88.
- [2] W. Almaguer Melian, J. Jas García, L. Francis Turner, I. Antunez Potashkina, J. Bergado Rosado, Estudio comparativo de la lesión de fimbria–fornix por aspiración y transección, *Rev. Neurol.* 29 (1999) 704–709.
- [3] J.A. Bergado, H. Moreno, J. Soto, O. Castellano, L. Castillo, Septal fetal tissue transplants restore long-term potentiation in the dentate

- gyrus of fimbria–formix-lesioned rats, *J. Neural Transplant. Plast.* 6 (1997) 31–40.
- [4] T.V.P. Bliss, G.L. Collingridge, A synaptic model of memory: long-term potentiation in the hippocampus, *Nature* 361 (1993) 31–39.
 - [5] J.-C. Cassel, E. Duconseille, H. Jeltsch, B. Will, The fimbria–formix/cingular bundle pathways: a review of neurochemical and behavioural approaches using lesions and transplantation techniques, *Prog. Neurobiol.* 51 (1997) 663–716.
 - [6] A. Desmedt, R. Garcia, R. Jaffard, Differential modulation of changes in hippocampal–septal synaptic excitability by the amygdala as a function of either elemental or contextual fear conditioning in mice, *J. Neurosci.* 18 (1998) 480–487.
 - [7] E.E. Wong, E.L. Derian, X.-H. Chen, N.L. Nowlin-Finch, L.A. Brothers, Neurophysiology of limbic system pathways in the rat: projections from the amygdala to the entorhinal cortex, *Brain Res.* 370 (1986) 273–284.
 - [8] U. Frey, Y.-Y. Huang, E.R. Kandel, Effects of cAMP simulate a late stage of LTP in hippocampal CA1 neurons, *Science* 260 (1993) 1661–1664.
 - [9] U. Frey, H. Matthies, K.G. Reymann, The effect of dopaminergic D1 receptor blockade during tetanization on the expression of long-term potentiation in the rat CA1 region in vitro, *Neurosci. Lett.* 129 (1991) 111–114.
 - [10] U. Frey, R.G.M. Morris, Synaptic tagging and long-term potentiation, *Nature* 385 (1997) 533–536.
 - [11] Y. Ikegaya, H. Saito, K. Abe, Amygdala *N*-methyl-D-aspartate receptors participate in the induction of long-term potentiation in the dentate gyrus in vivo, *Neurosci. Lett.* 192 (1995) 193–196.
 - [12] Y. Ikegaya, H. Saito, K. Abe, High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo, *Neurosci. Res.* 22 (1995) 203–207.
 - [13] Y. Ikegaya, H. Saito, K. Abe, Dentate gyrus field potentials evoked by stimulation of the basolateral amygdaloid nucleus in anesthetized rats, *Brain Res.* 718 (1996) 53–60.
 - [14] Y. Ikegaya, H. Saito, K. Abe, The basomedial and basolateral amygdaloid nuclei contribute to the induction of long-term potentiation in the dentate gyrus in vivo, *Eur. J. Neurosci.* 8 (1996) 1833–1839.
 - [15] I. Izquierdo, J.A. Quirrellfeldt, M.S. Zanatta, J. Quevedo, E. Schaeffer, P.K. Schmitz, J.H. Medina, Sequential role of hippocampus and amygdala, entorhinal cortex and parietal cortex in formation and retrieval of memory for inhibitory avoidance in rats, *Eur. J. Neurosci.* 9 (1997) 786–793.
 - [16] M. Krug, J. Bergado, H. Ruethrich, Long-term potentiation and postconditioning potentiation — the same mechanism?, *Biomed. Biochim. Acta* 49 (1990) 273–279.
 - [17] M. Krug, B. Lössner, T. Ott, Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats, *Brain Res. Bull.* 13 (1984) 39–42.
 - [18] J.E. LeDoux, The amygdala: contribution to fear and stress, *Sem. Neurosci.* 6 (1994) 237.
 - [19] S. Maren, Synaptic transmission and plasticity in the amygdala — an emerging physiology of fear conditioning circuits, *Mol. Neurobiol.* 13 (1996) 1–22.
 - [20] H. Matthies, Neuronale Grundlagen der Gedächtnisbildung, in: F. Klix, H. Spada (Eds.), *Enzyklopädie der Psychologie*, Vol. 6, Göttingen, Hogrefe Verlag für Psychologie. Kognition, 1998, pp. 15–42.
 - [21] H. Matthies, U. Frey, M. Krug, W. Pohle, K.G. Reymann, H. Rüthrich, Long-term synaptic potentiation and macromolecular changes in memory formation, *Ergeb. Exp. Med.* 51 (1990) 88–94.
 - [22] H. Matthies, H. Ruethrich, T. Ott, H.K. Matthies, R. Matthies, Low frequency perforant path stimulation as a conditioned stimulus demonstrates correlations between long-term synaptic potentiation and learning, *Physiol. Behav.* 36 (1986) 811–821.
 - [23] J.L. McGaugh, L. Cahill, B. Roozendaal, Involvement of the amygdala in memory storage: interaction with other brain systems, *Proc. Natl. Acad. Sci. U. S. A.* 93 (1996) 13508–13514.
 - [24] B. Milner, L.R. Squire, E.R. Kandel, Cognitive neuroscience and the study of memory, *Neuron* 20 (1998) 445–468.
 - [25] T. Seidenbecher, K.G. Reymann, D. Balschun, A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli, *Proc. Natl. Acad. Sci. U. S. A.* 94 (1997) 1494–1499.
 - [26] S.R. Thomas, S.Y. Assaf, S.D. Iversen, Amygdaloid complex modulates neurotransmission from the entorhinal cortex to the dentate gyrus of the rat, *Brain Res.* 307 (1984) 363–365.
 - [27] Y. Watanabe, Y. Ikegaya, H. Saito, K. Abe, Roles of GABA_A, NMDA and muscarinic receptors in induction of long-term potentiation in the medial and lateral amygdala in vitro, *Neurosci. Res.* 21 (1995) 317–322.

Capítulo 5: La desaferentación subcortical afecta el reforzamiento conductual de la potenciación sináptica duradera

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La hipótesis que da origen a este trabajo se deriva directamente de la anterior.

Si el sistema de la fimbria-fornix es un mediador importante del efecto reforzador de la amígdala sobre la LTP en el giro dentado, y la amígdala es un componente esencial del mecanismo de reforzamiento conductual, entonces el sistema fimbria-fornix debe ser también mediador del reforzamiento conductual.

Para comprobarla se empleó el modelo de reforzamiento conductual en animales privados de agua a los cuales se les había lesionado la fimbria-fornix una semana antes.

Los grupos de control mostraron que esta lesión no afectó los valores basales del componente evaluado en los potenciales de campo y tampoco afectó el curso temporal de la LTP sin reforzamiento (fig. 3B). Sin embargo el efecto de reforzamiento conductual si fue abolido completamente por la lesión de la fimbria-fornix.

Si bien este resultado demuestra que esta proyección es importante, y confirman la hipótesis, deben tenerse en cuenta las mismas limitaciones y la misma cautela que en el análisis del resultado precedente, para no extrapolar su validez injustificadamente.

SUBCORTICAL DEAFFERENTATION IMPAIRS BEHAVIORAL REINFORCEMENT OF LONG-TERM POTENTIATION IN THE DENTATE GYRUS OF FREELY MOVING RATS

W. ALMAGUER-MELIAN,^a J. C. ROSILLO,^a J. U. FREY^b
AND J. A. BERGADO^{a*}

^aCentro Internacional de Restauración Neurológica, Ave. 25 # 15805, Cubanacán, Playa 11300, Ciudad de La Habana, Cuba

^bLeibniz Institute for Neurobiology, D-39118 Magdeburg, Germany

Abstract—Long-term potentiation is a form of neural functional plasticity which has been related with memory formation and recovery of function after brain injury. Previous studies have shown that a transient early-long-term potentiation can be prolonged by direct stimulation of distinct brain areas, or behavioral stimuli with a high motivational content. The basolateral amygdala and other subcortical structures, like the medial septum and the locus coeruleus, are involved in mediating the reinforcing effect. We have previously shown that the lesion of the fimbria-fornix—the main entrance of subcortical afferents to the hippocampus—abolishes the reinforcing basolateral amygdala-effects on long-term potentiation in the dentate gyrus *in vivo*. It remains to be investigated, however, if such subcortical afferents may also be important for behavioral reinforcement of long-term potentiation. Young-adult (8 weeks) Sprague–Dawley male rats were fimbria-fornix-transected under anesthesia, and electrodes were implanted at the dentate gyrus and the perforant path. One week after surgery the freely moving animals were studied. Fimbria-fornix-lesion reduced the ability of the animals to develop long-term potentiation when a short pulse duration was used for tetanization (0.1 ms per half-wave of a biphasic stimulus), whereas increasing the pulse duration to 0.2 ms per half-wave during tetanization resulted in a transient early-long-term potentiation lasting about 4 h in the lesioned animals, comparable to that obtained in non-lesioned or sham-operated control rats. In water-deprived (24 h) control animals, i.e. in non-lesioned and sham-operated rats, early-long-term potentiation could be behaviorally reinforced by drinking 15 min after tetanization. However, in fimbria-fornix-lesioned animals long-term potentiation-reinforcement by drinking was not detected. This result indicates that the effect of behavioral–motivational stimuli to reinforce long-term potentiation is mediated by subcortical, heterosynaptic afferents. © 2005 Published by Elsevier Ltd on behalf of IBRO.

Key words: synaptic plasticity, behavioral reinforcement, fimbria-fornix, dentate gyrus, basolateral amygdala, rat.

*Corresponding author. Tel: +53-7-273-6844; fax: +53-7-273-2420. E-mail address: bergado@neuro.ciren.cu (J. A. Bergado).

Abbreviations: ACh, acetylcholine; AChE, acetyl cholinesterase; BLA, basolateral portion of the amygdala; BR, behavioral reinforcement; DG, dentate gyrus; EC, entorhinal cortex; E-LTP, early long-term potentiation; FF, fimbria-fornix; L-LTP, late long-term potentiation; LTP, long-term potentiation; NE, norepinephrine; NMDA, *N*-methyl-D-aspartate; PSA, population spike amplitude.

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Long-term potentiation (LTP) is a form of functional plasticity expressed as an increase of synaptic efficacy following brief, high frequency stimulation (Bliss and Gardner-Medwin, 1973; Bliss and Lomo, 1973). The phenomenon is not unitary, but consists of temporarily sequenced, partially overlapping phases. An initial, early-long-term potentiation phase (E-LTP) lasting less than 4 h is followed by a late-long-term potentiation (L-LTP) which depends on protein synthesis (Krug et al., 1984; Frey et al., 1988; Reymann et al., 1988).

It has been shown that an E-LTP induced by mild stimulation patterns in the dentate gyrus (DG) of freely moving rats, can be converted into an L-LTP by the application of behavioral stimuli with a high motivational value within a limited time window before or after LTP-induction (Seidenbecher et al., 1997; Straube et al., 2003b). This effect seems to depend on the ability of such stimuli to activate modulatory brain regions projecting to the DG, resulting in the synthesis of plasticity-related proteins required for L-LTP to occur. This is documented by the fact that the application of anisomycin, a protein synthesis inhibitor, prevented behaviorally-induced LTP-reinforcement (Bergado et al., 2003; Straube et al., 2003a).

Similarly, direct electrical stimulation of the basolateral portion of the amygdala (BLA) is also able to reinforce E-LTP into L-LTP (Frey et al., 2001) suggesting a role for this limbic structure during behavioral reinforcement (BR). This effect was also dependent on protein synthesis (Frey et al., 2001) and the temporal or permanent inactivation of the amygdala impaired behavioral LTP-reinforcement (Almaguer-Melian et al., 2003), strengthening the hypothesis that the BLA might be part of the BR system.

The BLA might project to the DG via the entorhinal cortex (EC) or the fimbria-fornix (FF) (Canning and Leung, 1997; Thomas et al., 1984; Maren, 1996; Maren and Quirk, 2004). The EC sends an excitatory, glutamatergic projection to the DG (Deller and Frotscher, 1997; Vizi and Kiss, 1998). The FF afferents to the DG are of subcortical origin, and include a GABAergic and a cholinergic component from septal origin, as well as noradrenergic fibers arising from the locus coeruleus (Adelmann et al., 1996; Cassel et al., 1997; Vizi and Kiss, 1998). We have previously shown that interrupting the FF abolishes the reinforcing effects of stimulating the BLA on LTP (Jas et al., 2000), but it is not known whether that lesion will have similar effects on BR.

Here, we present evidence that lesioning the FF, i.e. depriving the DG from subcortical innervation, abolishes BR of LTP.

EXPERIMENTAL PROCEDURES

Animals

Male adult Sprague–Dawley rats (obtained from CENPALAB, Havana, Cuba) weighing 250–270 g during electrode implantation were housed individually after surgery in plastic, translucent cages. Water and food (Rat Chow, CENPALAB) remained *ad libitum* through the experiment, except for the series investigating the effect of BR of LTP, in which rats were water-deprived for 24 h. Animals were handled according to the Cuban Regulations for Using Laboratory Animals and the U.S. National Institutes of Health guidelines for the use of laboratory animals. All efforts were made to reduce the number of animals and their suffering.

Surgery

Surgery was performed under chloral hydrate narcosis (420 mg/kg, intraperitoneally) according to the procedure described elsewhere (Almaguer-Melian et al., 2005b). The animals were mounted on a stereotactic frame (David Kopf Inst., Saint Louis, MO, USA) and the skull was exposed after cutting the skin.

Lesioning of the FF

A bilateral window was opened at the skull at coordinates (AP: -1.4 mm, ML: ± 0.5 to ± 5.2 mm). A reduced #11 surgical blade was lowered at 15° (with respect to the vertical direction) and a bilateral knife transection was carried out at the coordinates AP: -1.4 mm, ML: ± 0.8 to ± 3.1 mm, and DV: 5.0 mm. Though all the electrophysiological studies were done in the right hemisphere, lesions were made bilaterally to prevent recovery by sprouting of the contralateral fibers. Sham-operated animals received the same treatment, but no knife was inserted and no transection was performed. Control animals were not operated.

Electrode implantation

Stainless steel, Teflon-isolated monopolar electrodes for recording were implanted in the hilus of the DG (coordinates AP: -3.8 mm, ML: 2.0 mm, DV: -3.5 mm from bregma) and bipolar electrodes of the same wire for stimulation at the angular bundle of the PP (coordinates AP: -7.5 mm, ML: 4.0 mm, DV: -3.7 mm). Miniscrews were attached in the right frontal and left parietal bones to serve as indifferent and ground electrode respectively.

Electrophysiology

One week after surgery the animals were habituated to the test chamber for at least 4 h before obtaining an input–output curve on which the stimulus intensity was determined for each animal at 40% of the maximal population spike amplitude (PSA). The test chamber consisted of individual plastic concave boxes, in which the animals could move freely while remain connected to the amplifier (AB 621G, Nihon Kohden, Japan) and stimulator (AM Systems 2100, AM Systems, Sequim, WA, USA). The filtered signals (1–5000 Hz) were digitized (AD converter 1401+, CED, Cambridge, Great Britain) fed into a PC and averaged and measured by a special computer program (Intracell, Magdeburg, Germany). Lesioned (lesion) sham-operated (sham) and control animals were then randomly assigned to one of the experimental protocols. In the LTP protocol a weak tetanus (3×15 impulses at 200 Hz) was used to induce an E-LTP. In the initial design we have planned to use 0.1 ms pulse duration per half-wave for LTP induction, but we noticed that the level of LTP induced was too low

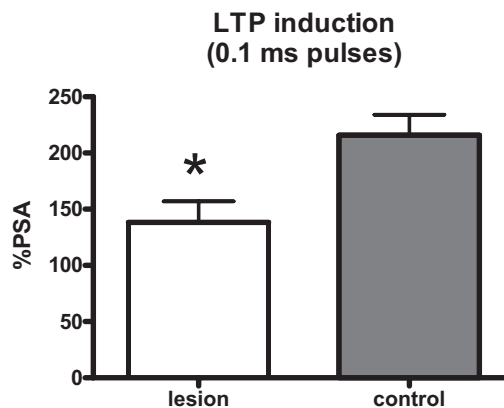


Fig. 1. Level of potentiation reached by FF-lesioned animals (lesion, $n=9$) using 0.1 ms pulse duration for the high frequency stimulation compared with a “historical” sample of non-lesioned animals (control, $n=24$). The percentual potentiation of the PSA (%PSA) is shown 5 min after LTP induction.

in comparison to the historical average in our laboratory (see Fig. 1). Therefore, we have changed to 0.2 ms pulse duration, re-started the group, and used it throughout the experiment. For the BR protocol animals were water deprived for 24 h before inducing E-LTP using the same stimulus pattern (0.2 ms). Fifteen minutes after LTP induction, water was made available until the end of the recording session. Control experiments, performed earlier in our laboratory, revealed no effects of water deprivation on test potentials recorded in a control group without tetanization [19].

Test recordings

Test recordings were made before and after induction of LTP. Twelve recordings were made during one hour before LTP. After LTP induction, test recordings were made 5 min after induction, and from 30 min on, up to 6 h, new test recordings were made every 15 min, as well as 24 h later. Each test recording represents the averaged value of five consecutive evoked potentials recorded at a frequency of 0.1 Hz.

Histology

After finishing the experiments the animals were perfused under narcosis, and the brains sectioned, and stained for the enzyme acetyl cholinesterase (AChE), to confirm the efficacy of the lesioning procedure.

Statistical analysis

A three-way ANOVA with one repeated measures factor (LESION, STIMULATION and TIME) was used. When significant differences were found a post hoc test using the Tukey honest significant differences was performed. Differences were considered significant only when $P < 0.05$.

RESULTS

The histological examination confirmed a severe reduction of AChE in the hippocampus and the DG of lesioned animals, indicating that the lesion procedure was effective in provoking an intense subcortical denervation of the hippocampal formation (Fig. 2).

An initial ANOVA, considering the factors type of LESION (i.e. lesion, sham or controls), and the STIMULATION protocol (whether it includes BR or not) as independent

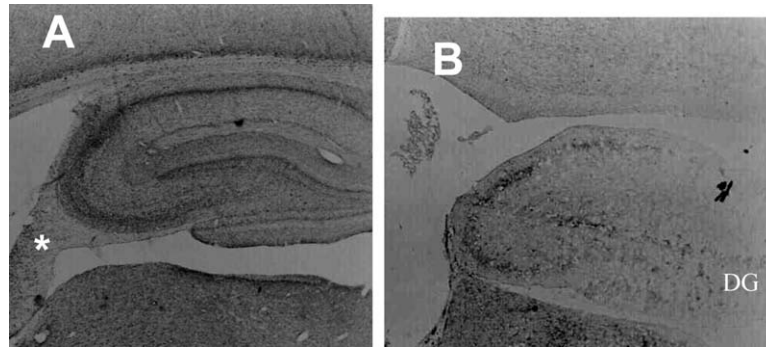


Fig. 2. Representative microphotographs of a control animal (A) and a FF-lesioned animal (B) stained for ACh esterase. Notice the intense reduction of staining in all subfields of the hippocampal formation including the DG in the lesioned animal. Notice also the absence of the fornix in B (marked with an asterisk in the control animal).

variables, with TIME as repeated measures factor dependent variable, confirmed significant differences due to LESION ($F_{2, 42}=3.91428$); the STIMULATION protocol ($F_{1, 42}=21.66280$); and TIME ($F_{7, 294}=33.19984$). A significant interaction between LESION and STIMULATION was also confirmed ($F_{2, 42}=6.98083$) as well as between LESION and TIME ($F_{14, 294}=2.50766$).

For the sake of simplicity in the presentation of results, we have separated both stimulation protocols in two graphs which are shown in the following paragraphs, with their corresponding ANOVA results.

Fig. 3 shows the results of the three lesion groups in the LTP stimulation protocol. A two-way ANOVA (LESION and TIME, repeated measures) showed no effect for LESION ($F_{2, 21}=0.46395$) a significant effect for TIME ($F_{7, 147}=32.11707$) and no interaction between factors ($F_{14, 147}=1.27327$) was also seen. The post hoc analysis for factor TIME showed that the records corresponding to 5 min and 1 h after LTP induction significantly differ from the rest. However, the value corresponding to 2 h do not differ from those of 3 and 4 h, but do differ from those taken from 5 h on.

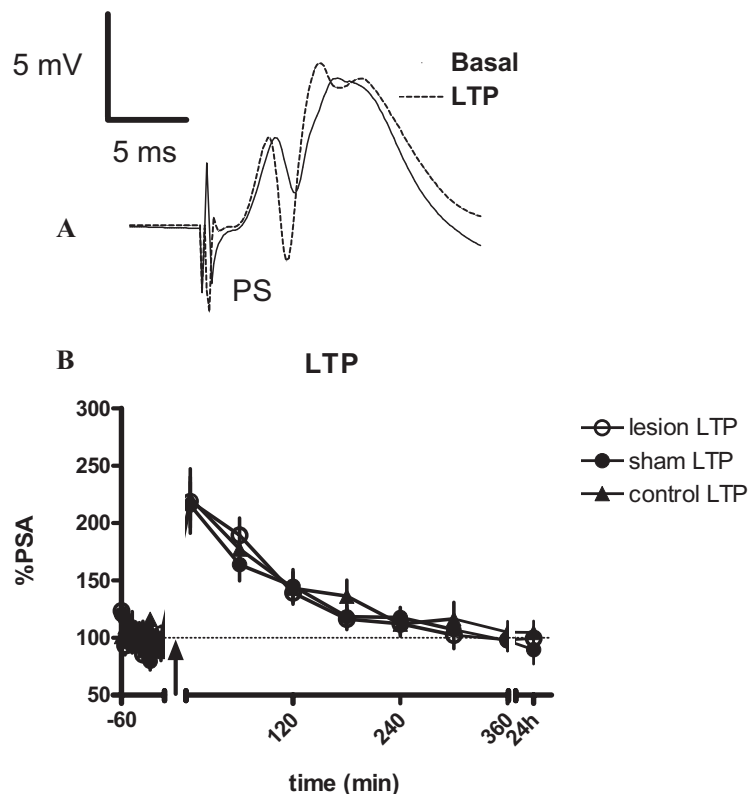


Fig. 3. (A) Representative recordings taken from one FF-lesioned animal 5 min before (basal: continuous line) and 5 min after (LTP: interrupted line) induction of LTP (PS: population spike). (B) Time course of LTP in FF-lesioned animals (lesion LTP, $n=9$), sham-operated (sham LTP, $n=10$) and control animals (control LTP, $n=10$). The ANOVA two way with repeated measures showed no differences due to the lesioning protocol. (The vertical arrow indicates the time of LTP induction.)

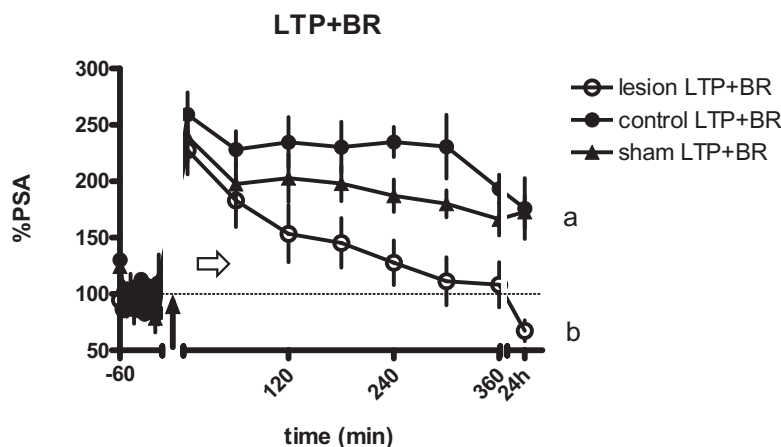


Fig. 4. Time course of LTP followed by BR (water access) 15 min later. The two way ANOVA with repeated measures confirmed differences among groups. FF-lesioned animals (lesion LTP+BR, $n=10$), sham-operated (sham LTP+BR, $n=10$) and control animals (control LTP+BR, $n=9$) are shown. The post hoc analysis showed no differences in the initial level of potentiation, but the reinforcing effect (a prolonged potentiation due to BR) was absent in lesioned animals. (The vertical arrow indicates the time of LTP induction; the open horizontal arrow indicates the time in which water was made available.)

These results confirm that the stimulation protocol induced a decaying E-LTP in all three groups of animals, and that the lesion of the FF induced no difference in the level and evolution of LTP.

Fig. 4 shows the results when behavioral motivational stimulus (drinking) was applied 15 min after inducing LTP with an identical train. A two-way ANOVA (LESION and TIME, repeated measures) showed significant effects for LESION ($F_{2, 21}=7.06803$) TIME ($F_{7, 147}=10.58999$) and the interaction between both ($F_{14, 147}=2.04525$). The post hoc analysis for the LESION $\times$ TIME interaction revealed no differences among groups during the first 2 h after LTP induction, confirming that a similar E-LTP was induced in the three groups irrespective of the fact that they were lesioned or not. However, after 3 h the lesioned animals differed significantly from the control ones, while from 4 h to 24 h they differed from both lesioned and sham-operated animals. These two groups (control and sham) do not show differences at any time point. This indicates that while the BR was able to transform the initial E-LTP into an L-LTP in animals bearing no or sham lesion, in those with a bilateral lesion of the FF such effect was abolished.

DISCUSSION

Our results confirm our hypothesis that the lesion of the FF should also abolish the reinforcement of LTP by motivational behavioral stimuli.

Application of the 0.2 ms pulse duration tetanization protocol resulted in the induction of a similar E-LTP in all groups. However, application of the weaker, 0.1 ms pulse duration tetanization protocol caused different plastic events in the different groups of animals. An early report about the effects of FF lesion on LTP showed an absence of potentiation in lesioned animals (Buzsáki and Gage, 1989). These authors employed 0.1 ms pulses at 200 Hz to induce LTP, but the number of impulses delivered was higher (eight trains of 20 pulses) than ours (three trains of

15 pulses). Similar results were reported later by a different group using the same frequency (Valjakka et al., 1991) although they do not mention the pulse duration employed. However, this impaired induction seems to be dependent on the stimulation protocol. We have studied LTP in FF-lesioned rats employing stronger stimuli (10 trains of 10 pulses at 400 Hz) and showed a normal induction of LTP of the population spike (but reduced in the slope of the EPSP) (Bergado et al., 1996). There is no clear interpretation for these results. It has been argued that a lower level of catecholamines might activate Ca^{2+} -dependent K channels, impairing thus the activation of *N*-methyl-D-aspartate (NMDA) receptors through high frequency stimulation. According to this, it is conceivable that a stronger stimulus (like a longer pulse) might overcome this deficit to reach a normal level of LTP.

Our LTP groups confirm this assumption. All three groups (lesion, sham and control) showed an identical level and time course of LTP when tetanized with the 0.2 ms protocol. This is consistent with the assumption that the stimulation pattern employed induced only an E-LTP (Seidenbecher et al., 1997; Frey et al., 2001).

The lesion of the FF had consequences on the mechanisms of induction of later phases of LTP (L-LTP) as demonstrated by our results combining high frequency stimulation and BR. The FF-lesioned animals develop an E-LTP which does not differ from the sham or control animals. However, while in two control groups the access to water prolonged LTP; in the FF-lesioned group LTP-reinforcement was completely absent. In fact the decay of LTP in these animals is identical to that of the groups in which no BR was applied. These data support the important role and the relevance of subcortical modulatory afferents for BR of the NMDA-dependent LTP induced by stimulation of the medial perforant pathway in the DG.

As L-LTP (Krug et al., 1984; Frey et al., 1988) and BR (Bergado et al., 2003) depend on protein synthesis, we

suggest to interpret our result in terms of the synaptic tagging hypothesis (Frey and Morris, 1997a,b). According to this, the induction of LTP sets a tag at the activated synapses which allows the recognition and insertion of the proteins required for a longer lasting enhancement of synaptic efficacy. BR might activate heterosynaptic afferents, releasing transmitter substances able to activate metabolic cascades leading to regulation of transcription, and therefore, the synthesis of plasticity proteins. The lack of reinforcement in the FF lesioned group, strongly suggests that these modulatory afferents reach the DG via the FF, and suggest further a subcortical origin for them.

Pharmacological studies have shown that propranolol, a β -adrenergic receptor antagonist, can effectively block BR (Seidenbecher et al., 1997; Straube et al., 2003a) and BLA-induced reinforcement (Frey et al., 2001), while atropine, a muscarinic receptor antagonist, only blocks the latter. Intraventricular administration of norepinephrine (NE) but not oxotremorine (a muscarinic agonist) reinforces LTP in a similar manner as after BR or BLA-stimulation (Almaguer-Melian et al., 2005b). The mentioned pharmacological studies lead to the hypothesis of an action of NE on the DG, converting an E-LTP into an L-LTP, while the possible role of acetylcholine (ACh) seemed not so clear with respect to the above mentioned forms of reinforcement. However, one should keep in mind that the i.c.v. administration opens the possibility that the effects of these substances may not necessarily take place within the DG. In a microdialysis study we have tried to solve this question studying changes in neurotransmitter release at the DG under BLA stimulation or BR. The results show an increase in the release of ACh and a reduction in NE under BLA-stimulation, while under BR, ACh and NE remained unchanged and a reduction in glutamate and GABA release was found (Almaguer-Melian et al., 2005a). While the large sampling time, and the low frequency of stimulation used, might have masked some short-term changes, this result suggests that effects of NE are indirect, acting perhaps on the amygdala to modulate its activity.

The amygdala appears as a key structure to mediate motivational effects on learning (Abe et al., 1998; Antoniadis and McDonald, 2000; Cahill, 1999; Hamann et al., 1999; Huff and Rudy, 2004; Izquierdo et al., 1997; McGaugh et al., 2002; McIntyre et al., 2003; Pare, 2003; Richter-Levin, 2004) and synaptic plasticity (Abe, 2001; Akirav and Richter-Levin, 1999, 2002; Almaguer-Melian and Bergado Rosado, 2002; Ikegaya et al., 1994, 1995), two processes that appear to be functionally linked. The lack of BR in FF-lesioned animals could be attributed to the interruption of one of the pathways through which, the BLA projects into the DG. However, other brain structures like the locus coeruleus (Harley et al., 1989; Berridge and Waterhouse, 2003; Kitchigina et al., 1997) sending a noradrenergic projection to the hippocampal formation via the FF should not be discarded. Experiments in progress, using topical application of substances within relevant limbic and forebrain structures, should contribute to a better understanding of the system and transmitters involved in the motivational modulation of LTP.

REFERENCES

- Abe K (2001) Modulation of hippocampal long-term potentiation by the amygdala: a synaptic mechanism linking emotion and memory. *Jpn J Pharmacol* 86:18–22.
- Abe K, Inokawa M, Kashiwagi A, Yanagihara T (1998) Amnesia after a discrete basal forebrain lesion. *J Neurol Neurosurg Psychiatry* 65:126–130.
- Adelmann G, Deller T, Frotscher M (1996) Organization of identified fiber tracts in the rat fimbria-fornix: an anterograde tracing and electron microscopic study. *Anat Embryol (Berl)* 193:481–493.
- Akirav I, Richter-Levin G (2002) Mechanisms of amygdala modulation of hippocampal plasticity. *J Neurosci* 22:9912–9921.
- Akirav I, Richter-Levin G (1999) Priming stimulation in the basolateral amygdala modulates synaptic plasticity in the rat dentate gyrus. *Neurosci Lett* 270:83–86.
- Almaguer-Melian W, Bergado Rosado JA (2002) Interactions between the hippocampus and the amygdala in synaptic plasticity processes. A key to understanding the relations between motivation and memory. *Rev Neurol* 35:586–593.
- Almaguer-Melian W, Cruz-Aguado R, De la Riva, Kendrick KM, Frey JU, Bergado J (2005a) Effect of LTP-reinforcing paradigms on neurotransmitter release in the dentate gyrus of young and aged rats. *Biochem Biophys Res Commun* 327:877–883.
- Almaguer-Melian W, Rojas-Reyes Y, Alvaré A, Rosillo JC, Frey JU, Bergado JA (2005b) Long-term potentiation in the dentate gyrus in freely moving rats is reinforced by intraventricular application of norepinephrine, but not oxotremorine. *Neurobiol Learn Mem* 83: 72–78.
- Almaguer-Melian W, Martínez-Martí L, Frey JU, Bergado JA (2003) The amygdala is part of the behavioral reinforcement system modulating long-term potentiation in rat hippocampus. *Neuroscience* 119:319–322.
- Antoniadis EA, McDonald RJ (2000) Amygdala, hippocampus and discriminative fear conditioning to context. *Behav Brain Res* 108: 1–19.
- Bergado JA, Almaguer-Melian W, Kostenko S, Frey S, Frey JU (2003) Behavioral reinforcement of long-term potentiation in rat dentate gyrus in vivo is protein synthesis-dependent. *Neurosci Lett* 351: 56–58.
- Bergado JA, Moreno H, Nuñez N (1996) Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biol Res* 29: 197–202.
- Berridge CW, Waterhouse BD (2003) The locus coeruleus-noradrenergic system: modulation of behavioral state and state-dependent cognitive processes. *Brain Res Brain Res Rev* 42:33–84.
- Bliss TV, Gardner-Medwin AR (1973) Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol (Lond)* 232:357–374.
- Bliss TV, Lomo T (1973) Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *J Physiol (Lond)* 232:331–356.
- Buzsáki G, Gage FH (1989) Absence of long-term potentiation in the subcortically deafferented dentate gyrus. *Brain Res* 484:94–101.
- Cahill L (1999) A neurobiological perspective on emotionally influenced, long-term memory. *Semin Clin Neuropsychiatry* 4:266–273.
- Canning KJ, Leung LS (1997) Lateral entorhinal, perirhinal, and amygdala-entorhinal transition projections to hippocampal CA1 and dentate gyrus in the rat: a current source density study. *Hippocampus* 7:643–655.
- Cassel J-C, Duconseille E, Jeltsch H, Will B (1997) The fimbria-fornix/cingulate bundle pathways: a review of neurochemical and behavioural approaches using lesions and transplantation techniques. *Prog Neurobiol* 51:663–716.
- Deller T, Frotscher M (1997) Lesion-induced plasticity of central neurons: sprouting of single fibers in the rat hippocampus after unilateral entorhinal cortex lesion. *Prog Neurobiol* 53:687–727.

- Frey S, Bergado JA, Seidenbecher T, Pape HC, Frey JU (2001) Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP. *J Neurosci* 21:3697–3703.
- Frey U, Krug M, Reymann KG, Matthies H (1988) Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Res* 452:57–65.
- Frey U, Morris RGM (1997a) Synaptic tagging and long-term potentiation. *Nature* 385:533–536.
- Frey U, Morris RGM (1997b) Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 21:181–188.
- Hamann SB, Ely TD, Grafton ST, Kilts CD (1999) Amygdala activity related to enhanced memory for pleasant and aversive stimuli. *Nat Neurosci* 2:289–293.
- Harley C, Milway JS, Lacaille JC (1989) Locus coeruleus potentiation of dentate gyrus responses: Evidence for two systems. *Brain Res Bull* 22:643–650.
- Huff NC, Rudy JW (2004) The amygdala modulates hippocampus-dependent context memory formation and stores cue-shock associations. *Behav Neurosci* 118:53–62.
- Ikegaya Y, Saito H, Abe K (1995) High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 22:203–207.
- Ikegaya Y, Saito H, Abe K (1994) Attenuated hippocampal long-term potentiation in basolateral amygdala-lesioned rats. *Brain Res* 656:157–164.
- Izquierdo I, Quirfeldt JA, Zanatta MS, Quevedo J, Schaeffer E, Schmitz PK, Medina JH (1997) Sequential role of hippocampus and amygdala, entorhinal cortex and parietal cortex in formation and retrieval of memory for inhibitory avoidance in rats. *Eur J Neurosci* 9:786–793.
- Jas J, Almaguer W, Frey JU, Bergado J (2000) Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res* 861:186–189.
- Kitchigina V, Vankov A, Harley C, Sara SJ (1997) Novelty-elicited, noradrenaline-dependent enhancement of excitability in the dentate gyrus. *Eur J Neurosci* 9:41–47.
- Krug M, Lössner B, Ott T (1984) Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 13:39–42.
- Maren S (1996) Synaptic transmission and plasticity in the amygdala: An emerging physiology of fear conditioning circuits. *Mol Neurobiol* 13:1–22.
- Maren S, Quirk GJ (2004) Neuronal signaling of fear memory. *Nat Rev Neurosci* 5:844–852.
- McGaugh JL, McIntyre CK, Power AE (2002) Amygdala modulation of memory consolidation: interaction with other brain systems. *Neurobiol Learn Mem* 78:539–552.
- McIntyre CK, Power AE, Roozendaal B, McGaugh JL (2003) Role of the basolateral amygdala in memory consolidation. *Ann N Y Acad Sci* 985:273–293.
- Pare D (2003) Role of the basolateral amygdala in memory consolidation. *Prog Neurobiol* 70:409–420.
- Reymann K, Frey U, Matthies H (1988) A multi-phase model of synaptic long-term potentiation in hippocampal CA1 neurones: protein kinase C activation and protein synthesis are required for the maintenance of the trace. In: *Synaptic plasticity in the hippocampus* (Haas HL, Buzsáki G, eds), pp 126–129. Berlin: Springer.
- Richter-Levin G (2004) The amygdala, the hippocampus, and emotional modulation of memory. *Neuroscientist* 10:31–39.
- Seidenbecher T, Reymann KG, Balschun D (1997) A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc Natl Acad Sci U S A* 94:1494–1499.
- Straube T, Korz V, Balschun D, Frey JU (2003a) Requirement of beta-adrenergic receptor activation and protein synthesis for LTP-reinforcement by novelty in rat dentate gyrus. *J Physiol* 552 (Pt 3):953–960.
- Straube T, Korz V, Frey JU (2003b) Bidirectional modulation of long-term potentiation by novelty-exploration in rat dentate gyrus. *Neurosci Lett* 344:5–8.
- Thomas SR, Assaf SY, Iversen SD (1984) Amygdaloid complex modulates neurotransmission from the entorhinal cortex to the dentate gyrus of the rat. *Brain Res* 307:363–365.
- Valjakka A, Lukkarinen K, Koivisto E, Lammintausta R, Airaksinen MM, Riekkinen P (1991) Evoked field responses, recurrent inhibition, long-term potentiation and immobility-related nonrhythmic EEG in the dentate gyrus of fimbria-fornix lesioned and control rats. *Brain Res Bull* 26:525–532.
- Vizi ES, Kiss JP (1998) Neurochemistry and pharmacology of the major hippocampal transmitter systems: synaptic and nonsynaptic interaction. *Hippocampus* 8:566–607.

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Capítulo 6: La estimulación del septo medial modula la potenciación sináptica duradera en el giro dentado

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Varios de los resultados antes descritos sugieren que la región del septo medial puede ser parte de los mecanismos de modulación de procesos neuroplásticos en el giro dentado:

1. La mediación demostrada del sistema de fimbria fornix tanto en el reforzamiento conductual como en el iniciado por estimulación de la amígdala.
2. El efecto de bloqueo por atropina del reforzamiento por estimulación de la amígdala ya que la proyección colinérgica en toda la formación del hipocampo es de origen septal.

Basados en esta evidencias se puede proponer la hipótesis de que *la proyección septo-hipocampal es parte del sistema neural de reforzamiento de procesos neuroplásticos*.

Si esto es cierto, debe ser posible provocar un efecto de reforzamiento similar al obtenido con estimulación de la amígdala, mediante la estimulación de la región medial del septum. El objetivo de este trabajo fue comprobar esa hipótesis.

Los resultados muestran que es posible reforzar la LTP en el giro dentado mediante la estimulación de la región medial del septum con la aplicación de estímulos de alta frecuencia al septum (fig. 2A) pero no con estímulos de baja frecuencia (2B). Lo primero mimetiza el efecto de la estimulación de la amígdala pero no lo segundo. El estudio de los mecanismos celulares mostró la participación de los mecanismos de síntesis de proteínas (ver figura 4) mientras que el estudio farmacológico demostró la mediación de aferentes noradrenérgicas (fig. 3B). Inesperadamente, el bloqueo de la transmisión colinérgica no tuvo efectos (fig. 3C). Este último resultado parece contradecir el efecto antes encontrado para la atropina con estimulación de la amígdala y choca con el hecho

de que la principal proyección del septum al hipocampo es colinérgica, aunque la estimulación eléctrica puede activar también fibras que pasan por el septum, como las noradrenérgicas procedentes del locus coeruleus.

Debe tenerse en cuenta que la aplicación de las sustancias bloqueadoras se realizó en el ventrículo cerebral derecho y, por esa razón sus efectos pueden ocurrir no solo en el giro dentado, que es la región de interés, sino en cualquier otra zona del encéfalo. Eso reduce las posibilidades de interpretación inequívoca de los resultados. Para lograr una caracterización más precisa de la contribución de esos neurotransmisores sería necesario su aplicación tópica en las regiones de interés como se mostrará más adelante.

MODULATION OF LATE PHASES OF LONG-TERM POTENTIATION IN RAT DENTATE GYRUS BY STIMULATION OF THE MEDIAL SEPTUM

S. FREY,^a J. A. BERGADO^b AND J. U. FREY^{a*}

^aLeibniz Institute for Neurobiology, Department of Neurophysiology, Brenneckestrasse 6, 39118 Magdeburg, Germany

^bInternational Center for Neurological Restoration, Ave 25, Playa 12100, Havana, Cuba

Abstract—The prolonged maintenance of hippocampal long-term potentiation (LTP) seems to require heterosynaptic events during its induction. We have previously shown that stimulation of the basolateral nucleus of the amygdala (BLA) within a distinct time window can reinforce a transient early-LTP into a long-lasting late-LTP in the dentate gyrus (DG) in freely moving rats. We have shown that this reinforcement was dependent on β -adrenergic and/or muscarinic receptor activation and protein synthesis. However, since the BLA does not directly stimulate the DG the question remained by which inputs such heterosynaptic processes are triggered. We have now directly stimulated the medial septal pathway 15 min after induction of early-LTP in the DG and show that this input is capable of reinforcing early into late-LTP in a frequency-dependent manner. This septal reinforcement of DG LTP was dependent on β -adrenergic receptor activation and protein synthesis. We suggest that the reinforcing effect of the BLA stimulation can, potentially, be mediated via the septal input to the DG, though it differs in its ability to induce or modulate functional plasticity. © 2003 IBRO. Published by Elsevier Science Ltd. All rights reserved.

Key words: late-LTP, hippocampus, reinforcement, memory formation, learning, septum.

Learning is a complex form of experience-dependent plasticity which allows animals, and humans, to modify and adapt their behaviour in response to changes in environmental conditions. Long-term potentiation (LTP) (Bliss and Lomo, 1970; Bliss and Gardner-Medwin, 1971) is an enduring change in synaptic efficacy produced by brief repetitive stimulation of specific afferents and has been considered as a cellular model of long-term memory. Best studied in the dentate gyrus (DG) and the CA1 region of the hippocampus, LTP has long been thought to be the result of the monosynaptic activation of glutamatergic receptors, mainly of the *N*-methyl-D-aspartate (NMDA)-re-

ceptor type (Bashir, et al., 1991). LTP, however, is not a unitary phenomenon. Like memory, LTP has phases that depend on different intracellular molecular mechanisms. Blocking protein synthesis with specific inhibitors allows the dissociation of a protein synthesis-independent early-LTP lasting less than 4 h, from a protein synthesis-dependent late-LTP (Krug et al., 1984; Frey et al., 1988). Whilst the role of NMDA-receptor activation for the induction of LTP seems to be well established, accumulating evidence suggests that other transmitter systems might exert important influences on the maintenance of LTP. Amongst others, acetylcholine (Segal and Auerbach, 1997) as well as catecholamines, like dopamine (Frey et al., 1990) and norepinephrine (Stanton and Sarvey, 1987) are able to influence LTP, mainly in its late components (Frey et al., 1990). Interestingly, these transmitters are thought to be part of the neural systems involved in evaluating and determining the emotional and motivational status of the individual (McGaugh and Introini-Collison, 1987; Haselmo, 1995; McGaugh et al., 1996; Nader and LeDoux, 1999). Recently it was demonstrated that emotional-motivational processes can influence the duration of hippocampal LTP (Seidenbecher et al., 1995, 1997). The main findings of those studies showed that behavioural stimuli with a strong emotional-motivational content, like drinking after 24 h of water deprivation, were able to transform an early-LTP induced by a weak tetanus into a long-lasting LTP beyond 8 h. This effect was abolished by propranolol an antagonist of norepinephrine β -receptors.

One candidate brain structure to subserve such 'behavioural reinforcement' effect is the amygdala. This limbic structure is considered to be part of an emotional memory system (Quirk et al., 1996) and to mediate emotional reinforcement of memory in other brain structures like the hippocampus (Wang and Arvanov, 1998). A functional link between the amygdala and hippocampal LTP has been demonstrated as well (Ikegaya et al., 1994, 1995a,b; Akirav and Richter-Levin, 1999a,b; Kamiya and Ozawa, 1998). These studies, however, concentrated on the induction or very early stages of early-LTP. In a recent report, we have assessed whether amygdala stimulation can also influence the maintenance of LTP. The main findings showed that amygdala stimulation is able to transform a transient early-LTP at the perforant path (PP)–DG synapses into a late-LTP when applied within a 30-min time window, before or after the induction of LTP (with one exception: simultaneous tetanization did not lead to a reinforcement). The effect was protein synthesis-dependent and could be abolished by noradrenergic or muscarinic

*Corresponding author. Tel: +49-391-6263-422; fax: +49-391-6263-421.

E-mail address: frey@ifn-magdeburg.de (J. U. Frey).

Abbreviations: BDNF, brain-derived neurotrophic factor; BLA, basolateral nucleus of the amygdala; DG, dentate gyrus; EPSP, excitatory postsynaptic potential; hfs, high-frequency stimulation; lfs, low-frequency stimulation; LTP, long-term potentiation; MS, medial septum; MSHfs, high-frequency stimulation of the medial septum; MSlfs, low-frequency stimulation of the medial septum; NMDA, *N*-methyl-D-aspartate; PP, perforant path; PPHfs, high-frequency stimulation of the perforant path; PSA, population spike amplitude.

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antagonists but not by dopaminergic or NMDA-receptor antagonists (Frey et al., 2001).

There are no known direct efferents linking the amygdala with the DG (Chen et al., 1999). Two alternative pathways, via the entorhinal cortex or the medial septum (MS), might sustain such interactions. We have shown that the interruption of the septo-hippocampal afferents by the lesion of the fimbria-fornix affects the late effects of stimulating the amygdala (Jas et al., 2000). However, this does not prove that stimulating the septum might reproduce the effects of stimulating the amygdala.

In the present paper we show results demonstrating the efficacy of the septal stimulation to prolong LTP at the DG.

EXPERIMENTAL PROCEDURES

Male Wistar rats (250–300 g at the time of preparation) were used. All experiments were performed in compliance with the relevant laws and institutional guidelines and have been approved by the Land Sachsen-Anhalt. After surgery for electrode implantation animals were housed individually in plastic translucent cages with free access to water and food.

Surgery was performed under pentobarbital narcosis (40 mg/kg, i.p.) supplemented with additional doses if required. The animals were mounted in a stereotactic frame (bregma 1 mm above lambda) and electrodes were lowered to their final positions under electrophysiological control of the evoked potentials to achieve the best sensitivity. A monopolar recording electrode was placed at the hilar region of the DG (coordinates, AP: –2.8 mm, ML: 1.8 mm, from bregma, DV: –3.5 mm from the dural surface) whilst bipolar stimulating electrodes were set at the PP (AP: –6.9 mm, ML: 4.1 mm from bregma, DV: –2.5 mm from the dura) the basolateral nucleus of the amygdala (BLA) (AP: –2.4 mm, ML: 5.0 mm from bregma, DV: –7.6 mm from the dura) and the MS (AP: +1.7 mm, ML: 1.8 mm, DV: –7.0 mm from the dura; tilted 15° toward the midline in the coronal plane). Miniscrews attached to the skull served as indifferent electrode and earth respectively.

After 2 weeks of recovery animals were habituated to the recording set for at least 4 h before starting the experiment.

LTP was studied by the evaluation of potentials evoked at the DG by stimulation of the PP. An input/output curve was constructed for each animal and the stimulation intensity required to induce a 40% of the maximal population spike amplitude (PSA) determined for each animal. This intensity was used for test recordings as well as for inducing LTP stimulating the PP. For stimulating the BLA or the MS 0.1-ms square pulses of constant intensity were used. The stimulation of BLA but not of the MS produced evoked potentials at the DG (Frey et al., 2001), but, as the recording electrode position was optimised for the PP input, they were small and showed flattened I/O relationships. Therefore a constant 400 μ A current was used for both inputs. PSA-recording and analysis were favoured against the slope of field excitatory postsynaptic potentials (field-EPSPs) since the latter is relatively unstable in freely moving animals, especially considering that the PP stimulation intensity was adjusted to obtain an optimal population spike, which negatively influences the field-EPSP. During high-frequency stimulation (hfs), i.e. during LTP induction the spike is required to induce late-LTP. We have previously studied the relationships between PSA potentiation and EPSP potentiation and found a positive correlation between both variables over a wide range of tetanization frequencies (Frey et al., 2001; López Planes et al., 1999). In addition, in a few experiments with potentials containing a relatively large proportion of an “EPSP,” we have analysed the time course of that response's initial positive slope after stimulus artefact. The slope of this potential followed the time course of the corresponding PSA, respectively (data not shown).

Test recordings of the PP–DG-evoked potentials were recorded during 1 h (baseline) before applying any of the stimulation protocols. Each recorded potential was the average of five consecutive single pulses (0.05 Hz). Hfs or low-frequency stimulation (lfs) protocols were applied to the PP, the BLA or MS after baseline recording. The averaged value of the PSA in baseline records was used to calculate the percentile change in this variable in test recordings taken during the following 8 h, and 24 h after stimulation.

Hfs consisted of three trains of 15 impulses at 200 Hz (10-s intertrain interval) whilst lfs consisted of 45 impulses at 0.1 Hz

The experimental protocol included the following stimulation groups

Single stimulation

- hfs of the PP (PPhfs), $n=8$
- hfs of the MS (MSHfs), $n=6$
- lfs of the MS (MSlfs), $n=6$

Double stimulation (15-min interval)

- PPhfs followed by MSHfs (PPhfs+MSHfs), $n=5$
- PPhfs followed by MSlfs (PPhfs+MSlfs), $n=8$

Pharmacology

The application procedure for drugs were the same as described earlier (Frey et al., 2001) when DG–LTP reinforcement was investigated at an interval between LTP induction in the DG and reinforcing stimulation of 15 min. There were no non-specific behavioural effects detected during or after drug application. The following pharmacological experiments were carried out:

- PPhfs+MSHfs and the application of NaCl, $n=6$
- PPhfs+MSHfs and the application of the β -adrenergic receptor blocker propranolol (6.76 nmol), $n=4$
- PPhfs+MSHfs and the application of the muscarinic receptor blocker atropine (1 nmol), $n=4$
- PPhfs+MSHfs and the application of the protein synthesis inhibitor anisomycin (0.905 mol), $n=5$

The animals were randomly assigned to the experimental groups. Some animals were used in more than one protocol. In those cases the second protocol was at least 1 week after the first, and after baseline values completely recovered their original level.

The statistical analysis was performed using the Wilcoxon *t*-test for paired samples for within group comparisons (test recording versus own baseline), whilst the Kruskal-Wallis non-parametric ANOVA was used for between-groups comparisons. In each case a two-tailed $P<0.05$ was established as statistically significant.

At the end of the experiments the brains of all animals were fixed by intracardiac perfusion under narcosis and the location of the electrodes was histologically confirmed. Only animals with a correct electrode location (i.e. within the structures of interest) were included in the final results.

All efforts were made to minimise the number of rats used and their suffering. All experiments have been performed with permission of the local legislative authorities of the Land Sachsen-Anhalt.

RESULTS

The group with single PPhfs ($n=8$) was introduced as control of the classical LTP that can be induced in the DG after stimulating the glutamatergic fibres arising from the entorhinal cortex. As shown in Fig. 1B, this stimulus paradigm induces an increase of the PSA relative to baseline recordings. This LTP, however, is decremental as shown in Fig. 1B. The level of potentiation slowly declines to baseline values within

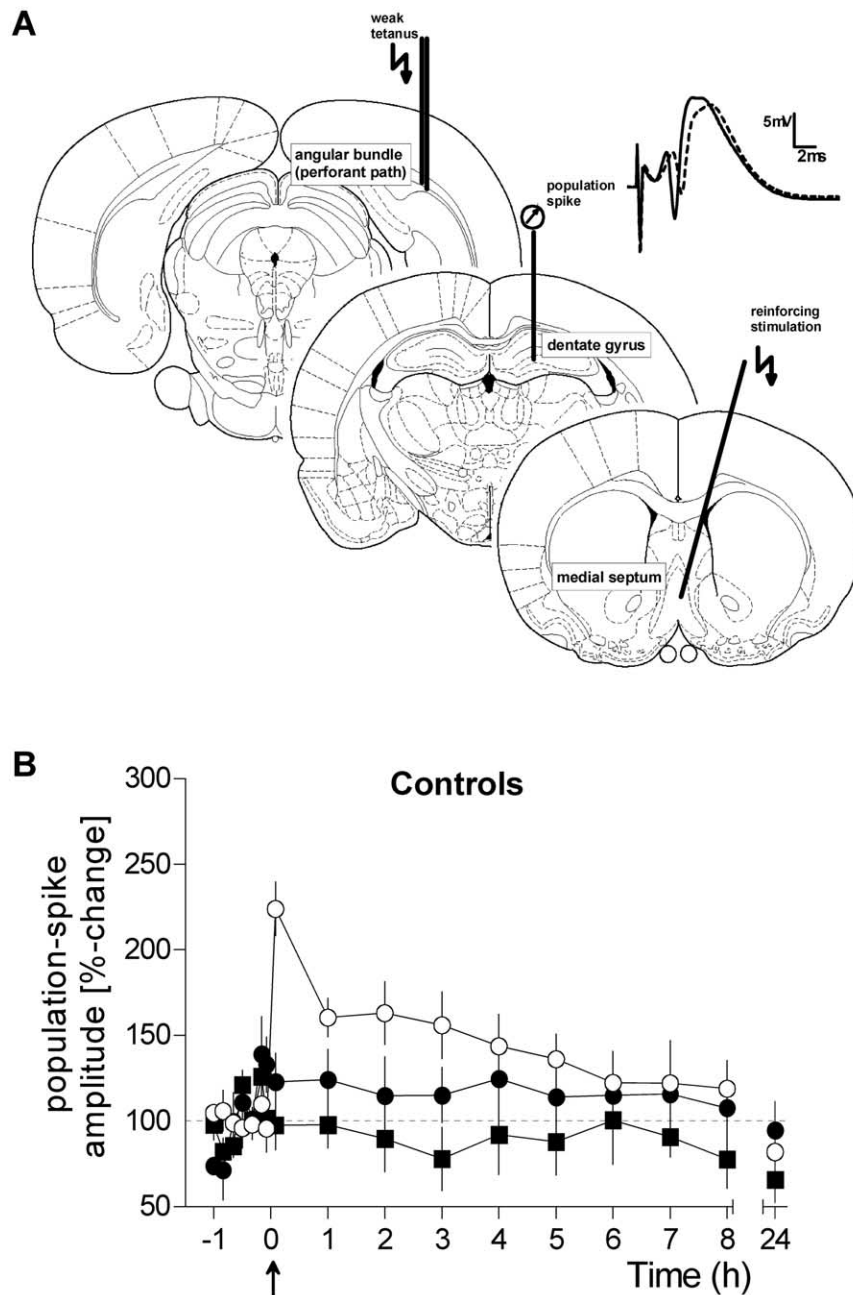


Fig. 1. Control experiments. (A) Schematic illustration of electrode localisation. Insets show analog examples of recordings obtained before (dotted line) and after LTP induction (filled line) of the perforant path (upper right). Notice the increase in the PSA (downwards spike-like deflection) and the small increase in the slope of the population EPSP (first upward deflection). The PSA was used as an indicator of potentiation. (B) Time course of early-LTP at the PP–DG synapses (control-LTP, open circles) and the influence of hfs (closed circles) or lfs (closed squares) of the MS on test potentials in the DG. The arrow indicates the time of tetanic stimulation or lfs of the PP or the MS, respectively.

the first 3–6 h. The Wilcoxon test confirmed a significant increase of the PSA only during the first 2 h after induction.

Hfs ($n=6$) or lfs ($n=6$) of the MS alone (Fig. 1B) produced no significant change on the PSA in test recordings during the same 24-h period. However, when the MS was stimulated with high frequency after the induction of LTP in DG (Fig. 2A, PPHfs+MSHfs, $n=5$) a significant modification in LTP time course appeared (Kruskal-Wallis test). Instead of declining to non-significant levels after 2 h, the

population spike remained statistically significantly potentiated even after 24 h. Such an effect of MS stimulation seemed to be frequency-dependent whilst lfs of this region was ineffective in influencing the maintenance of early DG–LTP (Fig. 2B, $n=8$). No significant differences between curves were found by the Kruskal-Wallis test. It is important to emphasise that no change was detected with respect to the level of initial potentiation when hfs or lfs was applied after induction of DG–early-LTP.

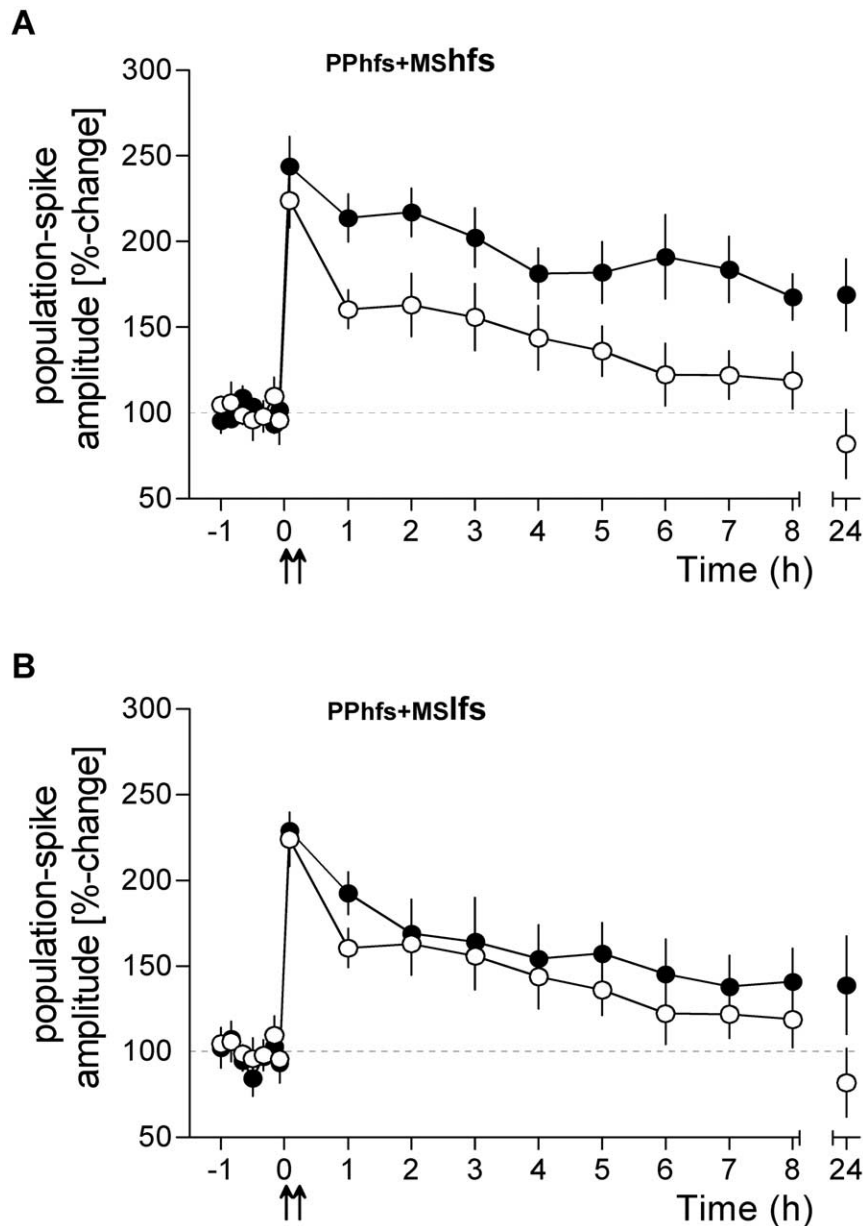


Fig. 2. MS stimulation can prolong LTP at the PP–DG synapses. (A) Hfs of the MS 15 min after induction of LTP at the PP–DG synapses (PPhfs+MShfs, filled circles) significantly prolonged the time course of LTP for at least 24 h. This curve was also significantly different from control early-LTP ($P < 0.05$, Kruskal–Wallis test). For comparison, the time course of DG early-LTP is presented (open circles). (B) Lfs of the MS (PPhfs+MSlfs, filled circles) has no reinforcing effect on LTP at the PP–DG synapses. No significant differences between this and the control LTP curve were observed (Kruskal–Wallis test). For comparison, the time course of DG early-LTP is presented in both graphs (open circles). The arrows indicate the time of stimulation of the PP or the MS.

Pharmacological interventions resulted in the following effects on a control reinforcement of DG–early-LTP by subsequent tetanization of the MS 15 min after the DG tetanization ($n=5$): Control experiments with the application of saline instead of drug treatment (Fig. 3A, NaCl controls, $n=6$) revealed no difference when compared with untreated control reinforced animals. However, the application of the β -adrenergic receptor antagonist propranolol (Fig. 3B, $n=4$) but not of the muscarinic receptor antagonist atropine (Fig. 3C, $n=4$), prevented the reinforcing effect of MShfs on DG–early-LTP. After the application of

propranolol a statistically significant potentiation was only observed for the first 2 h after the tetanization (Wilcoxon test). Similar results as with propranolol were obtained with the reversible protein synthesis inhibitor anisomycin (Fig. 4, $n=5$). Application of anisomycin prevented LTP after 1 h onward (Wilcoxon test).

DISCUSSION

The tetanization paradigm we have used in the present experiments effectively induced LTP at the PP–DG syn-

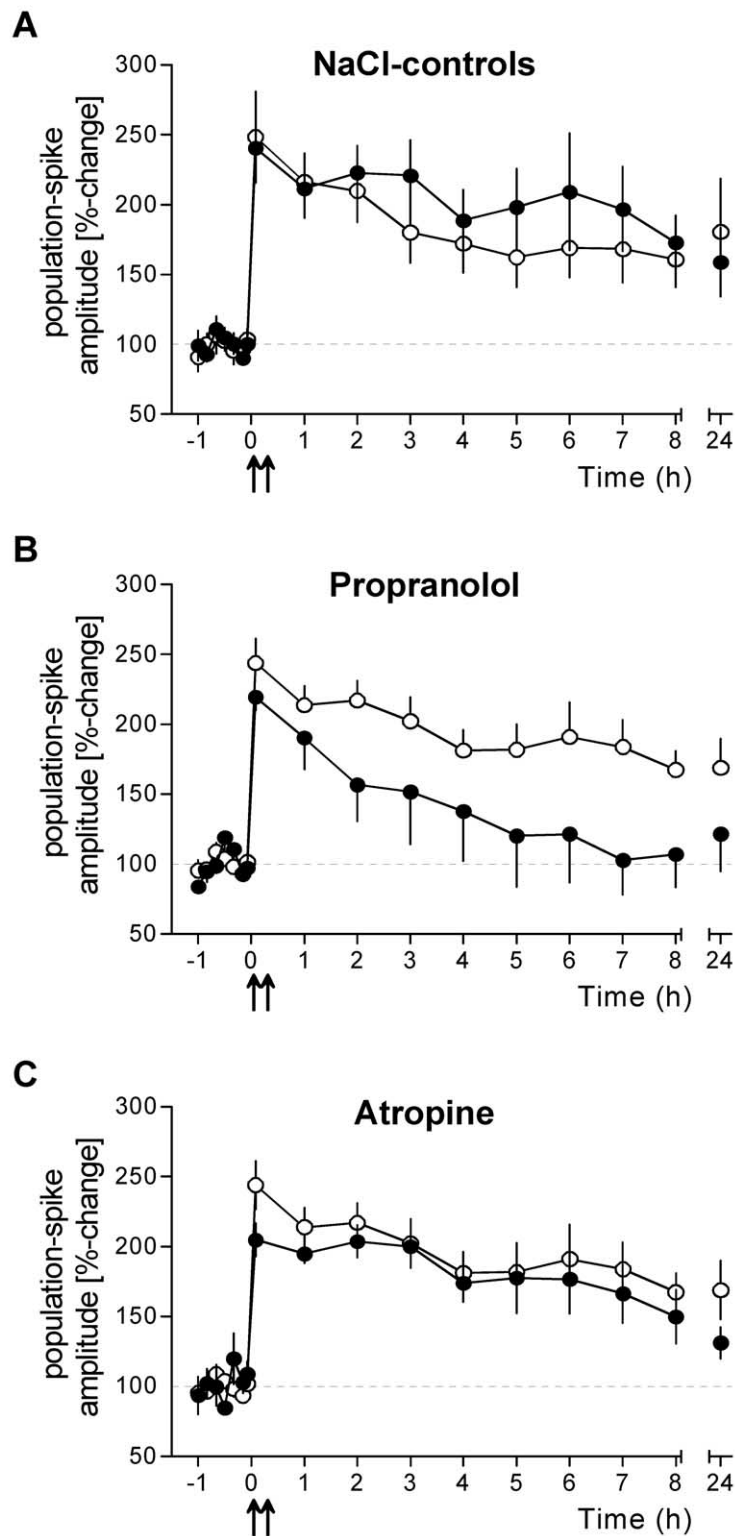


Fig. 3. The influence of pharmacological substances on the MS–hfs reinforcement of DG–early-LTP. (A) A repeated pattern of hfs of the MS 15 min after induction of LTP at the PP–DG synapses significantly prolongs the time course of early-LTP in untreated (open circles; see also Fig. 2A) as well as in animals in which saline (NaCl) was injected into the ventricle (filled circles). The curves were statistically significant different from control early-LTP ($P < 0.05$, Kruskal–Wallis test) for at least 24 h. (B) The application of the β -adrenergic receptor antagonist propranolol i.c.v. prevented (closed circles) the PPhfs–MShfs reinforcement from 3 h onward. (C) The application of the muscarinic receptor antagonist atropine (closed circles) was ineffective in influencing PPhfs–MShfs reinforcement. For comparison the time course of the PPhfs–MShfs-reinforced late-LTP (untreated and NaCl controls) is presented in all three graphs (open circles). The arrows indicate the time of tetanic stimulation of the PP or the MS.

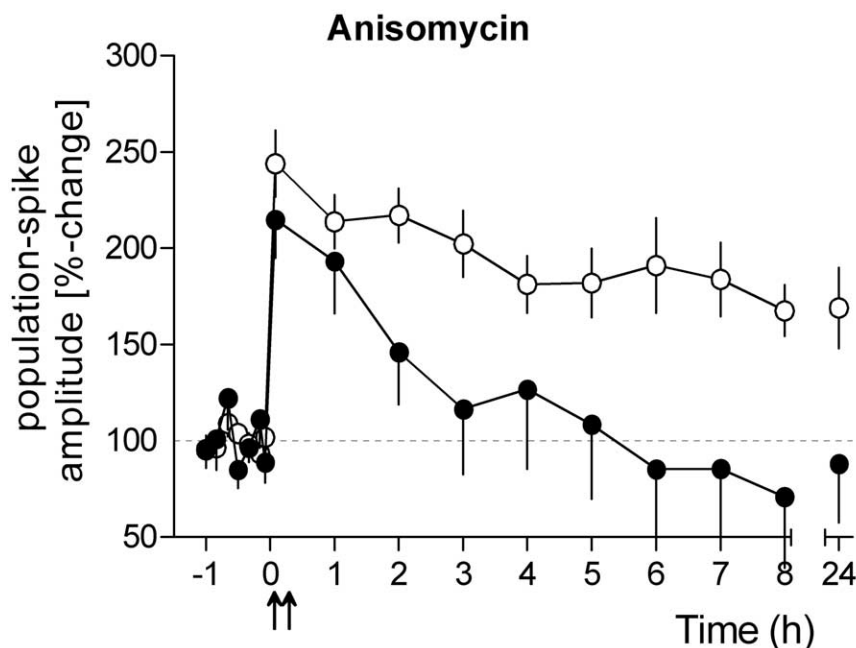


Fig. 4. The influence of anisomycin on PPhfs–MShfs-reinforced LTP. The application of the reversible protein synthesis inhibitor anisomycin i.c.v. prevented (closed circles) the PPhfs–MShfs reinforcement from 2 h onward. For comparison the time course of the PPhfs–MShfs-reinforced late-LTP (untreated and NaCl controls) is presented (open circles). The arrows indicate the time of tetanic stimulation of the PP or the MS.

apses. This potentiation was short lasting (<4 h) corresponding to early-LTP, similar to the one obtained in previous reports using an identical stimulation pattern (Seidenbecher et al., 1997; Frey et al., 2001). In contrast, an lfs pattern, as the one used in the present study, induced no increase of the PSA; instead a slight but significant slowly developing decrease (6 h) appeared after lfs of the PP alone (Frey et al., 2001). Interestingly, hfs or lfs of the BLA alone induced a slowly developing, slight depression of the PSA (Frey et al., 2001). Whether such effects might be causally related to the depression observed after lfs of the PP, and whether they could be considered a form of long-term depression deserves future examination.

Our results after hfs of the MS in the paired PPhfs–MShfs protocol (Fig. 2A) support our previous findings where we have shown that heterosynaptic, non-glutamatergic afferents can modulate the maintenance of LTP induced at a glutamatergic synaptic population. Septo-hippocampal interactions are well known to be involved in the production of θ rhythm in the hippocampal EEG (Bland et al., 1999), a cholinergic rhythm which can be induced by activation of the amygdala (Dringenberg and Vanderwolf, 1996). Septal stimulation can affect PP–DG-evoked potentials (Robinson and Racine, 1982; Carre and Harley, 2000) and hippocampal LTP (Robinson and Racine, 1982; Robinson, 1986) using a shorter interval between stimuli and restricted to the initial induction of LTP.

Electrical stimulation of the MS can activate local cholinergic fibres projecting to the hippocampus, but it can also activate noradrenergic, dopaminergic, serotonergic, GABA-ergic and histaminergic fibres passing through the septum and reaching the hippocampus via the fimbria-fornix. In our previous work we have shown that the block-

ade of cholinergic (muscarinic) and β -adrenergic, but not dopaminergic D1-receptors, abolishes the reinforcing effect of stimulating the BLA (Frey et al., 2001). Surprisingly, we found in this study only an effect of the β -adrenergic but not muscarinic receptor blocker on the reinforcement of DG LTP by stimulating the MS. An explanation for that could be that the location of the stimulation electrode in the MS was optimal to modulate noradrenergic but suboptimal to stimulate cholinergic inputs to the hippocampus. Future experiments will further investigate this hypothesis. However, we could also show that the reinforcement of DG LTP by MS stimulation required protein synthesis thusly demonstrating a transformation of early into late-LTP by β -adrenergic receptor stimulation.

Acetylcholine and norepinephrine are both able to produce an LTP-like, slowly developing potentiation if applied to the DG (Stanton and Sarvey, 1985b; Auerbach and Segal, 1994) very similar to the potentiation produced by the neurotrophin brain-derived neurotrophic factor (BDNF) (Kang and Schuman, 1995). All these forms of potentiation are protein synthesis-dependent (Stanton and Sarvey, 1985a; Kang and Schuman, 1996). Our working hypothesis is that the prolonged maintenance of LTP in general depends on heterosynaptic activity: glutamatergic events are mainly required to store afferent information transiently (Morris and Frey, 1997; Frey and Morris, 1998), i.e. generating early-LTP and to mark the activated synapses in a specific way, which makes them able to capture plasticity-related proteins necessary to transform early into late-LTP. Heterosynaptic events are involved in regulating the availability of plasticity-related proteins thusly guaranteeing the potential transformation of a short- into a long-term memory trace. In distinct cases heterosynaptic activity such as

local increases in cyclic AMP may cause the non-specific activation ("tagging") of synapses which would explain the non-specific drug-induced potentiation by different modulatory substances such as norepinephrine, dopamine, BDNF, protein kinase A activators, etc. (Stanton and Sarvey, 1985a; Frey et al., 1993; Auerbach and Segal, 1994; Kang and Schuman, 1995; Frey, 2001). All of these processes are included in the synaptic-tagging hypothesis (Frey and Morris, 1997) which provides a rationale to understand the heterosynaptic requirement for long-lasting plastic changes to occur.

The same events might explain the reinforcing effects of our double-stimulation paradigm. The first tetanus to the PP, along with inducing LTP, tags the activated synapses. The effect of the second tetanus is not to increase the level of potentiation but to activate molecular cascades leading to macromolecular synthesis (Frey et al., 1993, 1995) for instance by the further accumulation of intracellular Ca^{2+} by glutamatergic or other transmitter systems which may activate processes inducing protein synthesis, and the distribution and insertion of the newly synthesised proteins at the tagged synaptic contacts (Frey and Morris, 1997, 1998). Previous and the current results support our hypothesis that field stimulation and induction of LTP in the PP–DG synapses can be modified by heterosynaptic events (Dunwiddie et al., 1982; Stanton and Sarvey, 1987; Bramham et al., 1997).

Therefore, the stimulation of the septum reproduced some aspects of the reinforcing effects of the BLA. There are, however some important differences which must be considered in future experiments. Only hfs of the MS is effective to prolong DG LTP, whilst lfs of the BLA also resulted in an effective reinforcement (Frey et al., 2001). On the other hand, only hfs of the PP is able to induce a potentiation by itself, hfs of the amygdala or the septum can act only on and modulate an existing potentiation elsewhere, such as at the PP–DG synapses. The concept of instructive (glutamatergic PP) and modulatory synapses (septal and entorhinal inputs) is well illustrated in this example (Matthies, 1989). The knowledge obtained in this study is not only of theoretical value. We have recently shown that amygdala–hippocampus interactions (Almaguer et al., 2002), as well as behavioural reinforcement of LTP (Bergado et al., 2001) are impaired in aged rats with cognitive deficits. Further studies on these mechanisms should contribute to the development of new strategies to treat memory disorders and thusly, could have a direct practical value.

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REFERENCES

- Akirav I, Richter-Levin G (1999a) Biphasic modulation of hippocampal plasticity by behavioral stress and basolateral amygdala stimulation in the rat. *J Neurosci* 19:10530–10535.
- Akirav I, Richter-Levin G (1999b) Priming stimulation in the basolateral amygdala modulates synaptic plasticity in the rat dentate gyrus. *Neurosci Lett* 270:83–86.
- Almaguer W, Estupiñán B, Frey JU, Bergado JA (2002) Aging impairs amygdala–hippocampus interactions involved in hippocampal LTP. *Neurobiol Aging* 23:319–324.
- Auerbach JM, Segal M (1994) A novel cholinergic induction of long-term potentiation in rat hippocampus. *J Neurophysiol* 72:2034–2040.
- Bashir ZI, Alford S, Davies SN, Randall AD, Collingridge GL (1991) Long-term potentiation of NMDA receptor-mediated synaptic transmission in the hippocampus. *Nature* 349:156–158.
- Bergado JA, Almaguer W, Ravelo J, Rosillo JC, Frey JU (2001) Behavioral reinforcement of long-term potentiation is impaired in aged rats with cognitive deficiencies. *Neuroscience* 108:1–5.
- Bland BH, Oddie SD, Colom LV (1999) Mechanisms of neural synchrony in the septohippocampal pathway underlying hippocampal theta generation. *J Neurosci* 19:3223–3237.
- Bliss TV, Gardner-Medwin AR (1971) Long-lasting increases of synaptic influence in the unanesthetized hippocampus. *J Physiol* 216:32P–33P.
- Bliss TV, Lomo T (1970) Plasticity in a monosynaptic cortical pathway. *J Physiol* 207:61P.
- Bramham CR, Bacher-Svendsen K, Sarvey JM (1997) LTP in the lateral perforant path is β -adrenergic receptor-dependent. *Neuroreport* 8:719–724.
- Carre GP, Harley CW (2000) Glutamatergic activation of the medial septum complex: an enhancement of the dentate gyrus population spike and accompanying EEG and unit changes. *Brain Res* 861:16–25.
- Chen GQ, Kolbeck R, Barde YA, Bonhoeffer T, Kossel A (1999) Relative contribution of endogenous neurotrophins in hippocampal long-term potentiation. *J Neurosci* 19:7983–7990.
- Dringenberg HC, Vanderwolf CH (1996) Cholinergic activation of the electrocorticogram: an amygdaloid activating system. *Exp Brain Res* 108:285–296.
- Dunwiddie TV, Roberson NL, Worth T (1982) Modulation of long-term potentiation: effects of adrenergic and neuroleptic drugs. *Pharmacol Biochem Behav* 17:1257–1264.
- Frey JU (2001) Long-lasting hippocampal plasticity: cellular model for memory consolidation? In: *Cell polarity and subcellular RNA localization* (Richter D, ed), pp 27–40. Berlin: Springer-Verlag.
- Frey S, Bergado-Rosado J, Seidenbecher T, Pape HC, Frey JU (2001) Reinforcement of early long-term potentiation (early-LTP) in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction mechanisms of late-LTP. *J Neurosci* 21:3697–3703.
- Frey U, Huang Y-Y, Kandel ER (1993) Effects of cAMP simulate a late stage of LTP in hippocampal CA1 neurons. *Science* 260:1661–1664.
- Frey U, Krug M, Reymann KG, Matthies H (1988) Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Res* 452:57–65.
- Frey U, Morris RGM (1997) Synaptic tagging and long-term potentiation. *Nature* 385:533–536.
- Frey U, Morris RGM (1998) Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 21:181–188.
- Frey U, Schollmeier K, Reymann KG, Seidenbecher T (1995) Asymptotic hippocampal long-term potentiation in rats does not preclude additional potentiation at later phases. *Neuroscience* 67:799–807.
- Frey U, Schroeder H, Matthies H (1990) Dopaminergic antagonists prevent long-term maintenance of posttetanic LTP in the CA1 region of rat hippocampal slices. *Brain Res* 522:69–75.
- Hasselmo ME (1995) Neuromodulation and cortical function: modeling the physiological basis of behavior. *Behav Brain Res* 67:1–27.
- Ikegaya Y, Saito H, Abe K (1994) Attenuated hippocampal long-term potentiation in basolateral amygdala-lesioned rats. *Brain Res* 656:157–164.

- Ikegaya Y, Saito H, Abe K (1995a) High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 22:203–207.
- Ikegaya Y, Saito H, Abe K (1995b) Requirement of basolateral amygdala neuron activity for the induction of long-term potentiation in the dentate gyrus in vivo. *Brain Res* 671:351–354.
- Jas J, Almaguer W, Frey JU, Bergado J (2000) Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res* 861:186–189.
- Kamiya H, Ozawa S (1998) Kainate receptor-mediated inhibition of presynaptic Ca^{2+} influx and EPSP in area CA1 of the rat hippocampus. *J Physiol* 509:833–845.
- Kang H, Schuman EM (1995) Long-lasting neurotrophin-induced enhancement of synaptic transmission in the adult hippocampus. *Science* 267:1658–1662.
- Kang HJ, Schuman EM (1996) A requirement for local protein synthesis in neurotrophin-induced hippocampal synaptic plasticity. *Science* 273:1402–1406.
- Krug M, Lössner B, Ott T (1984) Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Res Bull* 13:39–42.
- López Planes J, Almaguer Melian W, Jas García J, Bergado Rosado JA (1999) Influencia de la frecuencia de estimulación sobre procesos de plasticidad sináptica en el giro dentado de la rata. *Arch Neurosci (Mex)* 4:9–20.
- Matthies H (1989) In search of cellular mechanisms of memory. *Prog Neurobiol* 32:277–349.
- McGaugh JL, Cahill L, Roozendaal B (1996) Involvement of the amygdala in memory storage: interaction with other brain systems. *Proc Natl Acad Sci USA* 93:13508–13514.
- McGaugh JL, Introini-Collison IB (1987) Hormonal and neurotransmitter interactions in the modulation of memory storage: involvement of the amygdala. *Int J Neurol* 21–22:58–72.
- Morris RGM, Frey U (1997) Hippocampal synaptic plasticity: role in spatial learning or the automatic recording of attended experience? *Philos Trans R Soc Lond Biol* 352:1489–1503.
- Nader K, LeDoux J (1999) The dopaminergic modulation of fear: quinpirole impairs the recall of emotional memories in rats. *Behav Neurosci* 113:152–165.
- Quirk GJ, Armony JL, Repa JC, Li XF, LeDoux JE (1996) Emotional memory: a search for sites of plasticity. *Cold Spring Harbor Symp Quant Biol* 61:247–257.
- Robinson GB (1986) Enhanced long-term potentiation induced in rat dentate gyrus by coactivation of septal and entorhinal inputs: temporal constraints. *Brain Res* 379:56–62.
- Robinson GB, Racine RJ (1982) Heterosynaptic interactions between septal and entorhinal inputs to the dentate gyrus: long-term potentiation effects. *Brain Res* 249:162–166.
- Segal M, Auerbach JM (1997) Muscarinic receptors involved in hippocampal plasticity. *Life Sci* 60:1085–1091.
- Seidenbecher T, Balschun D, Reymann KG (1995) Drinking after water deprivation prolongs “unsaturated” LTP in the dentate gyrus of rats. *Physiol Behav* 57:1001–1004.
- Seidenbecher T, Reymann KG, Balschun D (1997) A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc Natl Acad Sci USA* 94:1494–1499.
- Stanton PK, Sarvey JM (1985a) Blockade of norepinephrine-induced long-lasting potentiation in the hippocampal dentate gyrus by an inhibitor of protein synthesis. *Brain Res* 361:276–283.
- Stanton PK, Sarvey JM (1985b) Depletion of norepinephrine, but not serotonin, reduces long-term potentiation in the dentate gyrus of rat hippocampal slices. *J Neurosci* 5:2169–2176.
- Stanton PK, Sarvey JM (1987) Norepinephrine regulates long-term potentiation of both the population spike and dendritic EPSP in hippocampal dentate gyrus. *Brain Res Bull* 18:115–119.
- Wang RY, Arvanov VL (1998) M100907, a highly selective 5-HT_{2A} receptor antagonist and a potential atypical antipsychotic drug, facilitates induction of long-term potentiation in area CA1 of the rat hippocampal slice. *Brain Res* 779:309–313.

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Capítulo 7: La administración de norepinefrina, pero no de oxotremorina refuerza procesos de plasticidad sináptica

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En este trabajo intentamos una aproximación farmacológica diferente al tema que hemos venido desarrollando. En lugar de intentar bloquear determinados sistemas neurotransmisores con antagonistas, nos propusimos suplantar la estimulación directa de determinados núcleos por la administración de agonistas conocidos de los sistemas neurotransmisores seleccionados.

En este caso se trató de la transmisión noradrenérgica y colinérgica que fueron las que mostraron resultados de bloqueo del reforzamiento cuando se emplearon antagonistas específicos.

La hipótesis de trabajo fue la siguiente: *Si el bloqueo de la transmisión noradrenérgica y colinérgica impide el reforzamiento conductual, entonces la administración de agonistas directos de estos sistemas puede mimetizar el efecto del reforzamiento conductual.*

Como agonista noradrenérgico empleamos norepinefrina, mientras que para la transmisión colinérgica empleamos oxotremorina, un agonista de los receptores muscarínicos.

La administración intraventricular de norepinefrina 15 minutos después de la inducción de la LTP en el giro dentado mostró efectos similares a los del reforzamiento. Este efecto fue dosis dependiente y al parecer muestra el clásico patrón de U invertida. La dosis más baja empleada fue poco efectiva y la dosis más alta fue inefectiva. La oxotremorina no mostró el mismo efecto de reforzamiento para ninguna de las tres dosis empleadas.

Es posible mimetizar el efecto de reforzamiento afectivo mediante la administración de agonistas de la neurotransmisión noradrenérgica en dosis adecuadas.

Long-term potentiation in the dentate gyrus in freely moving rats is reinforced by intraventricular application of norepinephrine, but not oxotremorine

William Almaguer-Melian^a, Yeneissy Rojas-Reyes^a, Armando Alvare^a, Juan C. Rosillo^a,
Julietta U. Frey^{b,*}, Jorge A. Bergado^{a,b,*}

^a Department of Experimental Neurophysiology, International Center for Neurological Restoration (CIREN), Havana, Cuba

^b Department of Neurophysiology, Leibniz Institute for Neurobiology, Brennekestr. 6, 39118 Magdeburg, Germany

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Abstract

Growing evidence suggests that processes of synaptic plasticity, such as long-term potentiation (LTP) occurring in one synaptic population, can be modulated by consolidating afferents from other brain structures. We have previously shown that an early-LTP lasting less than 4 h (E-LTP) in the dentate gyrus can be prolonged by stimulating the basolateral amygdala, the septum or the locus coeruleus within a specific time window. Pharmacological experiments have suggested that noradrenergic (NE) and/or cholinergic systems might be involved in these effects. We have therefore investigated whether the direct intraventricular application of agonists for NE- or muscarinic receptors is able to modulate synaptic plasticity. E-LTP was induced at the dentate gyrus of freely moving rats using a mild tetanization protocol that induces only an E-LTP. NE or oxotremorine (OXO) were applied icv 10 min after the tetanus. Results show that low doses of NE (1.5 and 5 nM) effectively prolong LTP. A higher dose (50 nM) was not effective. None of the OXO doses employed (5, 25, and 50 nM) showed similar effects. These results stress the importance of transmitter-specific modulatory influences on the time course of synaptic plasticity, in particular NE whose application mimics the reinforcing effect of directly stimulating limbic structures on LTP.

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Keywords: Long-term potentiation; Reinforcement; Norepinephrine; Oxotremorine; Early-LTP; Late-LTP; Functional plasticity

1. Introduction

Long-term potentiation (LTP) is a form of functional neural plasticity which has been considered a cellular correlate of memory formation (Bliss & Collingridge, 1993; Matthies, 1982). LTP has phases, as does memory and the molecular mechanisms implicated in LTP phases are also implicated in memory. An initial early-

LTP (E-LTP) lasting less than 4 h and not depending on protein synthesis might eventually be converted into a late-LTP (L-LTP) extending beyond 4 h and requiring the synthesis of new proteins (Krug, Lössner, & Ott, 1984; Frey, Krug, Reymann, & Matthies, 1988; Matthies et al., 1989; Matthies, 1998).

Studies by Seidenbecher et al. for the first time showed that an E-LTP, elicited at the dentate gyrus of freely moving rats by a mild tetanus, is transformed into a L-LTP if, within a restricted time window, it is associated with a behavioral stimulus of strong motivational significance (Seidenbecher, Balschun, & Reymann, 1995; Seidenbecher, Reymann, & Balschun, 1997). This behavioral, or motivational reinforcement of LTP

* Corresponding authors. Fax: +49 391 6263 421 (J.U. Frey), +53 7 332420 (J.A. Bergado).

E-mail addresses: frey@ifn-magdeburg.de (J.U. Frey), bergado@neuro.ciren.cu, jorgebergado@yahoo.com (J.A. Bergado).

depends on the function of the amygdala (Almaguer-Melian, Martínez-Martí, Frey, & Bergado, 2003), is protein synthesis dependent (Bergado, Almaguer-Melian, Kostenko, Frey, & Frey, 2003) and is blocked by propranolol, a β -receptor blocker (Seidenbecher et al., 1997). These data supported our hypothesis that late-LTP requires heterosynaptic modulation—here by norepinephrine (NE)—in addition to previously activated glutamatergic synapses—for review see Matthies (1998) and Frey and Morris (1998).

A similar conversion of E-LTP into L-LTP as that seen with behavioral reinforcement can be obtained by a direct electrical stimulation of the amygdala shortly before or after the induction of an E-LTP by a mild tetanus. The effect is abolished by propranolol and by the muscarinic antagonist atropine (Frey, Bergado, Seidenbecher, Pape, & Frey, 2001). A further evidence for a role for cholinergic systems in the modulatory effect leading to reinforcement is the observation that lesioning the main cholinergic projection to the hippocampus abolishes the L-LTP promoting effects of the amygdala stimulation (Jas, Almaguer, Frey, & Bergado, 2000). However, propranolol, but not atropine, blocked the reinforcing effects on LTP of stimulating the septum (Frey, Bergado, & Frey, 2003).

All of the LTP-reinforcing actions—i.e., motivational, amygdalar or septal—seem to activate protein synthesis, a prerequisite for late-LTP to occur (Bergado et al., 2003; Frey et al., 2001, 2003). Both NE and acetylcholine (ACh) are able to activate protein synthesis via molecular cascades involving kinases regulating transcription, like MAPK (Greenwood & Dragunow, 2002; Hamilton & Nathanson, 2001; Massey, Bhabra, Cho, Brown, & Bashir, 2001; Rosenblum, Futter, Jones, Hulme, & Bliss, 2000; Stratton, Baraban, & Worley, 1988) or protein kinase A (Frey & Morris, 1998). It appears thus pertinent to test whether the direct administration of NE or a cholinergic agonist can mimic the reinforcing effects on LTP shown in the above-mentioned paradigms.

The results obtained showed that NE, but not oxotremorine (a muscarinic agonist), dose-dependently reinforce LTP in the dentate gyrus.

2. Materials and methods

2.1. Experimental animals

Male Sprague–Dawley rats, weighing 250 g by the time of surgery, were obtained from the Cuban national breeder (CENPALAB, La Habana) and kept in translucent Macrolon cages (five animals per cage before surgery) under controlled temperature (22–25 °C), humidity (lower than 60%), and light exposure conditions (12:12 h light:dark). All experiments were carried

out during the light phase of the cycle (8:00 am to 6:00 pm). Access to food and water were ad libitum during the experiment. Animal handling and experimental procedures were performed under observance of the Cuban Regulations for the Use of Laboratory Animals.

2.2. Surgery

Under narcosis (chloral hydrate 420 mg/kg, ip) the animals were implanted with a monopolar recording electrode at the dentate gyrus (AP: –3.8 mm, ML: 2.0 mm, DV: –3–7) a bipolar stimulating electrode at the medial perforant pathway (AP: –7.5 mm, ML: 4.0 mm, DV: –3.9 mm) and a guide cannulae directed at the lateral ventricle (AP: –0.3 mm, ML: 1.3 mm, DV: –2.4 mm). All implants were placed in the right hemisphere using bregma as reference (bregma and lambda were at the same horizontal plane). Additionally, two miniscrews were fixed over the parietal and frontal bones to serve as reference electrode and connection to earth. Electrodes were then connected to mini-sockets and all implanted material was secured with dental cement. After surgery the animals were individually housed and treated with antibiotics during 7 days to prevent infection (solution of 250 mg tetracycline dissolved in 500 ml water).

2.3. Electrophysiology

A week after surgery the animals were carried to the recording chamber (between 8:00 and 8:30 am) and connected to the stimulation and recording set through a swivel, which allowed the free movement of the animals. Stimulation and recording of the evoked potentials was started no less than after 4 h of habituation.

Each recording consisted of five averaged consecutive responses of 0.1 Hz. Responses were bandpass filtered (1–5000 Hz) and amplified using an AB-621G (Nihon-Kohden, Japan) bioelectric amplifier and passed through a CED 1401+ A/D converter to an IBM-compatible computer which allow visualization of the evoked potentials and the automatic measurement of its components with the aid of a self-made program (IntraCell for Windows, IfN Magdeburg). The same program was used to control the delivery of electrical stimulation to the perforant path by an isolated pulse stimulator (A-M Systems Mod. 2001, USA).

The amplitude of the population spike (PSA), the component reflecting the synchronous discharge of action potentials by the granule cell population, was measured and the changes in its relative amplitude (percent from baseline) were used to evaluate changes in synaptic efficacy. We have preferred to base on the PSA rather than the population excitatory post-synaptic potential (pEPSP) because of technical reasons while working in vivo and freely moving animals. On the other hand,

changes in PSA reflect the modifications in the output from the neuronal population that we have investigated, which is of a great significance in functional terms.

An Input/Output curve was first obtained from each animal, by recording the evoked responses in the dentate gyrus to stimulation (100, 200, 400, 600, and 800 μ A) of the perforant path. The intensity required to produce a 40% maximum PSA was determined and used for the rest of the experiments. Twelve recordings were taken at 5 min interval. The averaged PSA was used as baseline.

Long-term potentiation was induced by a mild tetanus consisting of three trains of 15 impulses at 200 Hz (10 s intertrain interval). Five minutes after induction of LTP a test recording was taken to evaluate the initial level of potentiation. Starting 30 min after tetanization test recordings were taken every 15 min for up to 6 h to follow the time course of LTP. A final series of four recordings separated by 15 min intervals was taken 24 h after LTP induction.

2.4. Substances and injection

NE, a natural ligand of α - and β -receptors, and oxotremorine, an agonist of muscarinic cholinergic receptors, were employed (both from Sigma, Saint Louis, USA). Both substances were tested for their efficacy before the beginning of the experiments using a different group of animals. NE was able to induce a significant increase in heart rate (ip 1 mg/kg) and oxotremorine induced sustained theta activity in the hippocampal EEG (ip 0.1 mg/kg). For the experiments the substances were applied in the right lateral ventricle in different doses. For NE, doses of 1.5, 5, and 50 nM were employed. NE was dissolved in saline solution containing ascorbic acid (2 mg/ml) to prevent oxidation. Oxotremorine, dissolved in saline, was prepared in concentrations of 5, 25, and 50 nM. The solutions were prepared fresh every day in a way that a final volume of 1 μ l was injected in every case. The injection was performed over 1 min using an injection cannula (1 mm longer than the guide cannula) connected to a micro-syringe (Hamilton, USA). The cannula was left in place for 5 min to reduce retrograde flux of the injected substance.

2.5. Experimental groups

Experimental groups were constituted according to the drug applied, the dose, and the stimulation pattern applied.

To test the stability of the evoked potentials over the time period studied and the effect of the applied substances and the vehicles, a series of control groups were studied. In these groups no LTP was induced. Substances were applied 5 min after 1-h baseline recordings.

One group (basal, $n = 10$) received no treatment. Two groups were injected with 1 μ l saline solution (basal + NaCl, $n = 10$) or the vehicle containing ascorbate (basal + Veh, $n = 8$). Additionally, two groups received 1 μ l of a solution containing 5 nM NE (basal + NE, $n = 9$) or the same volume of 25 nM oxotremorine (basal + OXO, $n = 8$).

To establish the time course of the LTP induced by the mild tetanus and the effects of the vehicles the following groups were created: a control LTP group ($n = 11$) in which LTP was induced and followed without injection, and two groups in which LTP was induced and 1 μ l of saline (LTP + NaCl, $n = 8$) or vehicle with ascorbic acid (LTP + Veh, $n = 8$) was injected icv 10 min later.

To study the effects of NE on LTP we used three groups, corresponding to the different doses (1.5 nM, $n = 6$; 5 nM, $n = 7$; and 50 nM, $n = 7$) of NE-injected icv 10 min after the induction of LTP with a mild tetanus. In the same way, three groups injected with oxotremorine were studied corresponding to each of the doses employed (5 nM, $n = 8$; 25 nM, $n = 8$; and 50 nM, $n = 9$).

2.6. Histology

After finishing the electrophysiological studies all animals were intracardially perfused with formalin under narcosis. The brains were extracted and processed for histology with cresyl violet to assess the correct location of the cannulae. The microscopic observation of the slices confirmed the correct location—in the direction of the lateral ventricle—in all cases.

2.7. Statistics

Data were analyzed using a two-way ANOVA with Group (treatment) and Time (repeated measures) as factors. Post hoc studies with the Duncan's test were performed when significant differences between groups were confirmed. When the factor Time was significant, paired t tests were carried out to confirm within group differences to baseline. In every case differences were considered significant only if $p < .05$.

3. Results

The comparison among control groups, without LTP induction (Fig. 1) showed no differences between groups ($F_{4,36} = 0.775722$), suggesting that none of the control treatments with vehicle or substance induced significant deviations from baseline. However and unexpectedly, the factor Time showed significant differences ($F_{7,252} = 2.083680$). The within groups analysis (paired t test) showed that this was caused only by the group treated with 5 nM NE. The animals that received NE

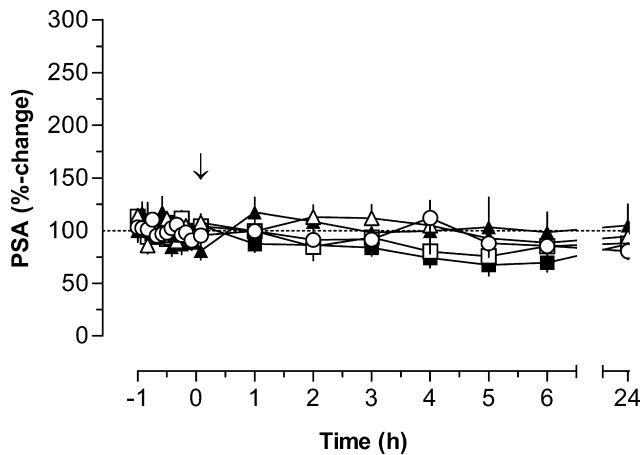


Fig. 1. Control recordings of the PSA and the influence of drugs investigated on baseline potentials measured as the population spike amplitude (PSA) during a period of 24 h. The mean PSA ($\pm$ SEM) relative to baseline and expressed in percentile values (PSA % change) for each time point and for each group are shown. No differences between groups were found (ANOVA, two-way). Factor Time showed significant differences only for the group treated with norepinephrine (basal + NE, filled squares) during 4–6 h. Other symbols: open circles: basal group; open triangles: basal + NaCl group; filled triangles: basal + OXO group; and open squares: basal + Veh group. The arrow ($\downarrow$) shows the time at which substances were applied, except for the group basal which received no treatment or handling at that time.

developed a slight depression of the PSA that was significant between 4 and 6 h after injection. Other groups (basal, basal + NaCl, basal + OXO, and basal + Veh) showed no significant deviation from baseline over time.

The control groups in which LTP was induced (control LTP, LTP + Veh, and LTP + NaCl) showed no significant differences to each other (Fig. 2). The two-way ANOVA revealed no differences for the factor Group ($F_{2,21} = 0.52733$). The factor Time showed a significant effect ($F_{8,168} = 19.32421$) confirming that a relevant change in the PSA occurred in all groups after tetanization. This potentiation remained significant for 3 h (Duncan's test).

The administration of NE in the lower doses (1.5 and 5 nM) significantly prolongs LTP, but not the higher dose of 50 nM, as shown in Fig. 3. The two-way ANOVA revealed significant effects for both factors, Group ($F_{2,17} = 6.29625$) and Time ($F_{8,136} = 16.33383$). The tetanization paradigm employed induced an increase in the PSA in all groups, but the duration of the potentiation was different according to the dose of NE employed. The post hoc Duncan's test showed that the groups receiving 1.5 and 5 nM NE were significantly different from the vehicle treated group while the animals injected with 50 nM NE did not differ from vehicle. The *t* test for paired samples proved that LTP was prolonged in the 1.5 and 5 nM groups to 5 and 6 h, respectively.

Oxotremorine produced no effects on the magnitude or the duration of LTP (Fig. 4). Treatments using different doses of oxotremorine did not reveal pronounced

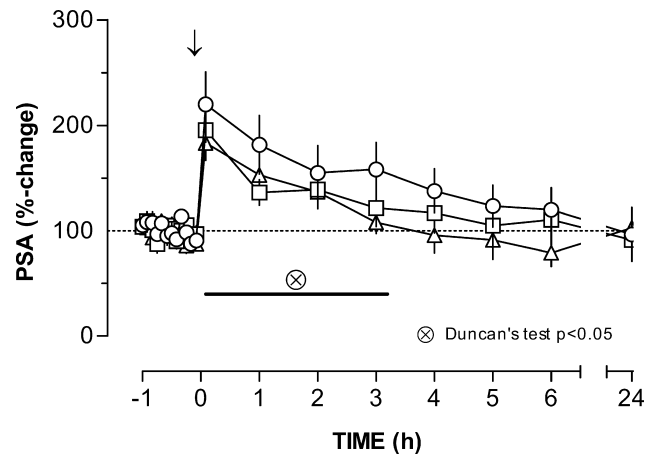


Fig. 2. Induction and maintenance of E-LTP and the influence of the vehicles employed. The mean PSA % change ($\pm$ SEM) for each time point before (time -1 to 0) and after tetanization (times 0–24) and for each group are shown. In all the groups the pattern of tetanization applied at time 0 induced an increase in the PSA which remained significant only up to the third hour after induction (Duncan's test). No differences between groups were found (ANOVA, two-way). Symbols: open circles: Control LTP group; open squares: LTP + Veh group; and open triangles: LTP + NaCl. The arrow ($\downarrow$) shows the time at which substances were applied, except for the group Control LTP which received no treatment or handling at that time.

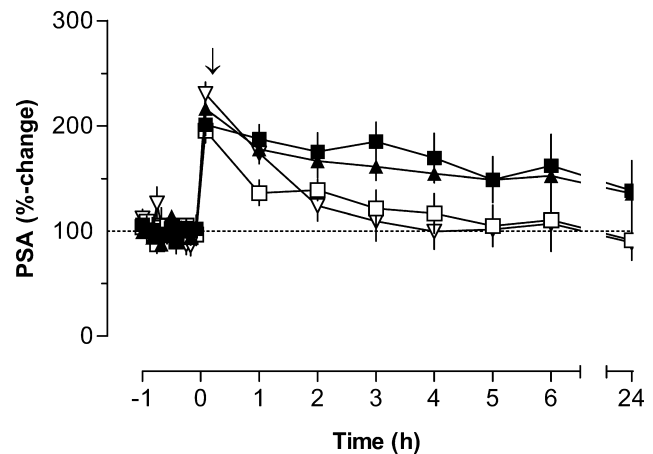


Fig. 3. Effect of norepinephrine on E-LTP. The mean PSA % change ($\pm$ SEM) for each time point before (time -1 to 0) and after tetanization (times 0–24) and for each group are shown. Significant differences between groups were found (ANOVA, two-way). All groups reached the same level of PSA potentiation, but they differ in how long the potentiated state holds. Groups treated with 1.5 or 5 nM norepinephrine (NE 1.5 filled squares; and NE 5 filled triangles) differed significantly from vehicle treated animals (Veh open squares) or those receiving a higher dose of NE (NE 50 open inverted triangles) as confirmed by the post hoc Duncan's test. Significant differences from baseline values were found up to 5 h after LTP-induction and up to 6 h for the NE 1.5 and NE 5 groups, respectively. The arrow ($\downarrow$) shows the time at which substances were applied.

changes on early-LTP when compared with the NaCl-injected control group as indicated by the two-way ANOVA ($F_{3,29} = 1.99304$). The factor Time, as expected

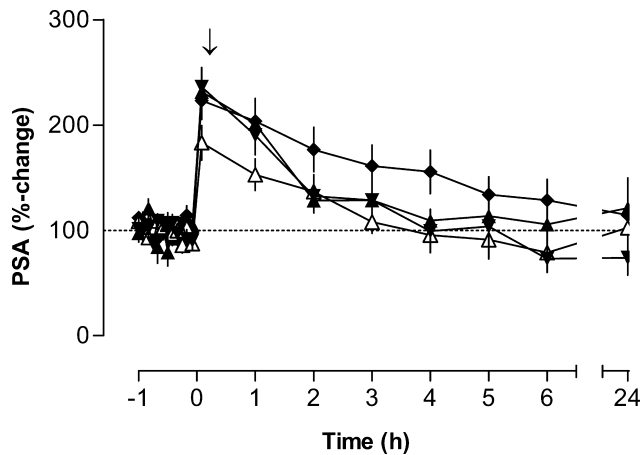


Fig. 4. Effect of oxotremorine on E-LTP. The mean PSA % change ($\pm$ SEM) for each time point before (time -1 to 0) and after tetanization (times 0 – 24) and for each group are shown. No major differences among groups were found (ANOVA two ways). All groups reached the same initial level of potentiation after LTP induction which decline to baseline within 3 h. Only the group receiving the higher dose of oxotremorine (LTP + OXO 50 filled diamonds) showed a slight, but significant Group/Time interaction. Other symbols: open triangles: LTP + NaCl group; filled inverted triangles: LTP + OXO 5 ; and filled triangles LTP + OXO 25 . The arrow ($\downarrow$) shows the time at which substances were applied.

after tetanizing, resulted in significant changes in PSA ($F_{8,232} = 58.43984$). The interaction Group $\times$ Time ($F_{24,232} = 1.89460$) was also significant. The post hoc Duncan's test determined that this slight, time-dependent effect of oxotremorine was caused by the highest dose (50 nM) used.

4. Discussion

The results in the basal group of animals, without tetanization, vehicle or drug treatment showed that the evoked potentials in the dentate gyrus induced by stimulation of the perforant path were stable over time. This is important to rule out a major effect of circadian factors inducing transient modifications in synaptic transmission. Furthermore, neither the injection procedure nor the volume injected affected the evoked potentials. None of the injected vehicles or oxotremorine produced changes in the PSA. However, the control injection of NE induced a late and slight, but significant decrease of the PSA. Earlier reports have shown that the administration of NE to hippocampal slices was able to increase the amplitude of the PSA (Lacaille & Harley, 1985) developing a delayed onset potentiation of the PSA (maximum was reached after about 1 h). Additionally, this NE-induced potentiation in vitro was dependent on protein synthesis (Stanton & Sarvey, 1985b). Therefore, the authors concluded that this form of potentiation resembled a LTP-like phenomenon. What are the reasons for the obvious contradiction between results obtained in vitro or—as in our case—in

vivo? There are a couple of possibilities such as the difference between the in vitro and in vivo conditions. Furthermore, the NE concentration employed was much higher (10 μ M) than the ones employed in our study in vivo. Additionally, extrahippocampal activity may interact in vivo more dramatically on evoked potentials than in vitro. In any case the depression reported here seemed to be the consequence of an interaction of NE-receptor activation on long-lasting plastic events rather than a direct effect of NE on membrane potentials. Interestingly, in previous studies we have shown a similar depression of the PSA recorded in the dentate gyrus either after low- or high-frequency stimulation of the amygdala (Frey et al., 2001) or in aged animals submitted to a behavioral reinforcement paradigm. Whether there is a functional or mechanistic link between them remains to be determined.

The control LTP group confirmed that the pattern of mild tetanization employed was only able to induce E-LTP, decaying within 4 h. The saline and vehicle groups failed to reveal any non-specific effect of manipulation, injection or volume on the time course of E-LTP. Any modification in LTP after substance application should, therefore, be attributed to a specific effect of the applied agonist.

Interestingly, NE dose-dependently influenced the time course of induced E-LTP if the mild tetanus preceded NE-application. The lower doses showed a reinforcing effect, consisting in a prolongation of the potentiated state and not in an increase of the level of PSA potentiation. This effect was similar to the one observed after behavioral reinforcement by motivational stimuli (Seidenbecher et al., 1997), and after direct, electrical stimulation of the basolateral amygdala (Frey et al., 2001). Both, motivational and amygdala stimulation induced a reinforcement of LTP which was blocked by the β -1-receptor antagonist propranolol. The effect reported here strengthens the hypothesis that NE might specifically be a part of the mechanisms mediating such reinforcement effects finally expressed in the dentate gyrus.

The mechanism for this effect of NE on LTP might be explained in terms of a modulatory-metabolic action of NE. NE is a strong activator of the cAMP-PKA cascade (Gereau & Conn, 1994; Stanton & Sarvey, 1985a) which may activate the protein synthesis required for the expression of L-LTP. This β -adrenergic reinforcement seemed particularly important to reinforce LTP induced by relatively weak tetanic stimuli (Straube & Frey, 2003). A functional relationship like the one suggested here may support and explain the multiple experiments showing a modulating-reinforcing action of NE on memory (McGaugh & Cahill, 1997). The action of NE might occur directly on the dentate gyrus, but it might involve other brain structures, like the amygdala (McGaugh, 2002). This limbic structure plays a critical role in emotional reactions and the motivational modu-

lation of memory and LTP (Richter-Levin, 2004; Richter-Levin & Akirav, 2003).

Regarding the action of the muscarinic agonist oxotremorine, our findings surprisingly do not confirm the expectation of a reinforcing effect of cholinergic-receptor activation on E-LTP in the dentate gyrus. None of the employed doses of this substance was able to significantly modify LTP, only a mild Group $\times$ Time interaction was found with the a relatively high dose. Cholinergic mechanisms have been proposed to sustain memory functions (Baratti, Opezzo, & Kopf, 1993; Blokland, 1995; Conner, Culbertson, Packowski, Chiba, & Tuszynsky, 2003; Ohno, Yamamoto, & Watanabe, 1994; Ramirez Lugo, Miranda, Escobar, & Bermudez-Rattoni, 2003; Warburton et al., 2003) and deterioration of cholinergic systems is considered as a major cause for age-related memory impairments (Bartus, 2000; Bartus, Dean III, Beer, & Lippa, 1982). Both lines of evidence have guided experimental and clinical trails to restore memory functions based on pharmaceuticals (Lyketsos, Corazzini, Steele, & Kraus, 1996; Sands, Katz, & Schneider, 1999) transplants (Fernández et al., 1994; Leanza, Martinez-Serrano, & Bjorklund, 1998) or trophic factors (Fischer, Sirevaag, Wiegand, Lindsay, & Bjorklund, 1994). The role of cholinergic mechanisms in LTP has been, however, controversial. While some reports describe a facilitatory, and even an inducing role (Auerbach & Segal, 1994; Boyd, Trepel, & Racine, 2000; Hess & Donoghue, 1999; Leung, Shen, Rajakumar, & Ma, 2003; Motooka et al., 2001; Segal & Auerbach, 1997; Ye, Qi, & Qiao, 2001) others found mild or no effects (Abe, Nakata, Mizutani, & Saito, 1994; Feasey-Truger, Li, & ten Bruggencate, 1992; Jouvenceau, Billard, Lamour, & Dutar, 1996; Kleschevnikov, Sinden, & Marchbanks, 1994). Previous results from our group have also provided contradictory results with respect to cholinergic influences via muscarinic receptors in LTP-reinforcement mechanisms. While atropine blocked reinforcement induced by stimulation of the basolateral amygdala (Frey et al., 2001), it did not block reinforcement induced by stimulation of the septal area (Frey et al., 2003) suggesting that some actions might involve other extrahippocampal structures. A targetted application instead of an icv injection might contribute to clarify this issue.

In summary, direct activation of β -adrenergic, but not muscarinic receptors, can convert early- into late-LTP in the dentate gyrus in vivo in freely moving animals. The possibility of modulating L-LTP by the direct application of agonistic drugs may lead to the development of drugs to treat memory disorders in the future.

References

- Abe, K., Nakata, A., Mizutani, A., & Saito, H. (1994). Facilitatory but nonessential role of the muscarinic cholinergic system in the generation of long-term potentiation of population spikes in the dentate gyrus in vivo. *Neuropharmacology*, 33, 847–852.
- Almaguer-Melian, W., Martínez-Martí, L., Frey, J. U., & Bergado, J. A. (2003). The amygdala is part of the behavioral reinforcement system modulating long-term potentiation in rat hippocampus. *Neuroscience*, 119, 319–322.
- Auerbach, J. M., & Segal, M. (1994). A novel cholinergic induction of long-term potentiation in rat hippocampus. *Journal of Neurophysiology*, 72, 2034–2040.
- Baratti, C. M., Opezzo, J. W., & Kopf, S. R. (1993). Facilitation of memory storage by the acetylcholine M₂ muscarinic receptor antagonist AF-DX 116. *Behavioral and Neural Biology*, 60, 69–74.
- Bartus, R. T. (2000). On neurodegenerative diseases, models, and treatment strategies: Lessons learned and lessons forgotten a generation following the cholinergic hypothesis. *Experimental Neurology*, 163, 495–529.
- Bartus, R. T., Dean III, R. L., Beer, B., & Lippa, A. S. (1982). The cholinergic hypothesis geriatric memory dysfunction. *Science*, 217, 408–417.
- Bergado, J. A., Almaguer-Melian, W., Kostenko, S., Frey, S., & Frey, J. U. (2003). Behavioral reinforcement of long-term potentiation in rat dentate gyrus in vivo is protein synthesis-dependent. *Neuroscience Letters*, 351, 56–58.
- Bliss, T. V. P., & Collingridge, G. L. (1993). A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature*, 361, 31–39.
- Blokland, A. (1995). Acetylcholine: A neurotransmitter for learning and memory? *Brain Research and Reviews*, 21, 285–300.
- Boyd, T. E., Trepel, C., & Racine, R. J. (2000). Cholinergic modulation of neocortical long-term potentiation in the awake, freely moving rat. *Brain Research*, 881, 28–36.
- Conner, J. M., Culbertson, A., Packowski, C., Chiba, A., & Tuszynsky, M. H. (2003). Lesions of the basal forebrain cholinergic system impair task acquisition and abolish cortical plasticity associated with motor skill learning. *Neuron*, 38, 819–829.
- Feasey-Truger, K. J., Li, B.-H., & ten Bruggencate, G. (1992). Lesions of the medial septum which produce deficits in working/spatial memory do not impair long-term potentiation in the CA3 region of the rat hippocampus in vivo. *Brain Research*, 591, 296–304.
- Fernández, C. I., Bergado, J., De la Cuétara, K., Nunez, N., Castellanos, O., González, O., Torres, A., Macías, R., Alvarez, L., & Molina, H. (1994). Brain aging and neurotransplantation. II. Effects of septal cell suspension grafts to hippocampus of aged rodents on learning and memory impairments. *Archives of Gerontology and Geriatrics*, 18(Suppl. 4), 59–65.
- Fischer, W., Sirevaag, A., Wiegand, S. J., Lindsay, R. M., & Bjorklund, A. (1994). Reversal of spatial memory impairments in aged rats by nerve growth factor and neurotrophins 3 and 4/5 but not by brain-derived neurotrophic factor. *Proceedings of the National Academy of Sciences of the United States of America*, 91, 8607–8611.
- Frey, S., Bergado, J. A., & Frey, J. U. (2003). Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum. *Neuroscience*, 118, 1055–1062.
- Frey, S., Bergado, J. A., Seidenbecher, T., Pape, H. C., & Frey, J. U. (2001). Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP. *Journal of Neuroscience*, 21, 3697–3703.
- Frey, U., Krug, M., Reymann, K. G., & Matthies, H. (1988). Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro. *Brain Research*, 452, 57–65.
- Frey, U., & Morris, R. G. M. (1998). Synaptic tagging: Implications for late maintenance of hippocampal long-term potentiation. *Trends Neuroscience*, 21, 181–188.
- Gereau, R. W., & Conn, P. J. (1994). A cyclic AMP-dependent form of associative synaptic plasticity induced by coactivation of b-adren-

- ergic receptors and metabotropic glutamate receptors in rat hippocampus. *Journal of Neuroscience*, 14, 3310–3318.
- Greenwood, J. M., & Dragunow, M. (2002). Muscarinic receptor-mediated phosphorylation of cyclic AMP response element binding protein in human neuroblastoma cells. *Journal of Neurochemistry*, 82, 389–397.
- Hamilton, S. E., & Nathanson, N. M. (2001). The M₁ receptor is required for muscarinic activation of MAP kinase in murine cerebral cortical neurons. *Journal of Biological Chemistry*.
- Hess, G., & Donoghue, J. P. (1999). Facilitation of long-term potentiation in layer II/III horizontal connections of rat motor cortex following layer I stimulation: Route of effect and cholinergic contributions. *Experimental Brain Research*, 127, 279–290.
- Jas, J., Almaguer, W., Frey, J. U., & Bergado, J. (2000). Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Research*, 861, 186–189.
- Jouveceau, A., Billard, J. M., Lamour, Y., & Dutar, P. (1996). Persistence of CA1 hippocampal LTP after selective cholinergic denervation. *Neuroreport*, 7, 948–952.
- Kleschevnikov, A. M., Sinden, J. D., & Marchbanks, R. (1994). Fimbria-fornix lesions impair spatial performance and induce epileptic-like activity but do not affect long-term potentiation in the CA1 region of rat hippocampal slices. *Brain Research*, 656, 221–228.
- Krug, M., Lössner, B., & Ott, T. (1984). Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats. *Brain Research Bulletin*, 13, 39–42.
- Lacaille, J. C., & Harley, C. W. (1985). The action of norepinephrine in the dentate gyrus: Beta-mediated facilitation of evoked potentials in vitro. *Brain Research*, 358, 210–220.
- Leanza, G., Martinez-Serrano, A., & Bjorklund, A. (1998). Amelioration of spatial navigation and short-term memory deficits by grafts of foetal basal forebrain tissue placed into the hippocampus and cortex of rats with selective cholinergic lesions. *The European Journal of Neuroscience*, 10, 2353–2370.
- Leung, L. S., Shen, B., Rajakumar, N., & Ma, J. (2003). Cholinergic activity enhances hippocampal long-term potentiation in CA1 during walking in rats. *The Journal of Neuroscience*, 23, 9297–9304.
- Lyketsos, C. G., Corazzini, K., Steele, C. D., & Kraus, M. F. (1996). Guidelines for the use of tacrine in Alzheimer's disease: Clinical application and effectiveness. Johns Hopkins Dementia Research Clinic. *Journal of Neuropsychiatry and Clinical Neurosciences*, 8, 67–73.
- Massey, P. V., Bhabra, G., Cho, K., Brown, M. W., & Bashir, Z. I. (2001). Activation of muscarinic receptors induces protein synthesis-dependent long-lasting depression in the perirhinal cortex. *The European Journal of Neuroscience*, 14, 145–152.
- Matthies, H. Neuronale Grundlagen der Gedächtnisbildung. Klix, F. and Spada, H. Enzyklopädie der Psychologie [6], 15–42. 1998. Göttingen, Hogrefe Verlag für Psychologie. Kognition. Ref Type: Serial (Book, Monograph).
- Matthies, H. (1982). Plasticity in the nervous system. An approach to memory research. In C. Ajmone-Marsan & H. Matthies (Eds.), *Neuronal plasticity and memory formation* (pp. 1–15). New York: Raven Press.
- Matthies, H., Reymann, K. G., Krug, M., Frey, U., Loessner, B., & Popov, N. (1989). Multiple stages of posttetanic LTP and memory. In Rahmann (Ed.), *Fundamentals of memory formation: Neuronal plasticity and brain function* (pp. 395–396). Stuttgart, New York: Gustav Fischer Verlag.
- McGaugh, J. L. (2002). Memory consolidation and the amygdala: A systems perspective. *Trends in Neurosciences*, 25, 456.
- McGaugh, J. L., & Cahill, L. (1997). Interaction of neuromodulatory systems in modulating memory storage. *Behavioral Brain Research*, 83, 31–38.
- Motooka, Y., Kondoh, T., Nomura, T., Tamaki, N., Tozaki, H., Kanno, T., & Nishizaki, T. (2001). Selective cholinergic denervation inhibits expression of long-term potentiation in the adult but not infant rat hippocampus. *Brain Research. Developmental Brain Research*, 129, 119–123.
- Ohno, M., Yamamoto, T., & Watanabe, S. (1994). Blockade of hippocampal M₁ muscarinic receptors impairs working memory performance of rats. *Brain Research*, 650, 260–266.
- Ramirez Lugo, L., Miranda, I., Escobar, L., & Bermudez-Rattoni, F. (2003). The role of cortical cholinergic pre- and post-synaptic receptors in taste memory formation. *Neurobiology of Learning and Memory*, 79, 184–193.
- Richter-Levin, G. (2004). The amygdala, the hippocampus, and emotional modulation of memory. *Neuroscientist*, 10, 31–39.
- Richter-Levin, G., & Akirav, I. (2003). Emotional tagging of memory formation in the search for neural mechanisms. *Brain Research. Brain Research Reviews*, 43, 247–256.
- Rosenblum, K., Futter, M., Jones, M., Hulme, E. C., & Bliss, T. V. (2000). ERK1/II regulation by the muscarinic acetylcholine receptors in neurons. *The Journal of Neuroscience*, 20, 977–985.
- Sands, L. P., Katz, I., & Schneider, L. (1999). Assessing individual patients for cognitive benefits from acetylcholinesterase inhibitors. *Alzheimer Disease and Associated Disorders*, 13, 26–33.
- Segal, M., & Auerbach, J. M. (1997). Muscarinic receptors involved in hippocampal plasticity. *Life Science*, 60, 1085–1091.
- Seidenbecher, T., Balschun, D., & Reymann, K. G. (1995). Drinking after water deprivation prolongs unsaturated LTP in the dentate gyrus of rats. *Physiology and Behavior*, 57, 1001–1004.
- Seidenbecher, T., Reymann, K. G., & Balschun, D. (1997). A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proceedings of the National Academy of Sciences of the United States of America*, 94, 1494–1499.
- Stanton, P. K., & Sarvey, J. M. (1985a). The effect of high-frequency electrical stimulation and norepinephrine on cyclic AMP levels in normal versus norepinephrine-depleted rat hippocampal slices. *Brain Research*, 358, 343–348.
- Stanton, P. K., & Sarvey, J. M. (1985b). Blockade of norepinephrine-induced long-lasting potentiation in the hippocampal dentate gyrus by an inhibitor of protein synthesis. *Brain Research*, 361, 276–283.
- Stratton, K. R., Baraban, J. M., & Worley, P. F. (1988). Norepinephrine stimulation of adenylate cyclase potentiates protein kinase C action: Electrophysiological studies in the dentate gyrus. *Synapse*, 2, 614–618.
- Straube, T., & Frey, J. U. (2003). Involvement of beta-adrenergic receptors in protein synthesis-dependent late long-term potentiation (LTP) in the dentate gyrus of freely moving rats: The critical role of the LTP induction strength. *Neuroscience*, 119, 473–479.
- Warburton, E. C., Koder, T., Cho, K., Massey, P. V., Duguid, G., Barker, G. R., Aggleton, J. P., Bashir, Z. I., & Brown, M. W. (2003). Cholinergic neurotransmission is essential for perirhinal cortical plasticity and recognition memory. *Neuron*, 38, 987–996.
- Ye, L., Qi, J. S., & Qiao, J. T. (2001). Long-term potentiation in hippocampus of rats is enhanced by endogenous acetylcholine in a way that is independent of N-methyl-D-aspartate receptors. *Neuroscience Letters*, 300, 145–148.

Capítulo 8: Efecto de paradigmas de reforzamiento de la plasticidad sináptica sobre la liberación de neurotransmisores en el giro dentado

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Los estudios farmacológicos previos habían mostrado la necesidad de participación de sistemas de neurotransmisión colinérgico y noradrenérgico. Sin embargo, en algunos trabajos aparecieron resultados contradictorios, sobre todo con relación al papel de la acetil colina. El estudio de estimulación de la amígdala muestra que la atropina bloquea el efecto. Sin embargo, no fue así en el caso de la estimulación del septum y tampoco la oxotremorina mimetizó los efectos del reforzamiento.

Con todas esas interrogantes en consideración, intentamos resolver las contradicciones midiendo directamente la liberación de estos y otros neurotransmisores en el giro dentado cuando se estimula la amígdala basolateral o cuando se someten los animales a reforzamiento conductual.

Este fue un estudio exploratorio. Más que testar una hipótesis intentaba encontrar pistas para interpretar las contradicciones antes mencionadas. Para ello empleamos la técnica microdialisis cerebral que permite la recogida de muestras de fluido del cerebro *in situ* e *in vivo*. Se aprovechó además la disponibilidad de ratas viejas con deficiencias cognitivas comprobadas en estudios conductuales para explorar los efectos de la edad sobre estos procesos.

Es justo reconocer que, en el interés de abarcar la mayor cantidad de sistemas neurotransmisores posibles (aminoácidos, aminas biógenas y acetil colina) se hizo un diseño experimental que requería la recogida de muestras por períodos de tiempo relativamente largos. Eso, por una parte, puede “diluir” cualquier variación aguda provocada por el reforzamiento conductual o la estimulación, y por otra, obligó a emplear

frecuencias bajas de estimulación de la amígdala, muy diferentes a las utilizadas en los estudios anteriores. Tal vez por esas razones los resultados no arrojaron las respuestas esperadas y, en algunos casos hicieron aún más compleja la interpretación.

Las variaciones detectadas durante la estimulación de la amígdala mostraron un aumento en los niveles acetil colina y una disminución de noradrenalina y serotonina. Más sorprendente aún, el reforzamiento conductual disminuye los niveles de glutamato y GABA.

No obstante estos problemas, el trabajo tiene valor por cuanto muestra una tabla comparativa de los niveles basales de neurotransmisores y metabolitos asociados entre ratas jóvenes y ratas viejas con deficiencias cognitivas a partir de muestras directas de fluido cerebral y cuantificadas con las técnicas más modernas disponibles.

Otro resultado interesante fue que las ratas viejas no mostraron alteración alguna de los niveles de ninguna de las sustancias estudiadas, ni ante la estimulación de la amígdala, ni por efecto del reforzamiento conductual.

Effect of LTP-reinforcing paradigms on neurotransmitter release in the dentate gyrus of young and aged rats

W. Almaguer-Melian^a, R. Cruz-Aguado^a, C. de la Riva^b, K.M. Kendrick^b,
J.U. Frey^c, J. Bergado^{a,*}

^a *International Centre for Restorative Neurology (CIREN), Havana, Cuba*

^b *Cognitive and Behavioural Neuroscience, The Babraham Institute, Cambridge, UK*

^c *Leibniz Institute for Neurobiology, Magdeburg, Germany*

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Abstract

Long-term potentiation (LTP) is considered a cellular correlate of memory processing. A short-lasting early-LTP can be prolonged into a late-LTP (>4 h) by stimulation of the basolateral amygdala (BLA) or motivational behavioral stimuli in young, but not in aged, cognitively impaired rats. We measured the changes in transmitter release-induced by BLA or behavioral reinforcement—in young and aged cognitively impaired rats, after implanting a microdialysis cannula at the dentate gyrus. Samples were taken under baseline conditions and during stimulation of BLA. Rats were water deprived and tested again next day, taking samples after allowing access to water. Higher concentrations of choline, HIAA, aspartate, glutamate, and glycine were found in baseline samples from young animals compared to aged. In young animals, BLA stimulation increased the levels of ACh and reduced nor-epinephrine and serotonin, while behavioral reinforcement reduced the levels of glutamate and glycine. These effects were absent among aged rats, suggesting that this reduced neurochemical response might be linked to the impaired LTP-reinforcement reported previously.

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Keywords: LTP; Amygdala; Behavioral reinforcement; Neurotransmitters; Microdialysis

The hippocampus is a cortical structure involved in explicit and spatial memory in rodents and humans [19,39,48]. The inputs to this limbic structure are highly organized according to origin and neurotransmitter system. An excitatory-glutamatergic input reaches the dentate gyrus, and other hippocampal subfields, via the perforant pathway (PP), while other, mainly modulatory, transmitters arising in the medial septum and brain stem nuclei converge in the fimbria–fornix system (FF) to enter the hippocampal formation [37,58]. The latter include acetylcholine (ACh), γ -aminobutyric acid (GABA), norepinephrine (NA), dopamine (DA), and serotonin (5-HT), among others.

Long-term potentiation (LTP)—first described in the early 1970s [13,14]—represents a long-lasting increase in synaptic efficacy and is considered a cellular correlate of memory [15,31,33,34]. In the dentate gyrus, the CA1 subfield, and other brain regions, LTP is initiated by the activation of NMDA–glutamatergic receptors which allows Ca^{2+} to enter the postsynaptic terminal [7]. The Ca^{2+} -mediated activation of protein kinases, like the calcium–calmodulin-dependent kinase (CaM-kinase), provokes local changes in receptor proteins, like the AMPA–glutamatergic receptor, which are considered to be responsible for the initial increase in synaptic efficacy [27,36,54]. At later stages, LTP requires the synthesis of new proteins for its maintenance [25,30,38].

Based on this background, a two phase model of LTP has been proposed. An early, protein synthesis

* Corresponding author. Fax: +53 7332420.

E-mail address: bergado@neuro.ciren.cu (J. Bergado).

independent phase, lasting for about 4 h (E-LTP), is followed by a longer-lasting, protein synthesis-dependent late-LTP (L-LTP) [30,32].

While glutamatergic synapses appear able to induce LTP (instructive synapses), this does not preclude a cooperative action of non-glutamatergic afferents to the same population (modulatory synapses) [31]. Such modulation appears to be of particular importance for L-LTP. In the CA1, a role for DA has been well documented [23,24,26,43]. In the dentate gyrus, behavioral manipulations known to be associated with the activation of modulatory inputs are able to prolong an E-LTP induced by a mild tetanus to the PP, into a L-LTP [47,53].

Our group has studied the neural and neurochemical mechanisms involved in LTP reinforcement. The basolateral amygdala (BLA) seems to play a key role because its temporal or permanent inactivation abolishes reinforcement [3]. Moreover, stimulating the BLA can produce a similar reinforcing effect to that seen with behavioral stimuli [22]. This effect is mediated by the FF system [28]. Pharmacological studies have consistently reported a role for NA [22,52], however, the results for ACh have been contradictory. While atropine blocks the reinforcing effects of BLA-stimulation [22], it does not influence the reinforcing effects of stimulating the medial septum [21].

All these studies have been carried out using pharmacological approaches, i.e., injecting antagonists in the cerebral ventricles, thus raising the possibility that the applied substances might be exerting their effects on the dentate gyrus indirectly through other brain structures.

We carried out the present study in an attempt to obtain direct evidence concerning which transmitter systems might be involved in both behavioral and BLA-stimulation reinforcement of E-LTP. We used *in vivo* microdialysis sampling to measure neurochemical release in the dentate gyrus under both reinforcing paradigms.

A main goal of our study was to compare the results obtained in young animals with those of aged, cognitively impaired rats. It is known that the main difference in LTP between young and aged rats is the faster decay, and consequent shorter duration, of LTP [4,5]. In line with this, our group has already shown that reinforcing paradigms are impaired in aged rats which have demonstrable memory deficits [2,9]. Changes in the pattern of activation of modulating inputs under both reinforcing paradigms could be responsible for their reduced efficacy.

Materials and methods

Animal housing and handling. Male Sprague–Dawley rats obtained from a local professional breeder (CENPALAB, Havana) were used.

Young (2–3 months) and aged animals (24–27 months) obtained as retired breeders were kept in plastic translucent cages (five animals per cage) under controlled environmental conditions (temperature, humidity, and light cycle) with free access to water and food throughout the experiment, except for 1 day of water deprivation during the behavioral reinforcement paradigm. All efforts were made to reduce pain or discomfort of the animals under strict observance of the Cuban Regulations for the Use of Laboratory Animals.

Memory test. Young ($n = 6$) and aged animals were first tested for their spatial memory abilities using the Morris Water Maze (MWM) as described elsewhere [1]. The average escape latency of aged rats over the five days of training (8 trials daily) was compared with that of young animals. Aged rats showing values above the mean escape latency plus two standard deviations of the young controls ($n = 6$) were considered as cognitively impaired.

Surgery. Under ketamine narcosis (33.3 mg/kg, ip.) rats were implanted with a microdialysis cannula (CMA/12, Sweden) in the hilus of the dentate gyrus (AP: -3.8 mm, ML: 2.0 mm, DV: -3.5 mm). A stainless steel stimulating electrode was positioned at the BLA (AP: -2.8 mm, ML: 5.0 mm, DV: 8.5 mm). All co-ordinates were calculated from bregma, with bregma and lambda at the same height.

Sample collection. All samples were collected from freely moving animals. The day after surgery rats were placed in a plastic bowl and connected through a swivel to the microinfusion pump (CMA/100, CMA Microdialysis, Sweden). Artificial cerebrospinal fluid (125 mM NaCl, 2.5 mM KCl, 0.5 mM NaH_2PO_4 , 5 mM Na_2HPO_4 , 1 mM $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, 0.2 mM ascorbic acid, and 1.2 mM $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$) was infused at $2 \mu\text{L}/\text{min}$ using a syringe pump (CMA Microdialysis, Sweden). Sample volumes of $90 \mu\text{L}$ fluid were collected over 45 min periods and stored at -80°C until used for analysis.

Stimulation and reinforcing paradigms. After a 1 h habituation to the experimental set-up, three consecutive samples were collected to determine the baseline extracellular concentrations for each transmitter. Low frequency stimulation to the BLA was then delivered at 0.1 Hz (0.1 ms square pulses, $400 \mu\text{A}$) by a SS104-J Isolation Unit coupled to a SEN 3301 Electronic Stimulator (both Nihon Kohden, Japan) for 45 min.

The behavioral reinforcement paradigm was carried out the following day. A 1 h habituation was permitted and two new baseline samples were collected for that day. After 24 h of water deprivation, access to water was allowed and a sample was collected during the following 45 min.

Biochemical analysis. Amino acid levels in dialysates were determined by high performance liquid chromatography (HPLC) with fluorescence detection following automated derivatization (autosampler Gilson 231, Villiers-le-Bel, France) with *o*-phthalaldehyde (OPA). The derivatization reaction was started by mixing $10 \mu\text{L}$ of the sample and $3 \mu\text{L}$ of the OPA solution (35 mM OPA containing 114 mM of 3-mercaptopropanol and 10% methanol in 0.4 M sodium tetraborate buffer, weekly prepared) for 2 min. Ten microliters of the derivatized sample was injected into a $15.0 \text{ cm} \times 3.2 \text{ mm}$ Sphere-Clone ODS(2) reversed phase column with $3 \mu\text{m}$ particle size (Phenomenex, Macclesfield, UK) kept at 38°C . A gradient of eluents at a $0.525 \text{ mL}/\text{min}$ flow rate was accomplished by means of a binary pump (Beckman 126, Fullerton, CA, USA). Eluent A was 39.8 mM sodium acetate with 10% methanol at pH 5.7 and eluent B was methanol with 20% of eluent A at pH 6.7. The gradient started at 23% B, rose to 45% B in 0.5 min, to 70% B at 6.5 min, to 97% B at 10 min, and returned to 23% B at 16 min. The fluorescence detector (CMA/280, CMA Microdialysis, Sweden) was set at an excitation wavelength of 340 nm and an emission wavelength of 495 nm. Chromatograms were recorded and the amino acid content in the samples was quantified by means of the external standard procedure with the 32 Karat software (Beckman, Fullerton, CA, USA).

Monoamines and metabolites were measured by HPLC with electrochemical detection. The detector (Waters, Milford, USA), with an adapted 6-mm glassy carbon cell (Bioanalytical Systems, West

Lafayette, USA), was set at +0.65 V for the oxidation of monoamines and metabolites. The mobile phase contained 10 mM NaCl, 80 mM sodium acetate, 0.8 mM octanesulfonic acid, 0.3 mM EDTA, and 5% methanol (pH 4.5). A flow rate of 0.210 mL/min was set by an isocratic pump (Gynkotek M480, Macclesfield, UK). Compounds were separated in a 15.0 cm $\times$ 2.0 mm SphereClone ODS(2) reversed phase column with 3 μ m particle size kept at 43 °C. A refrigerated autosampler (CMA/200, CMA Microdialysis, Sweden) injected 9.6 μ L of the samples. Monoamine concentrations were calculated with reference to external standards using GynkoSoft software (Gynkotek, Macclesfield, UK).

Acetylcholine concentrations were determined by HPLC with electrochemical detection using a wired electrode system and a microbore column and enzyme reactor (BAS, West Lafayette, USA). The mobile phase consisted of 85 mM NaH₂PO₄, 6.6 mM NaCl, and 0.5% Kathon (antibacterial agent) at pH 8.5. The fluid was pumped at 0.120 mL/min. by an isocratic liquid chromatograph (CMA/250, Stockholm, Sweden) through a UniJet microbore column sized 530 $\times$ 1 mm. At the end of the column, an enzyme electrode kit was attached. This device consisted of an immobilized-enzyme reactor where acetylcholinesterase and choline oxidase catalyzed-reactions produced hydrogen peroxide and a 6-mm glassy carbon electrode with coated horseradish peroxidase, which was replaced weekly. The signal derived from the reduction of the hydrogen peroxide by the peroxidase was detected by an electrochemical detector (LC-4C, BAS, West Lafayette, USA) set at 0 V reducing potential. A refrigerated autosampler (CMA/200) injected 7 μ L of the samples. Chromatograms were recorded and the ACh concentrations in the samples were quantified by means of the external standard procedure with the GynkoSoft software (Gynkotek, Macclesfield, UK).

Statistical analysis. The values measured for each substance under the different stimulation or reinforcing protocols for young and aged rats were compared using Student's *t* test for independent samples.

Within group comparisons between the different stimulation protocols were referred to the baseline of the corresponding day and compared by Student's *t* test for paired samples. Differences were considered significant only if $p < 0.05$.

Results

Table 1 contains the mean baseline concentration for all of the substances measured. Statistical analysis

confirmed significant differences between age groups for some of the measured substances. Basal concentrations of choline, 5-hydroxyindolacetic acid (5-HIAA), aspartate, glutamate, glycine, and GABA were significantly reduced in aged animals. There were similar trends towards age-associated reductions in DA, homovanillic acid (HVA), NA, 5-HT, citrulline, and taurine but these just failed to achieve significance. In the case of ACh a slight increase was apparent among aged rats, but this was associated with a greater variability as indicated by the standard error and was not significant. The reinforcing paradigms induced modifications in the release of several transmitters.

Fig. 1 shows the changes induced by the stimulation of the BLA. ACh release was significantly increased during the electrical stimulation of this structure. On the contrary, the release of NA and 5-HT was significantly reduced. Other transmitters and metabolites showed no significant alterations.

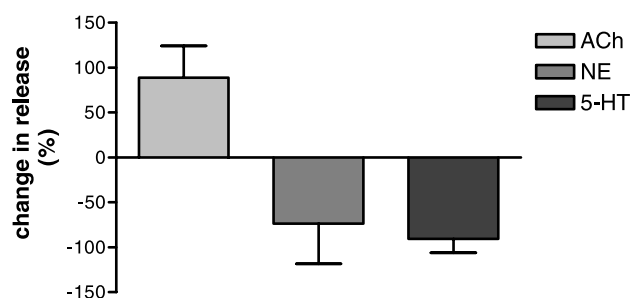


Fig. 1. Changes in the concentration of transmitter substances (% with respect to basal release) at the dentate gyrus during low frequency electrical stimulation of the basolateral amygdala. ACh, acetylcholine; NE, norepinephrine; and 5-HT, serotonin. Other transmitters or metabolites measured showed no significant changes under this condition.

Table 1

Baseline concentration (average of three consecutive samples) of neurotransmitters and related substances in the dentate gyrus of young and aged rats

Substance (units)	Young (mean $\pm$ SE)	Aged (mean $\pm$ SE)
Acetylcholine (nM)	0.34 $\pm$ 0.04	0.81 $\pm$ 0.39
Choline (nM)	389.57 $\pm$ 55.24* ($t = 2.00$)	209.12 $\pm$ 55.51
Dopamine (pg/ml)	791.48 $\pm$ 390.33	576.18 $\pm$ 199.24
DOPAC (nM) (pg/ml)	11390.28 $\pm$ 3426.65	13944.79 $\pm$ 3271.82
HVA (pg/ml)	3777.00 $\pm$ 553.64	3059.52 $\pm$ 838.17
Norepinephrine (pg/mL)	459.10 $\pm$ 125.67	365.42 $\pm$ 136.76
5-HT (pg/mL)	359.12 $\pm$ 144.74	188.31 $\pm$ 70.46
5-HIAA (pg/mL)	9461.05 $\pm$ 520.26* ($t = 3.26$)	6149.04 $\pm$ 980.43
Aspartate (nM)	129.55 $\pm$ 15.16* ($t = 2.13$)	77.97 $\pm$ 9.19
Glutamate (nM)	915.87 $\pm$ 152.54* ($t = 1.89$)	465.89 $\pm$ 45.23
Citrulline (nM)	287.23 $\pm$ 46.91	175.27 $\pm$ 63.34
Glycine (nM)	1188.19 $\pm$ 192.74* ($t = 2.85$)	318.21 $\pm$ 92.00
Taurine (nM)	3386.96 $\pm$ 480.02	1480.00 $\pm$ 230.47
GABA (nM)	163.98 $\pm$ 63.20* ($t = 1.43$)	13.55 $\pm$ 3.32

* Significant difference between groups, $p < 0.05$, Student's *t* test.

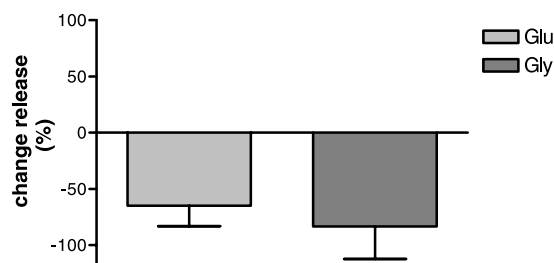


Fig. 2. Changes in the concentration of transmitter substances (% with respect to basal release) at the dentate gyrus after access to water in deprived animals. Glu, glutamate; Gly, glycine. Other transmitters or metabolites measured showed no significant changes under this condition.

The behavioral reinforcing protocol induced a different variation. As shown in Fig. 2, a significant decrease in glutamate and glycine was observed, while other transmitters and metabolites remained unaltered.

Such changes were not observed among aged rats which failed to show any significant variation in transmitter release after BLA stimulation or behavioral reinforcement.

Discussion

Baseline levels: young vs. aged

Brain aging involves not only the loss of neurons and synapses, but also a consequent reorganization in neuronal circuitries [8]. Therefore, the changes in neurotransmitter release are the overall result of the death or atrophy of specific neuronal populations, compensatory synaptic rearrangements, and also the modifications in general metabolism that accompany the aging process. In this work, we observed a decrease in the hippocampal microdialysate concentrations of several molecules related to neurotransmission in aged, cognitively impaired rats. The measurement of compounds in microdialysis samples from brain tissue provides a relative estimate of neurotransmitter levels at the synaptic cleft. However, these measurements can be also influenced by non-synaptic release of transmitters, spontaneous leakage, and inter-cellular exchange of substances for metabolic processes [51]. The latter factor could explain the decrease in the levels of several amino acids that we found in the microdialysates from aged rats, where a decrease in general metabolic activity has been described [11,45]. The reduced glutamate concentration might also be related to the dysfunctional pyramidal cell activity and impaired synaptic transmission in the aged hippocampus, where stereological studies have ruled out major cell loss [17]. Early studies have shown that both glutamate and GABA levels are decreased in Alzheimer's disease brain [20]. This mirrors our results in aged, cognitively impaired rats, a plausible animal model of

Alzheimer's disease. A more recent and detailed analysis of the hippocampus of Alzheimer's patients has revealed impairment in glutamate–glutamine cycle and a tissue redistribution of glutamine synthetase, a glutamate consuming enzyme [42]. In addition, the recently reported age-related decrease in rat hippocampal glutamate decarboxylase isoform 67 [49] could be responsible for the reduced GABA release in the present experiments.

The fact that we failed to find a decrease in the dialysate concentration of ACh in the aged-cognitively impaired rats of the present experiment, while on the contrary, a non-significant tendency to increased levels was apparent, is surprising given the recognized cholinergic dysfunction in the aged brain [35]. Basal dialysate acetylcholine levels have been found decreased in the dorsal hippocampus of aged rats [57]. Possible differences in the neurochemical changes during aging among different hippocampal subfields may account for this discrepancy. However, the recent findings of unimpaired or up-regulated cholinergic markers in the brains of mild Alzheimer's disease patients, which challenge and adds complexity to the traditional cholinergic hypothesis [56] are more compelling.

In addition, our finding of decreased extracellular 5-HIAA levels in the hippocampus of aged rats is consistent with previous reports of impaired 5-HT neurotransmission and metabolism in aged animals [12,41,55]. We failed to observe a corresponding decrease in 5-HT concentrations, perhaps due to the higher turnover of extracellular neurotransmitters compared with neurotransmitter metabolites. Serotonergic activity in hippocampus has been correlated with cognitive performance in rats [50], while neurodegenerative changes in the nucleus raphe dorsalis have been described in Alzheimer's brains [59]. These results underscore a role for serotonergic deficit in age-related brain functional impairment.

BLA induced modifications in young adult rats

The low frequency stimulation of the BLA induced an increase in ACh release and a reduction of NA and 5-HT release in the dentate gyrus of young animals. Previous results have shown that the reinforcing effect on E-LTP of stimulating the amygdala can be blocked by atropine (a muscarinic receptor blocking agent) and propranolol (a β -1 receptor antagonist) [22]. However, it must be considered that, those drugs were applied in the ventricle, thus away from the dentate gyrus (the presumed structure of action) where we have now directly measured transmitter release via microdialysis.

Present results support a direct involvement of cholinergic mechanisms in the reinforcement of E-LTP at the dentate gyrus by stimulation of the amygdala, while the actions of NA and 5-HT appear to be, more likely,

indirect, perhaps on other structures projecting to the dentate gyrus. It must be kept in mind that the stimulation pattern used in this study (0.1 Hz during 45 min) was different from the one used in the mentioned experiments, in which a high frequency was used (3 trains of 15 impulses at 200 Hz). Considering the complex interactions modulating transmitter release in the hippocampus, due to presynaptic inhibition or facilitation [58] the differences in stimulation patterns might lead to differences in the release of neurotransmitters in both conditions.

An increase in NA release at the dentate gyrus after induction of LTP at the dentate gyrus has been reported previously using microdialysis sampling methods [16]. The modulating role of NA from the locus coeruleus on electrophysiological and behavioral processes is well documented (for a review see [10]). Similar effects on LTP have been described for the median raphe-serotonergic inputs [44]. Our results do not disclose a role for those systems on reinforcing mechanisms of LTP at the dentate gyrus, but suggest an indirect action.

BR induced modifications in young adult rats

Reinforcement of an E-LTP into a L-LTP can be obtained by applying behavioral stimuli with a strong motivational value [29,46,47]. Electrophysiologically this effect is quite similar to the reinforcement induced by stimulating the amygdala. In both cases, a longer-lasting LTP, without modifications in amplitude, was obtained. Similar time windows have also been described for both reinforcing paradigms. Moreover, we have demonstrated that the amygdala is a key structure for behavioral reinforcement of LTP [3].

Based on those analogies we had expected to obtain a similar change in the pattern of neurotransmitter release after both, electrical stimulation of the amygdala and behavioral reinforcement. The results have shown, however, completely different neurochemical modifications. No change was observed in ACh, NE or 5-HT release after behavioral reinforcement, but a significant reduction in glutamate and glycine transmission.

This suggests that despite the mentioned analogies, the mechanism of action of behavioral reinforcement might be different to that observed for the amygdala stimulation. A decisive involvement of hypothalamus in the behavioral paradigm seemed likely. Changes in dopamine and acetylcholine release have been measured in the lateral hypothalamus associated with drinking after deprivation [40].

However, we cannot exclude the possibility that the modulatory action of behavioral reinforcement occurs at time points different to that which we have measured here. Further experiments are required to clear these apparent contradictions.

BLA and BR induced modifications in aged, cognitively impaired rats

Aged rats with cognitive impairments showed deficient mechanisms of behavioral and amygdala-stimulation reinforcement of LTP [2,9]. Our results showed that none of the reinforcement protocols induced any change in transmitter release. This might simply be an expression of the reduced basal levels of many transmitters among aged rats caused by the deafferentation of this structure with aging, or may reflect an additional functional defect of those systems that reduce their ability to react in physiological adaptive responses. A similar lack of neurochemical reactivity in aged animals has been observed in other brain regions [18].

LTP impairments with age are mostly related with its maintenance, while the induction mechanisms are better preserved [6]. The impaired neurochemical response demonstrated by aged rats in both situations might be functionally related to the lack of reinforcing effect. Again, it is also possible that we just did not investigate the time window in which the modulatory action takes place.

The growing knowledge on the functional impairments associated with aging and the characterization of their neurochemical correlates might be useful in the development of rational drugs to alleviate such deficits.

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References

- [1] W. Almaguer Melian, J. Jas García, L. Francis Turner, I. Antúnez Potashkina, J. Bergado Rosado, Estudio comparativo de la lesión de fimbria-fornix por aspiración y transección, *Rev. Neurol.* 29 (1999) 704–709.
- [2] W. Almaguer, B. Estupiñán, J.U. Frey, J.A. Bergado, Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP, *Neurobiol. Aging* 23 (2002) 319–324.
- [3] W. Almaguer-Melian, L. Martínez-Martí, J.U. Frey, J.A. Bergado, The amygdala is part of the behavioral reinforcement system modulating long-term potentiation in rat hippocampus, *Neuroscience* 119 (2003) 319–322.
- [4] C.A. Barnes, Plasticity in the aging central nervous system, *Int. Rev. Neurobiol.* 45 (2001) 339–354.
- [5] C.A. Barnes, B.L. McNaughton, An age comparison of the rates of acquisition and forgetting, *Behav. Neurosci.* 99 (1985) 1040–1048.
- [6] C.A. Barnes, G. Rao, B.L. McNaughton, Functional integrity of NMDA-dependent LTP induction mechanisms across the lifespan of F-344 rats, *Learn. Mem.* 3 (1996) 124–137.
- [7] Z.I. Bashir, S. Alford, S.N. Davies, A.D. Randall, G.L. Collingridge, Long-term potentiation of NMDA receptor-mediated

- synaptic transmission in the hippocampus, *Nature* 349 (1991) 156–158.
- [8] J.A. Bergado, W. Almaguer, Aging and synaptic plasticity. a review, *Neural. Plast.* 9 (2002) 217–232.
 - [9] J.A. Bergado, W. Almaguer, J. Ravelo, J.C. Rosillo, J.U. Frey, Behavioral reinforcement of long-term potentiation is impaired in aged rats with cognitive deficiencies, *Neuroscience* 108 (2001) 1–5.
 - [10] C.W. Berridge, B.D. Waterhouse, The locus coeruleus-noradrenergic system: modulation of behavioral state and state-dependent cognitive processes, *Brain Res. Brain Res. Rev.* 42 (2003) 33–84.
 - [11] C. Bertoni-Freddari, P. Fattoretti, U. Caselli, R. Paoloni, W. Meier-Ruge, Age-dependent decrease in the activity of succinic dehydrogenase in rat CA1 pyramidal cells: a quantitative cytochemical study, *Mech. Ageing Dev.* 90 (1996) 53–62.
 - [12] A. Birthelmer, J. Stemmelin, R. Jackisch, J.C. Cassel, Presynaptic modulation of acetylcholine, noradrenaline, and serotonin release in the hippocampus of aged rats with various levels of memory impairments, *Brain Res. Bull.* 60 (2003) 283–296.
 - [13] T.V. Bliss, A.R. Gardner-Medwin, Long-lasting potentiation of synaptic transmission in the dentate area of the unanaesthetized rabbit following stimulation of the perforant path, *J. Physiol. (Lond)* 232 (1973) 357–374.
 - [14] T.V. Bliss, T. Lomo, Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path, *J. Physiol. (Lond)* 232 (1973) 331–356.
 - [15] T.V.P. Bliss, G.L. Collingridge, A synaptic model of memory: long-term potentiation in the hippocampus, *Nature* 361 (1993) 31–39.
 - [16] J.D. Bronzino, P. Kehoe, R. Hendriks, L. Vita, B. Golas, C. Vivona, P.J. Morgane, Hippocampal neurochemical and electrophysiological measures from freely moving rats, *Exp. Neurol.* 199 (1999) 150–155.
 - [17] M.E. Calhoun, D. Kurth, A.L. Phinney, J.M. Long, J. Hengemihle, P.R. Mouton, D.K. Ingram, M. Jucker, Hippocampal neuron and synaptophysin-positive bouton number in aging C57BL/6 mice, *Neurobiol. Aging* 19 (1998) 599–606.
 - [18] A. Del Arco, G. Segovia, F. Mora, Dopamine release during stress in the prefrontal cortex of the rat decreases with age, *Neuroreport* 12 (2001) 4019–4022.
 - [19] H. Eichenbaum, The hippocampus and mechanisms of declarative memory, *Behav. Brain Res.* 103 (1999) 123–133.
 - [20] D.W. Ellison, M.F. Beal, M.F. Mazurek, E.D. Bird, J.B. Martin, A postmortem study of amino acid neurotransmitters in Alzheimer's disease, *Ann. Neurol.* 20 (1986) 616–621.
 - [21] S. Frey, J.A. Bergado, J.U. Frey, Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum, *Neuroscience* 118 (2003) 1055–1062.
 - [22] S. Frey, J.A. Bergado, T. Seidenbecher, H.C. Pape, J.U. Frey, Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala. Heterosynaptic induction of late-LTP, *J. Neurosci.* 21 (2001) 3697–3703.
 - [23] U. Frey, S. Hartmann, H. Matthies, Domperidone, an inhibitor of the D2-receptor, blocks a late phase of an electrically induced long-term potentiation in the CA1-region in rats, *Biomed. Biochim. Acta* 48 (1989) 473–476.
 - [24] U. Frey, M. Krug, R. Brödemann, K. Reymann, H. Matthies, Long-term potentiation induced in dendrites separated from rat's CA1 pyramidal somata does not establish a late phase, *Neurosci. Lett.* 97 (1989) 135–139.
 - [25] U. Frey, M. Krug, K.G. Reymann, H. Matthies, Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro, *Brain Res.* 452 (1988) 57–65.
 - [26] U. Frey, H. Matthies, K.G. Reymann, The effect of dopaminergic D1 receptor blockade during tetanization on the expression of long-term potentiation in the rat CA1 region in vitro, *Neurosci. Lett.* 129 (1991) 111–114.
 - [27] K. Fukunaga, D. Muller, E. Miyamoto, CaM kinase II in long-term potentiation, *Neurochem. Int.* 28 (1996) 343–358.
 - [28] J. Jas, W. Almaguer, J.U. Frey, J. Bergado, Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus, *Brain Res.* 861 (2000) 186–189.
 - [29] V. Korz, J.U. Frey, Stress-related modulation of hippocampal long-term potentiation in rats: involvement of adrenal steroid receptors, *J. Neurosci.* 23 (2003) 7281–7287.
 - [30] M. Krug, B. Lössner, T. Ott, Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats, *Brain Res. Bull.* 13 (1984) 39–42.
 - [31] H. Matthies, *Neuronale Grundlagen der Gedächtnisbildung*, F. Klux, H. Spada, *Enzyklopädie der Psychologie* [6], 15–42, 1998. Göttingen, Hogrefe Verlag für Psychologie. Kognition.
 - [32] H. Matthies, K.G. Reymann, M. Krug, U. Frey, B. Loessner, N. Popov, Multiple stages of posttetanic LTP and memory, in: Rahmann (Ed.), *Fundamentals of memory formation: Neuronal plasticity and brain function*, Gustav Fischer Verlag, Stuttgart, New York, 1989, pp. 395–396.
 - [33] H.J. Matthies, Neurobiological aspects of learning and memory, *Ann. Rev. Psychol.* 40 (1989) 381–404.
 - [34] H.J. Matthies, In search of cellular mechanisms of memory, *Prog. Neurobiol.* 32 (1989) 277–349.
 - [35] E.J. Mufson, S.D. Ginsberg, M.D. Ikonomic, S.T. DeKosky, Human cholinergic basal forebrain: chemoanatomy and neurologic dysfunction, *J. Chem. Neuroanat.* 26 (2003) 233–242.
 - [36] R.A. Nicoll, R.C. Malenka, Expression mechanisms underlying NMDA receptor-dependent long-term potentiation, *Ann. N.Y. Acad. Sci.* 868 (1999) 515–525.
 - [37] J. O'Keefe, L. Nadel, *The Hippocampus as a Cognitive Map*, Oxford University Press, New York, 1978.
 - [38] S. Otani, W.C. Abraham, Inhibition of protein synthesis in the dentate gyrus, but not the entorhinal cortex, blocks maintenance of long-term potentiation in rats, *Neurosci. Lett.* 106 (1989) 175–180.
 - [39] A.J. Parkin, Human memory: the hippocampus is the key, *Curr. Biol.* 6 (1996) 1583–1585.
 - [40] M. Puig de Parada, X. Paez, M.A. Parada, L. Hernández, G. Molina, E. Murzi, Q. Contreras, Changes in dopamine and acetylcholine release in the rat lateral hypothalamus during deprivation-induced drinking, *Neurosci. Lett.* 227 (1997) 153–156.
 - [41] G. Richter-Levin, M. Segal, Age-related cognitive deficits in rats are associated with a combined loss of cholinergic and serotonergic functions, *Ann. N.Y. Acad. Sci.* 695 (1993) 254–257.
 - [42] S.R. Robinson, Neuronal expression of glutamine synthetase in Alzheimer's disease indicates a profound impairment of metabolic interactions with astrocytes, *Neurochem. Int.* 36 (2000) 471–482.
 - [43] S. Sajikumar, J.U. Frey, Late-associativity, synaptic tagging, and the role of dopamine during LTP and LTD, *Neurobiol. Learn. Mem.* 82 (2004) 12–25.
 - [44] A. Sarihi, Y. Fathollahi, F. Motamedi, N. Naghdi, A. Rashidy-Pour, Effects of lidocaine reversible inactivation of the median raphe nucleus on long-term potentiation and recurrent inhibition in the dentate gyrus of rat hippocampus, *Brain Res.* 962 (2003) 159–168.
 - [45] G. Segovia, A. Porras, A. Del Arco, F. Mora, Glutamatergic neurotransmission in aging: a critical perspective, *Mech. Ageing Dev.* 122 (2001) 1–29.
 - [46] T. Seidenbecher, D. Balschun, K.G. Reymann, Drinking after water deprivation prolongs "unsaturated" LTP in the dentate gyrus of rats, *Physiol. Behav.* 57 (1995) 1001–1004.
 - [47] T. Seidenbecher, K.G. Reymann, D. Balschun, A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli, *Proc. Natl. Acad. Sci. USA* 94 (1997) 1494–1499.
 - [48] L.R. Squire, The hippocampus and spatial memory, *Trends Neurosci.* 16 (1993) 56–57.

- [49] D.P. Stanley, A.K. Shetty, Aging in the rat hippocampus is associated with widespread reductions in the number of glutamate decarboxylase-67 positive interneurons but not interneuron degeneration, *J. Neurochem.* 89 (2004) 204–216.
- [50] J. Stemmelin, C. Lazarus, S. Cassel, C. Kelche, J. Cassel, Immunohistochemical and neurochemical correlates of learning deficits in aged rats, *Neuroscience* 96 (2000) 275–289.
- [51] C.O. Stiller, B.K. Taylor, B. Linderoth, H. Gustafsson, A.A. Warsame, E. Brodin, Microdialysis in pain research, *Adv. Drug Deliv. Rev.* 55 (2003) 1065–1079.
- [52] T. Straube, J.U. Frey, Involvement of alfa-adrenergic receptors in protein synthesis-dependent late long-term potentiation (ltp) in the dentate gyrus of freely moving rats: the critical role of the ltp induction strength, *Neuroscience* 119 (2003) 473–479.
- [53] T. Straube, V. Korz, J.U. Frey, Bidirectional modulation of long-term potentiation by novelty-exploration in rat dentate gyrus, *Neurosci. Lett.* 344 (2003) 5–8.
- [54] S.-E. Tan, R.J. Wenthold, T.R. Soderling, Phosphorylation of AMPA-type glutamate receptors by calcium/calmodulin-dependent protein kinase II and protein kinase C in cultured hippocampal neurons, *J. Neurosci.* 14 (1994) 1123–1129.
- [55] H. Tanila, T. Taira, T.P. Piepponen, A. Honkanen, Effect of sex and age on brain monoamines and spatial learning in rats, *Neurobiol. Aging* 15 (1994) 733–741.
- [56] A.V. Terry Jr., J.J. Buccafusco, The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development, *J. Pharmacol. Exp. Ther.* 306 (2003) 821–827.
- [57] L. Trabace, T. Cassano, L. Steardo, C. Pietra, G. Villetti, K.M. Kendrick, V. Cuomo, Biochemical and neurobehavioral profile of CHF2819, a novel, orally active acetylcholinesterase inhibitor for Alzheimer's disease, *J. Pharmacol. Exp. Ther.* 294 (2000) 187–194.
- [58] E.S. Vizi, J.P. Kiss, Neurochemistry and pharmacology of the major hippocampal transmitter systems: synaptic and nonsynaptic interaction, *Hippocampus* 8 (1998) 566–607.
- [59] Y. Yang, H.P. Schmitt, Frontotemporal dementia: evidence for impairment of ascending serotonergic but not noradrenergic innervation. Immunocytochemical and quantitative study using a graph method, *Acta Neuropathol. (Berl)* 101 (2001) 256–270.

Capítulo 9: El reforzamiento conductual de la potenciación sináptica duradera se afecta por el envejecimiento

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El proceso de envejecimiento, tanto en animales como en humanos afecta las capacidades de memoria en todas sus formas y esto se asocia a deficiencias en los mecanismos de neuroplasticidad. También es conocido que el envejecimiento altera el balance afectivo asociándose frecuentemente con ansiedad y depresión.

Nuestro trabajo ha demostrado que los procesos de neuroplasticidad son modulados por factores afectivos mediante mecanismos que involucran a estructuras de las que se conoce, sufren despoblación neuronal severa, como la amígdala y la región septal.

También hemos mostrado que sistemas de neurotransmisión noradrenérgica y colinérgica son mediadores importantes de estos procesos. Ambos sistemas se encuentran afectados como consecuencia del envejecimiento.

Nuestra hipótesis para este trabajo y el que sigue fue: *los mecanismos de reforzamiento de procesos neuroplásticos por factores afectivos se encuentran deteriorados como resultado del envejecimiento.*

En este trabajo exploramos esa hipótesis en el modelo de reforzamiento conductual y en el que sigue lo hicimos mediante estimulación de la amígdala.

Los resultados de este trabajo muestran que, en efecto, en los animales viejos con deficiencias cognitivas los mecanismos de reforzamiento conductual no funcionan de manera adecuada. La causa de esta inoperancia pudiera estar en el deterioro funcional que sufren los sistemas colinérgico y noradrenérgico que demostramos en el estudio de microdiálisis.



Letter to Neuroscience

BEHAVIORAL REINFORCEMENT OF LONG-TERM POTENTIATION IS IMPAIRED IN AGED RATS WITH COGNITIVE DEFICIENCIES

J. A. BERGADO,^a W. ALMAGUER,^a J. RAVELO,^a J. C. ROSILLO^a and J. U. FREY^{b*}

^aInternational Center for Neurological Restoration (CIREN), La Habana, Cuba

^bDepartment of Neurophysiology, Leibniz Institute for Neurobiology, Brennekestrasse 6, P.O. Box 1860, 39008 Magdeburg, Germany

Key words: long-term potentiation, aging, reinforcement, dentate gyrus, learning, cognitive impairment.

Behavioral stimuli with emotional/motivational content can reinforce long-term potentiation in the dentate gyrus, if presented within a distinct time window. A similar effect can be obtained by stimulating the basolateral amygdala, a limbic structure related to emotions. We have previously shown that aging impairs amygdala–hippocampus interactions during long-term potentiation. In this report we show that behavioral reinforcement of long-term potentiation is also impaired in aged rats with cognitive deficits. While among young water-deprived animals drinking 15 min after induction of long-term potentiation leads to a significant prolongation of potentiation, cognitively impaired aged rats are devoid of such reinforcing effects. In contrast, a slight but statistically significant depression develops after drinking in this group of animals. We suggest that an impaired mechanism of emotional/motivational reinforcement of synaptic plasticity might be functionally related to the cognitive deficits shown by aged animals. © 2001 IBRO. Published by Elsevier Science Ltd. All rights reserved.

Since its discovery, long-term potentiation (LTP) has been the most popular cellular model of learning and memory formation (Bliss and Collingridge, 1993; Krug et al., 1990). Various reports have shown that LTP is negatively influenced by aging, similarly to memory (Barnes, 1979; Bergado et al., 1997a,b).

In the hippocampus and the dentate gyrus, LTP is normally induced by tetanic stimulation of glutamatergic afferents (Collingridge, 1992). However, a role of hetero-

synaptic, non-glutamatergic afferents on late phases of LTP is gaining increased recognition (Bergado Rosado and Almaguer Melian, 2000; Frey and Morris, 1997; Matthies, 1998). Heterosynaptic influences of this kind may mediate the recently described behavioral reinforcement of LTP (Seidenbecher et al., 1997). In this study, behavioral stimuli, with strong emotional/motivational content, applied within a defined time window after the induction of a protein synthesis-independent short-lasting early-LTP (EARLY-LTP < 4 h) were able to prolong it into a protein synthesis-dependent late-LTP (LATE-LTP > 4 h).

In a recent study we have shown that the stimulation of the amygdala can affect LTP in the dentate gyrus in a similar reinforcing manner (Frey et al., 2001). The amygdala is considered as an important neurological substrate of emotions (LeDoux, 1993) and is known to enhance memory consolidation (McGaugh et al., 1996), suggesting that the similarities between both of these types of reinforcements, i.e. behavioral and amygdala-dependent, might be causal, not casual.

The amygdala–hippocampus interactions on synaptic plasticity are likely mediated via septo-hippocampal afferents as the lesion of the fimbria-fornix abolishes the effects of amygdala stimulation on LATE-LTP (Jas et al., 2000). We have recently studied amygdala–hippocampus interactions in aged rats and found that aging severely impairs such interactions (Almaguer et al., 2001). It appears, therefore, interesting to study whether aging affects the behavioral reinforcement of LTP.

Early stages of LTP were similar between young and aged, cognitively impaired rats (see Experimental Procedures; Fig. 1A). Both groups reach a similar level of potentiation 5 min after tetanization, and the *U*-test showed no significant difference between groups at any time point for the population spike amplitude (PSA). Notably similar results were obtained for the field excitatory postsynaptic potential (EPSP) measured in a single series with a relatively large EPSP component (level

*Corresponding author. Tel.: +49-391-6263422; fax: +49-391-6263421.

E-mail address: frey@ifn-magdeburg.de (J. U. Frey).

Abbreviations: EARLY-LTP, protein synthesis-independent LTP (< 4 h); EPSP, excitatory postsynaptic potential; LATE-LTP, protein synthesis-dependent LTP (> 4 h); LTP, long-term potentiation; PSA, population spike amplitude.

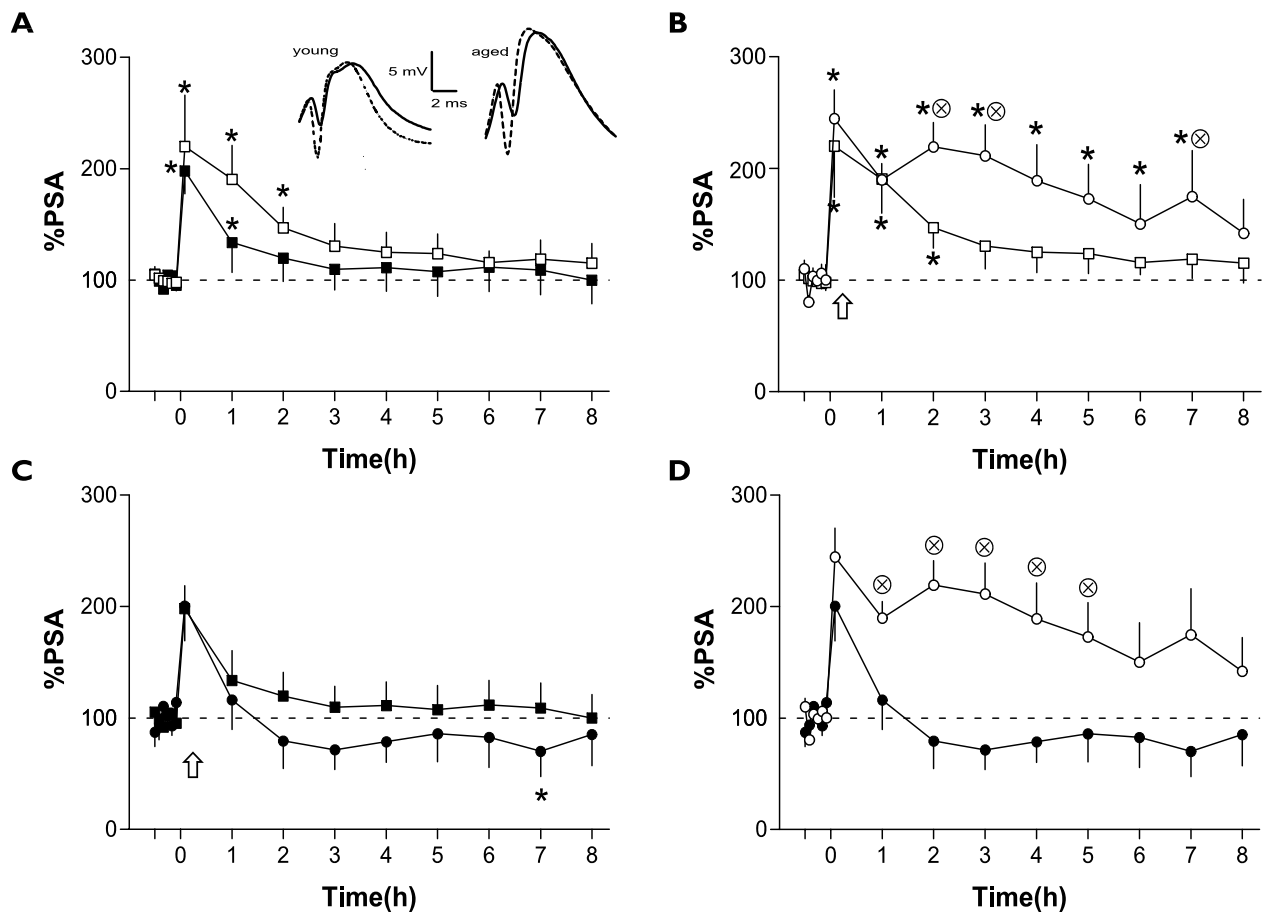


Fig. 1. Time course of LTP, expressed as the relative magnitude of the PSA (%PSA) to baseline value. Each time point is represented by the mean PSA and the S.E.M. Symbols: open squares, control LTP from young animals ($n=6$); filled squares, control LTP from old animals ($n=9$); open circles, behavioral reinforcement from young animals ($n=5$); filled circles, behavioral reinforcement from old animals ($n=6$). The arrows indicate the time at which access to water was allowed in the behavioral reinforcement groups. *Statistically significant differences to baseline values before tetanization within the same group (Wilcoxon test for paired samples, $P < 0.05$). ⊗ Statistically significant differences between groups (U -test, $P < 0.05$). (A) Control LTP in young and aged animals. No significant differences were found between groups, however among groups, LTP seemed to decay faster, according to the Wilcoxon test it remained increased only for the first hour after induction. The inserts show analog traces of recordings taken from a representative young animal (left) before (continuous line) and 5 min after LTP-induction (broken line). The right insert represents traces from an aged cognitively impaired rat, respectively. The amplitude was measured between the first positive and the subsequent negative peak. (B) Behavioral reinforcement of LTP. In young animals, drinking after water-deprivation prolongs LTP, which remained significant up to 7 h after induction. (C) Behavioral reinforcement was absent in old animals, instead a slight depression appears after allowing access to water. (D) Comparative results of behavioral reinforcement between young and old animals. The initial level of LTP does not differ between groups, however significantly higher levels were found among young animals during the subsequent hours (U -test).

of tetanized field EPSP measured as percentage change of the slope 5 min after tetanization: 119.07% in young rats and 118.04% in aged, cognitively impaired rats). The main difference seemed to be the decay rate of LTP. While among the young rats the Wilcoxon test identified a significant difference in the increase of the PSA until 2 h after LTP-induction; aged rats retain LTP only for the first hour (field EPSP as above but 1 h after LTP-induction: 117.21% in young versus 107.91% in aged rats).

Comparing young and aged animals, there were no detectable differences in the input/output characteristics except a slight, statistically non-significant smaller PSA in aged rats.

Allowing young water-deprived rats to drink 15 min after tetanization, does not change the level of synaptic potentiation, but produces a significant lengthening of

LTP (Fig. 1B) up to 7 h after tetanization (Wilcoxon test). The U -test identified statistically significant differences with control LTP at 2, 3 and 7 h (similar results were obtained for the slope of the field EPSP: e.g. % of potentiation in young rats versus motivationally treated young rats: 117.21% versus 121.93% at 2 h and 110.19% versus 100.00% after 7 h). Drinking alone did not influence field potentials in the dentate gyrus as shown by previous results (Seidenbecher et al., 1997).

Such reinforcing effect of a behavioral-motivational stimulus on LTP was absent among aged, cognitively impaired animals (Fig. 1C). The U -test confirmed no significant differences between control LTP and behaviorally reinforced LTP at any time point (examples for the slope of the field EPSP in aged control rats versus motivationally treated aged rats: 99.49% versus 96.86%

Table 1. Results of the behavioral test to select cognitively impaired aged rats

Group	N	Latency (s)			Crossings		
		mean	S.D.	S.E.M.	mean	S.D.	S.E.M.
Young controls	10	23.04	4.38	1.46	8.5	3.24	1.03
Aged impaired	12	45.27 ^a	5.27	1.46	1.42 ^a	1.38	0.39

N: number of animals per group. Values of latency resulting from averaging the data over all trials. Crossings are the group average of the number of times the animals crossed over the area where the platform used to be located.

^aStatistically significant differences between groups (*U*-test, $P < 0.05$; for details see Experimental procedures).

after 2 h and 92.49% versus 84.06% after 7 h). On the contrary, the behavioral stimulus, resulted in a slight tendency towards depression. While control LTP in aged rats is statistically significant up to 1 h after its induction, behaviorally reinforced animals showed only a significant increase 5 min after induction (Wilcoxon test). For the rest of the observation period, behaviorally reinforced animals remained under the baseline level and that reduction was significant at 7 h (Wilcoxon test).

According to this, behavioral reinforcement of LTP was different between young and aged, cognitively impaired rats. As shown in Fig. 1D, significant differences were found from 1 to 5 h after tetanization between those groups (*U*-test).

The results in young rats confirm previous studies reporting an effect of behaviorally relevant stimuli on LTP (Seidenbecher et al., 1995, 1997). Both positive and mild negative reinforcers were able to prolong LTP, while water deprivation itself was shown to have no effect. We have used the same tetanization paradigm which normally induces only an EARLY-LTP, lasting less than 4 h. If this potentiation was associated with drinking 15 min after tetanization, the EARLY-LTP was transformed into a prolonged potentiation. As mentioned previously, the stimulation of the basolateral amygdala within a comparable time window, produces an almost identical result, i.e. an extension of the time span of LTP (Frey et al., 2001). This effect of the amygdala is protein synthesis-dependent and likely to be mediated by heterosynaptic, noradrenergic and cholinergic, afferents (Frey et al., 2001) reaching the hippocampus via the fimbria-fornix (Jas et al., 2000). These results suggest that limbic structures like the amygdala and probably the septum, are parts of the behavioral reinforcement system.

It is well known that aging is very often characterized by cognitive impairments. Impaired synaptic plasticity has been suggested as one of the causes for the age-dependent decline in memory (Barnes, 1993; deToledo-Morrell et al., 1988). While impaired synaptic plasticity is likely to be multicausal, our results suggest that an altered emotional/motivational modulation of late stages of LTP might importantly contribute to that functional deficit.

In previous experiments we have shown that amygdala-hippocampus interactions in synaptic plasticity are impaired in old animals with cognitive impairments (Almaguer et al., 2001). In the present report we extend this observations to show that also behavioral reinforcement of LTP is impaired in a similar way by the aging

process. The behavioral stimulus not only failed to prolong LTP, but seemed to have some slight but long-lasting depressive effect. While the mechanisms of such an effect are unclear, it seems interesting to mention that, in previous control experiments, we have found that stimulation of the amygdala alone induces a slight, but significant long-lasting depression of the PSA (Frey et al., 2001), similar in magnitude and time course to the depression observed in aged rats after behavioral reinforcement.

Emotional alterations are also a major consequence of aging in rodents and humans (Miyagawa et al., 1998; Davidson and Irwin, 1999). We suggest that this age-dependent emotional/motivational impairment might be causally related to cognitive dysfunction because it impairs the reinforcement of ongoing changes in synaptic efficacy.

Structural and functional alterations within the amygdala have been reported as a consequence of aging (Aggleton, 1993; Hyman et al., 1990) and might contribute to the impaired emotional/motivational influence on the late stages of LTP observed in the present study. In addition, the septal cholinergic projection to the hippocampus is reduced during aging (Baxter et al., 1999; Taylor and Griffith, 1993). This modulatory input plays an important role on cognition (Bartus, 2000; Blokland, 1995) and synaptic plasticity (Bergado et al., 1996, 1997a; Buzsáki and Gage, 1989) and is probably involved in the amygdala-hippocampal interactions in LATE-LTP (Jas et al., 2000). We hypothesize that the observed lack of behavioral reinforcement among aged rats with cognitive impairments is due to a dysfunctional amygdalo-septal influence on the dentate gyrus, resulting in the prevention of the modulation of molecular cascades leading to protein synthesis and the reinforcement of EARLY-LTP into LATE-LTP.

These results open new ways to understand the physiological basis of geriatric memory dysfunction and stress the relevance of emotional/motivational factors and their impaired influence on late phases of hippocampal LTP in the chain of alterations that lead to cognitive impairments. Such an approach could also be useful for designing new therapeutic interventions to ameliorate one of the most widespread and invalidating consequences of aging.

EXPERIMENTAL PROCEDURES

Aged Sprague-Dawley male rats (24–27 months at the begin-

ning of the experiments) and a group of young control animals (8 weeks old at the beginning of the experiments) were used. Animals were kept in translucent cages (five animals/cage) with free access to water (except for the 24-h period preceding behavioral reinforcement experiments) and food. Efforts were made to minimize pain or discomfort to the animals, under observance of the Cuban Regulations for the Use of Laboratory Animals.

Aged rats were classified as cognitively impaired according to their results in the Morris water maze. In this behavioral test rats have to learn to find a submerged platform using extra maze cues. Eight trials were performed each day during four consecutive days (60 s maximal search time per trial, 30 s rest at the platform between trials). On day 5 only four trials were performed. The fifth trial on that day was a probe trial in which the platform was removed and the number of crossings over the location, where originally the platform was placed, were counted. The criteria to consider a rat as cognitively impaired was an average latency over the 5 days of training (eight trials per day) exceeding in two standard deviations the average latency of young controls. Since the total number of aged rats without cognitive impairment was too low for a separate group to study, only the group with cognitively impaired animals were used (Table 1). A control test with visible platform revealed no difference between aged and young animals excluding sensory or motor impairments as cause of the longer latencies.

After the end of training the animals were stereotactically implanted, under chloral hydrate narcosis (420 mg/kg), with one recording electrode at the dentate gyrus (coordinates: anteroposterior = -3.8 mm; mediolateral = 2.0 mm; dorsoventral = -3.5 mm, from bregma) and one stimulating electrode at the perforant pathway (coordinates: anteroposterior = -7.5 mm; mediolateral = 4.0 mm; dorsoventral = -4.5 mm, from bregma). In aged animals the recording electrodes had to be inserted slightly deeper than in young animals. Since the implan-

tation was performed under electrophysiological control the final, optimal positioning for the electrodes was evaluated by electrophysiological means. Miniature screws were fixed on the skull and served as reference electrode and earth, respectively. The whole assembly was secured with dental cement. Animals were allowed 1 week recovery from the operation before starting the LTP studies.

Before the electrophysiological experiments, animals were habituated to the recording chamber for at least 4 h. An input/output curve was then constructed and the intensity required for inducing a 40% maximal population spike determined and used for further test recording and induction of LTP.

On the following day a baseline was established, recording every 5 min for 30 min, before inducing LTP with three trains of 15 impulses at 200 Hz. In control LTP experiments, test recordings were performed every 15 min up to 8 h after induction. For behavioral reinforcement experiments, rats were water-deprived for 24 h before inducing LTP with an identical protocol. Fifteen minutes after LTP-induction rats were allowed to drink freely up to the end of the recording period. All deprived animals, young or aged, drank abundantly when the water pipet was placed at the experimental set.

The PSA relative to the baseline average value (expressed as percentage of baseline; % PSA) was used as an indirect measure of synaptic efficacy. In a few experiments with a larger field EPSP we have checked whether this component showed similar time courses as the PS. This was always the case suggesting that the recorded and analyzed PS is not just a measure of changes in excitability. Where possible, i.e. in a few series with a large EPSP component, data of the time course of the field EPSP are given from single experiments.

The Wilcoxon paired samples test was used to confirm significant increases above baseline within groups. For between-groups comparisons the Mann-Whitney *U*-test was used. In both cases a two-tailed significance level of $P < 0.05$ was used.

REFERENCES

- Aggleton, J.P., 1993. The contribution of the amygdala to normal and abnormal emotional states. *Trends Neurosci.* 16, 328–333.
- Almaguer, W., Estupiñán, B., Frey, J.U., Bergado, J., 2001. Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP. *Neurobiol. Aging*, in press.
- Barnes, C.A., 1979. Memory deficits associated with senescence: a neurophysiological and behavioural study in the rat. *J. Comp. Physiol. Psychol.* 93, 74–104.
- Barnes, C., 1993. Electrophysiological changes in hippocampus of aged rodents: predictions for functional change in aged primate. *Neurobiol. Aging* 14, 645–646.
- Bartus, R.T., 2000. On neurodegenerative diseases, models, and treatment strategies: lessons learned and lessons forgotten a generation following the cholinergic hypothesis. *Exp. Neurol.* 163, 495–529.
- Baxter, M.G., Frick, K.M., Price, D.L., Breckler, S.J., Markowska, A.L., Gorman, L.K., 1999. Presynaptic markers of cholinergic function in the rat brain: relationship with age and cognitive status. *Neuroscience* 89, 771–779.
- Bergado Rosado, J.A., Almaguer Melian, W., 2000. Cellular mechanisms of neural plasticity (Spanish). *Rev. Neurol.* 31, 1074–1095.
- Bergado, J.A., Fernández, C.I., Gómez-Soria, A., González, O., 1997a. Chronic intraventricular infusion with NGF improves LTP in old cognitively-impaired rats. *Brain Res.* 770, 1–9.
- Bergado, J.A., Moreno, H., Nuñez, N., 1996. Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biol. Res.* 29, 197–202.
- Bergado, J.A., Moreno, H., Soto, J., Castellano, O., Castillo, L., 1997b. Septal fetal tissue transplants restore long-term potentiation in the dentate gyrus of fimbria-fornix-lesioned rats. *J. Neural Transplant. Plast.* 6, 31–40.
- Bliss, T.V.P., Collingridge, G.L., 1993. A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361, 31–39.
- Blokland, A., 1995. Acetylcholine: a neurotransmitter for learning and memory? *Brain Res. Rev.* 21, 285–300.
- Buzsáki, G., Gage, F.H., 1989. Absence of long-term potentiation in the subcortically deafferented dentate gyrus. *Brain Res.* 484, 94–101.
- Collingridge, G.L., 1992. The mechanism of induction of NMDA receptor-dependent long-term potentiation in the hippocampus. *Exp. Physiol.* 77, 771–797.
- Davidson, R.J., Irwin, W., 1999. The functional neuroanatomy of emotion and affective style. *Trends Cogn. Sci.* 3, 11–21.
- deToledo-Morrell, L., Geinisman, Y., Morrell, F., 1988. Age-dependent alterations in hippocampal synaptic plasticity: relation to memory disorders. *Neurobiol. Aging* 9, 581–590.
- Frey, S., Bergado-Rosado, J., Seidenbecher, T., Pape, H.C., Frey, J.U., 2001. Reinforcement of early long-term potentiation (early-LTP) in dentate gyrus by stimulation of the basolateral amygdala: heterosynaptic induction mechanisms of late-LTP. *J. Neurosci.* 21, 3697–3703.
- Frey, U., Morris, R.G.M., 1997. Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci.* 21, 181–188.
- Hyman, B.T., Van Hoesen, G.W., Damasio, A.R., 1990. Memory-related neural systems in Alzheimer's disease: an anatomic study. *Neurology* 40, 1721–1730.
- Jas, J., Almaguer, W., Frey, J.U., Bergado, J., 2000. Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res.* 861, 186–189.

- Krug, M., Bergado, J., Ruethrich, H., 1990. Long-term potentiation and postconditioning potentiation: the same mechanism? *Biomed. Biochim. Acta* 49, 273–279.
- LeDoux, J.E., 1993. Emotional memory systems in the brain. *Behav. Brain Res.* 58, 69–79.
- Matthies, H., 1998. Neural basis of memory formation (German). In: Klix, F., Spada, H. (Eds.), *Enzyklopädie der Psychologie*, vol. 6. Hogrefe Verlag für Psychologie, Göttingen, pp. 15–42.
- McGaugh, J.L., Cahill, L., Roozendaal, B., 1996. Involvement of the amygdala in memory storage: interaction with other brain systems. *Proc. Natl. Acad. Sci. USA* 93, 13508–13514.
- Miyagawa, H., Hasegawa, M., Fukuta, T., Amano, M., Yamada, K., Nabeshima, T., 1998. Dissociation of impairment between spatial memory, and motor function and emotional behavior in aged rats. *Behav. Brain Res.* 91, 73–81.
- Seidenbecher, T., Balschun, D., Reymann, K.G., 1995. Drinking after water deprivation prolongs ‘unsaturated’ LTP in the dentate gyrus of rats. *Physiol. Behav.* 57, 1001–1004.
- Seidenbecher, T., Reymann, K.G., Balschun, D., 1997. A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proc. Natl. Acad. Sci. USA* 94, 1494–1499.
- Taylor, L., Griffith, W.H., 1993. Age-related decline in cholinergic synaptic transmission in hippocampus. *Neurobiol. Aging* 14, 509–515.

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Capítulo 10: El envejecimiento afecta las interacciones entre hipocampo y amígdala implicados en el reforzamiento de la potenciación sináptica duradera

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En este artículo abordamos la misma hipótesis anterior pero empleando en este caso la estimulación de la amígdala como agente de reforzamiento.

Los resultados demostraron que en los animales viejos con deficiencias cognitivas tampoco esta forma de reforzamiento fue efectiva.

En estos animales se exploraron también formas de plasticidad sináptica de corta duración mediante la técnica de pulsos pareados. No se encontraron diferencias entre animales jóvenes y viejos lo que excluye que deficiencias de los mecanismos básicos de transmisión sináptica sean la causa de la falta de reforzamiento.

En conjunto estos dos trabajos demuestran que los mecanismos de reforzamiento de procesos neuroplásticos por factores afectivos se encuentran afectados en animales viejos que muestran ya deficiencias cognitivas.

Teniendo en cuenta la frecuencia con que deficiencias cognitivas y alteraciones afectivas concurren en individuos envejecidos, estos resultados sugieren que existe una relación disfuncional entre las esferas cognoscitiva y afectiva que hace más grave el deterioro.

Este postulado puede ser importante para entender más cabalmente el deterioro funcional que acompaña al envejecimiento normal o patológico, sobre todo en enfermedades que afectan severamente la esfera cognoscitiva como las demencias.

Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP

William Almaguer^a, Bárbara Estupiñán^a, J. Uwe Frey^b, Jorge A. Bergado^{*, a}

^a International Center for Neurological Restoration (CIREN), Ave. 25 # 15805, Playa 12100, Ciudad de La Habana, Cuba

^b Leibniz Institute for Neurobiology, Brennekesstr. 6, PO Box 1860, D-39008, Magdeburg, Germany

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Abstract

Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP. NEUROBIOL. AGING. We have recently shown that the stimulation of the basolateral nucleus of the amygdala (BLA) is able to prolong early-LTP (<4h) into late-LTP (>4h) in the dentate gyrus. To study whether aging affects this interaction, aged (24–27 months) rats were used, classified as cognitively impaired (I), or non-impaired (N) by means of their results in the Morris water maze. Paired pulses (30–90 ms interval) showed no differences among age groups. Among young controls, the early-LTP induced in the dentate gyrus by stimulation of the perforant path (PP) was prolonged in a late-LTP when the BLA was stimulated 15 min later. In aged-impaired rats the stimulation of the PP induced a reduced LTP, decaying to baseline in less than 2 h. BLA stimulation was without effect. Aged non-impaired rats showed an early-LTP identical to that of young animals; however, stimulation of the BLA showed no effect. These results suggest that deficient synaptic plasticity and memory functions in aged animals might be caused, in part by impaired mechanisms of heterosynaptic reinforcement. © 2002 Elsevier Science Inc. All rights reserved.

Keywords: Aging; Long-term potentiation; Hippocampus; Amygdala; Rat; Cognition; Emotion

1. Introduction

Aging can affect learning and memory, as well as forms of neuronal plasticity, like long-term potentiation (LTP) suggesting that impaired plasticity might be involved in deteriorating processes of memory formation during aging [5,8].

Recent results from our laboratories have shown that the amygdala, a limbic structure considered as a neurological substrate of emotions [27], contributes to the induction of late-LTP. Our cellular studies revealed that stimulation of the basolateral nucleus of the amygdala within a distinct time window is able to reinforce a transient early-LTP (which has, under normal conditions, a duration of less than 4h and is protein synthesis-independent) into the long lasting, protein synthesis-dependent late-LTP in the dentate gyrus. This effect was blocked by cholinergic receptor antagonists [18] and could be abolished by lesioning the fimbria-fornix [25], the major cholinergic input to the hippocampus.

A reduced or dysfunctional cholinergic input to the hippocampus is considered as one of the main factors causing age-related cognitive and plasticity deficits [11,34]. If this input mediates the amygdala effects on hippocampal LTP, it can be hypothesized that aging might impair amygdala-hippocampal interactions in synaptic plasticity.

2. Materials and methods

To this purpose, we have studied LTP at perforant path (PP)-granule cells synapses in aged male Sprague Dawley rats (24–27 months old at the beginning of the experiments) previously classified as cognitively impaired (I) or not impaired (N) animals according to their performance in a spatial memory task (Fig. 1). The behavioral test for classifying aged rats was performed in the Morris water maze. Rats have to learn to find a submerged platform to escape from water. Escape latencies were measured in each of 8 trials during 5 consecutive daily training sessions. The data

* Corresponding author. Tel.: +53-7-216385, +53-7-216844; fax: +53-7-332420.

E-mail address: bergado@neubas.sld.cu (J.A. Bergado).

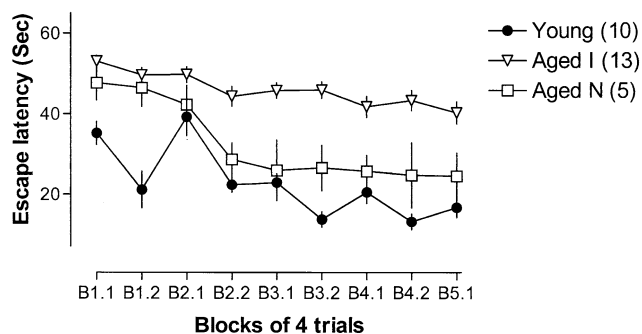


Fig. 1. Results of the behavioral test. Average (mean $\pm$ SEM) latencies to find the platform and escape from water are presented for each of the animal groups, grouped in blocks of 4 trials (2 blocks for each training day). Open squares: young animals ($n = 10$); open triangles: aged rats without cognitive impairment ($n = 5$); filled circles: aged rats cognitively impaired ($n = 13$). For further details see Materials and Methods.

of 4 trials were averaged and presented as blocks (B1.1, B1.2, B2.1, ...); the first number indicates the day of training. Aged rats were considered as cognitively impaired (aged I) if their mean average latency over the 5 days exceeds by 2 standard deviations that of young controls (2 months old animals of the same sex and strain), otherwise they were considered as not impaired (aged N). In previous studies we have confirmed that these differences are not caused by sensorimotor disturbances of the aged rats [15].

One week after behavioral testing a monopolar recording electrode was implanted (under chloral hydrate narcosis, 420 mg/kg), at the hilar region of the dentate gyrus (AP: -3.8 mm, ML: 2.0 mm, DV: -3.5 mm), and bipolar stimulating electrodes were positioned at the perforant path (AP: -7.5 mm, ML: 4.0 mm, DV: 3.9 mm) and the basolateral amygdala (BLA) (AP: -2.4 mm, ML: 5.0 mm, DV: -8.5 mm), respectively (all coordinates to bregma, with bregma and lambda at the same level). The electrodes were connected to a socket and the assembly was fixed to the skull with dental cement (Paladur, Wehrheim, Germany). All electrodes were made from 125 μ m epoxy-isolated stainless steel.

After 7–10 days recovery, but one day previous to the first experiment, the rats were habituated to the recording chamber for at least 4 h. On that day an I/O curve was recorded and from it the intensity needed to obtain a 40% max. population spike was determined and used for test recordings, paired pulse studies and tetanization, i.e. LTP induction.

To evaluate short-term synaptic interactions paired-pulse stimuli were delivered to the PP with interstimulus intervals ranging from 30 to 90 ms. Five consecutive impulses at 0.01 Hz were averaged on line with each interval. The relative amplitude of the second population spike (P2) to the first one (P1) was calculated and expressed as percent (%P2/P1).

Long-term synaptic plasticity studies were started at the following day. A baseline (5 consecutive impulses at 0.01 Hz per time point) were averaged on line every five

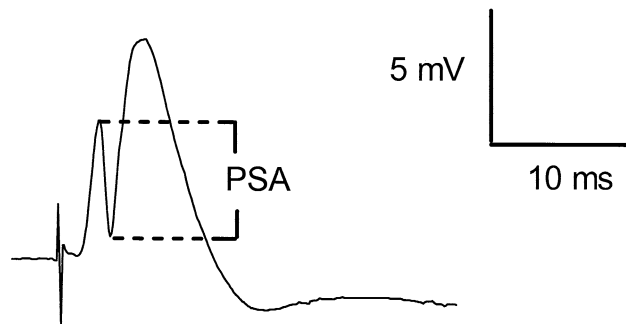


Fig. 2. Representative field potential evoked in the dentate gyrus by stimulation of the perforant pathway taken from an aged, cognitively impaired rat. PSA: population spike amplitude.

minutes) was recorded for 30 min before tetanizing the PP with 3 trains of 15 impulses each (0.1 ms square pulses) at 200 Hz. Five minutes after this first tetanization a record was made to assess the initial level of LTP. Further test recordings were taken every 15 min up to 8 h after tetanization. The population spike amplitude (PSA) was measured (Fig. 2) and its relative value to pre-tetanization baseline values was calculated (%PSA). One week later, after recovery of the baseline values, the same animals were again tetanized using the same protocol. Fifteen minutes after tetanizing the PP, the BLA was stimulated using an equal paradigm. The %PSA was calculated, as the measure of the change in synaptic efficacy, during the following 8 h period.

The animals were randomly assigned to one of the electrophysiological paradigms (i.e. paired pulses, control LTP or BLA stimulation) and some of them were used again for a different protocol under the following conditions: at least seven days delay between studies and a complete recovery of baseline values.

After the end of the study the location of the electrodes was histologically confirmed. Although all animals showed an apparently correct placement of the electrodes according to histology, 2 animals from the aged-impaired group were retired from the BLA-stimulation group because they consistently failed to produce an evoked potential at the dentate gyrus after stimulation of the BLA. This response consists of a 1–2 mV monophasic wave with a latency of 20 ms. Other animals, young or aged included in that study showed this potential, and no age-related differences were observed.

Statistical analysis included the Wilcoxon test for paired samples for within group comparisons relative to baseline values. Between groups comparisons were carried out using the Kruskal-Wallis test for comparing whole curves (i.e. all points in one curve, after LTP induction were compared to all points in the second curve), followed by the Mann-Whitney *U* test to establish significant differences at individual time points. In every test a two-tailed, $P < 0.05$ significance level was used.

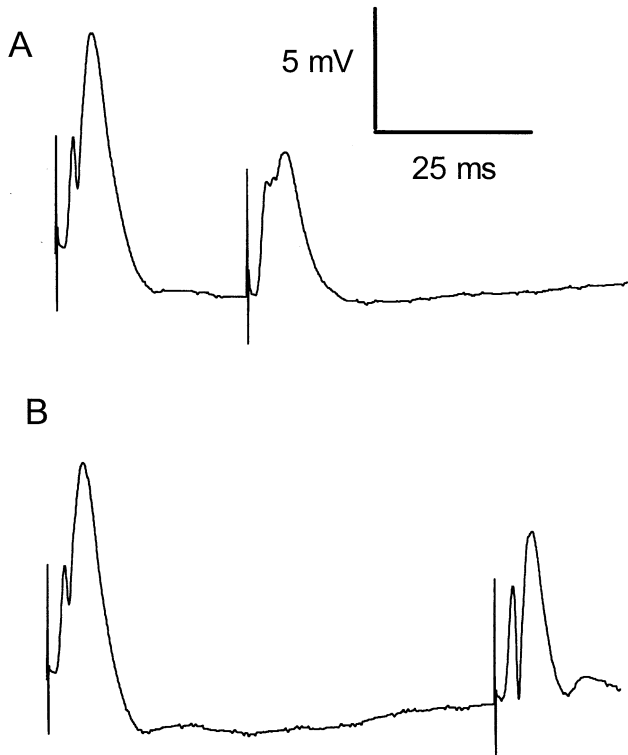


Fig. 3. Representative examples of recordings after paired pulses stimulation of the perforant pathway taken from an aged, cognitively impaired rat. A) 30 ms interstimulus interval. Notice the strong depression of the population spike on the second potential. B) 70 ms interstimulus interval. The second population spike is facilitated.

3. Results

The paired pulse study (Fig. 3) showed the typical relationship of inhibition with short stimulus interval (30 ms) and facilitation at longer intervals (70 and 90 ms). All groups behaved similarly in this study (Fig. 4). Aged rats, whether cognitively impaired or not, showed a similar pattern of inhibition and facilitation as young controls. No significant differences between groups were found at any interstimulus interval (*U* test). The apparent greater variability among aged rats could be a consequence of smaller number of animals per group.

As shown in Fig. 5A, early-LTP induced in the dentate gyrus by stimulation of the PP was reinforced into late-LTP (lasting for more than 4 h) when the BLA was stimulated 15 min after tetanization of the PP. The tetanization of the PP alone induced a significant increase of the population spike that declined to baseline level after two hours (Wilcoxon test). After the sequenced stimulation of the PP and the BLA 15 min later, the increase in synaptic efficacy remained significantly increased during the 8 h recording period (Wilcoxon test). The Kruskal-Wallis showed significant differences between both curves, while the *U* test confirmed significant differences 4 to 7 h after tetanization.

In aged rats with cognitive impairment (Fig. 5B) the stimulation of the PP induced an LTP whose initial value, 5

min after induction, does not differ significantly from that of young animals (*U* test). However, it seemed to decay faster. Statistically significant increases of the PSA were found only during the first hour after tetanization (Wilcoxon test). When the whole curve was compared to that of young animals the Kruskal-Wallis test showed significant differences, strengthening the suggestion of an accelerated decay rate of LTP in the group of aged-impaired rats. Interestingly, the concurrent stimulation of the amygdala does not change the level of synaptic potentiation or its decay rate among these animals. Again, an increase of synaptic efficacy lasting only one hour (Wilcoxon test) was observed. The Kruskal-Wallis test showed no difference to the control LTP curve of the same group, but it differed significantly from that of young animals after BLA stimulation. Significant differences between these two groups were demonstrated at 1 and 4 to 7 h after tetanization (*U* test). This suggests that aging impairs the reinforcing effect of the amygdala on hippocampal LTP.

On the other hand, aged rats without cognitive impairment showed a control LTP that does not differ from that of the young rats (Fig. 5C). The initial level and duration of synaptic potentiation appeared also similar to that of the young animals, i.e. a statistically significant LTP up to 2 h after its induction was observed (Wilcoxon test). The Kruskal-Wallis test showed no differences between this curve and the one corresponding to control LTP of the young rats, but significant differences to aged impaired animals. However, among these animals the stimulation of the amygdala apparently failed to prolong LTP. The period of increased synaptic efficacy remained two hours (Wilcoxon test), the same as in control LTP. The Kruskal-Wallis test found no significant differences between the BLA and control LTP curves among these animals. This finding may suggest that reinforcing effects of BLA stimulation might be failing also in these animals, even before cognitive impairments appear. However, this should be taken with caution because of the small number of animals in this group and the fact that no significant differences were found when

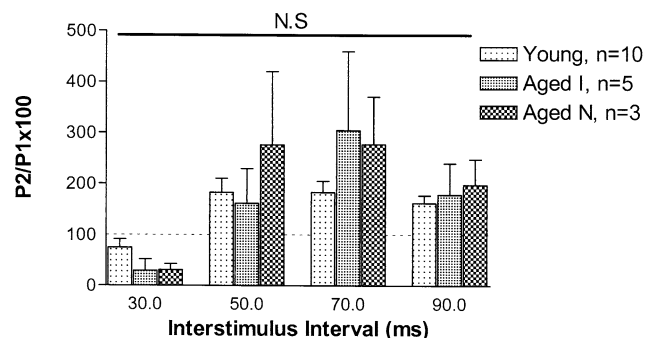


Fig. 4. Results of the paired pulse study in the different age groups. The values of the relative amplitude of the population spike amplitude in the second potential with respect to the first potential ($P2/P1 \times 100$) are shown (mean $\pm$ SEM) for the different groups. N.S.: no significant differences between groups were found with the Mann-Whitney's *U*-test.

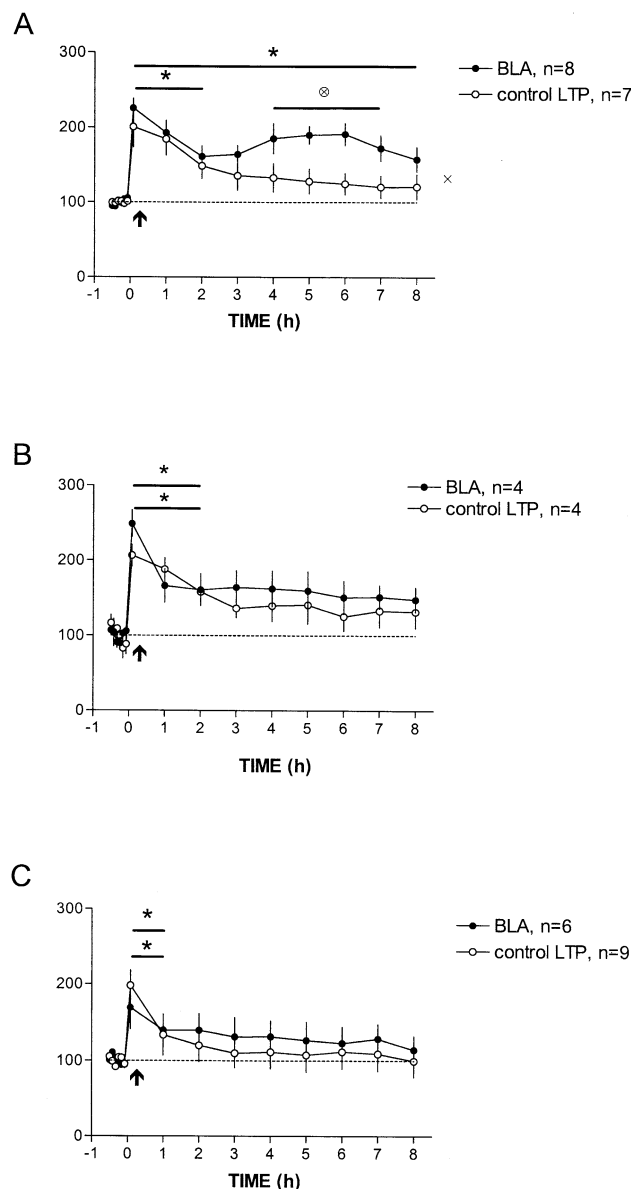


Fig. 5. Amygdala-hippocampus interactions in LTP. The values (mean $\pm$ SEM) of the population spike amplitude, relative to the baseline values (%PSA) are presented. Open symbols: control LTP after tetanization of the perforant path (PP) only; filled symbols: BLA, tetanization of the PP followed by tetanization of the BLA 15 min later. A) Young animals, B) aged rats without cognitive impairment, C) aged rats with cognitive impairments. * significant differences to own baseline ($P < 0.05$ Wilcoxon test for paired samples). The horizontal line below the asterisk indicates how long significant differences were obtained for each curve (inferior line: control LTP, superior line: BLA). X significant differences between curves ($P < 0.05$, Kruskal-Wallis test). $\otimes$, significant differences between groups at the time points indicated ($P < 0.05$, Mann-Whitney's U test). LTP induction by PP tetanization was performed at time 0. The arrow indicate the time of tetanization the BLA.

compared (Kruskal-Wallis) with the curve of young animals after BLA stimulation. In general terms, aged animals without cognitive impairments behave intermediate between young animals and impaired old rats.

4. Discussion

Changes in the excitatory post-synaptic potential (EPSP) are considered by some authors to be a more reliable indicator of modifications in synaptic efficacy. We have, in fact observed a general tendency of the EPSP slope to increase after LTP induction in the present study, however, we have used the population spike amplitude to this purpose instead of the EPSP for several reasons. The recording electrodes were positioned at the hilar region of the dentate gyrus, which is the usual location for chronically implanted electrodes. This is, however, distant from the optimal point to evaluate EPSP's. The size of the granular cell layer in the dentate gyrus makes EPSP-recordings in the intact animal quite unreliable, if compared with the CA1-region. In the hilar region each EPSP is masked by the dipole of the overlaying population spike. Calculating the EPSP from these potentials in freely moving animals is inappropriate because it varies strongly with the behavioral state of the animal. Finally, many LTP studies in slices, both at the CA1 and the dentate gyrus, as well as in vivo studies in narcotized animals in which behavioral influences are absent, have shown that both, the EPSP and the population spike potentiation strongly and positively correlate. We have tested this correlation over a wide range of frequencies to induce LTP and found always a positive correspondence of the EPSP and the population spike potentiation, except for very high frequencies (400 Hz) [28]. For these reasons we considered the PSA as a reliable indicator of changes in synaptic efficacy under our experimental conditions.

The reinforcing effect of activation of the BLA on LTP in the dentate gyrus in young rats is in good agreement with our previous findings [18] and those of other [2,24]. Early-LTP can be reinforced into a late-LTP by heterosynaptic modulatory inputs. As we have shown in those earlier experiments, this process depends on protein synthesis and is blocked by cholinergic and noradrenergic receptor antagonists. Those transmitter systems are potent activators of protein kinases, like PKC and PKA, known to be part of molecular cascades that regulate gene expression and protein synthesis [19], and therefore, essential for late-LTP.

LTP duration appears to be reduced in aged-impaired, but not in aged-not-impaired rats. Comparable results were described in classic LTP studies [4,5,12] and using the model of calcium-induced LTP [13]. The reasons for such different synaptic plastic response are unclear and might involve differences in calcium homeostasis [16,17] or downstream molecular cascades, particularly important for late-LTP. The results of our paired pulse study disclose differences in retroactive inhibition or short-term facilitation as cause of the reduced plasticity shown in LTP experiments.

The reinforcing effect of BLA prolonging LTP, however, was absent in aged rats with cognitive impairment, and likely also in not impaired aged animals. A reduced emotional reactivity has been demonstrated in both, impaired and not-impaired aged rats [30]. The amygdala is consid-

ered to be a key element of an emotional-motivational system [26], which contributes to memory reinforcement in the hippocampus within defined time windows [29,32]. It is tempting to speculate that impairments in emotional reinforcement of LTP occur before cognitive deficits, and that aged non-impaired rats are using some compensatory mechanisms, i.e. an increased excitability of granule cells [31] to overcome the impaired emotional reinforcement. This, however, requires a closer evaluation of cognition and emotion, using a combination of memory tasks, particularly considering the stressful properties attributed to the water maze.

In addition to the results presented here, we had recently shown that septo-hippocampal projections might be involved in the reinforcing effect of the amygdala on LTP [25]. Interestingly, a cooperative interaction of the amygdala and the septum during different learning tasks has been recently described [23]. The fact that the amygdala activates the hippocampus via cholinergic afferents [14] supports the idea that the amygdala-septal-hippocampal projection might be responsible for the reinforcing effect of BLA stimulation on LTP. Such would also be in line with the so called 'cholinergic hypothesis of geriatric memory dysfunction' [6] and the abundant literature demonstrating a link between aging, cholinergic deafferentation to the hippocampus, and cognitive decline [22,33]. Other subcortical inputs arising from brain stem nuclei provide the hippocampus with aminergic innervation. These systems are also able to influence late phases of LTP [21] and are also affected by aging [3] and should be considered along with cholinergic mechanisms to interpret our present data.

Our results suggest that beyond the intrinsic impact that this loss of afferents might produce on hippocampal plasticity [9,10], it could also impair the mechanisms of heterosynaptic modulation of late phases of LTP, like those arising from the amygdala. Both effects could cooperate in causing the cognitive impairments observed in these animals.

This heterosynaptic, associative mechanism may be impaired in aged rats due to neuronal loss and atrophy within the amygdala itself [1], at the septal relay level [11], or both.

The need of a heterosynaptic modulation of late-LTP is becoming increasingly accepted [7] and stresses the relevance of activating cellular cascades, which modulate protein synthesis and an adequate delivery mechanism of these proteins to the activated synapses for late-LTP to occur. The synaptic tagging hypothesis [20] offers a plausible explanation for such mechanisms. We suggest that the failure of reinforcement observed among aged animals could be mainly due to an inadequate or insufficient activation of the molecular cascades, through modulatory inputs, which lead to the synthesis of the plasticity-related proteins. Whether the setting of molecular 'tags' at the activated synapses to capture the newly synthesized proteins is also deficient in aged rats with cognitive impairments should be addressed in future experiments.

The results presented here point to the existence of functional links between two major negative consequences of

aging: reduced emotional/motivational reactivity and impaired cognition. The further study of these processes will open new ways to understand the complex phenomenon of aging and its impact on brain function.

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References

- [1] Aggleton JP. The contribution of the amygdala to normal, and abnormal emotional states. *Trends Neurosci* 1993;16:328–33.
- [2] Akirav I, Richter-Levin G. Priming stimulation in the basolateral amygdala modulates synaptic plasticity in the rat dentate gyrus. *Neurosci Lett* 1999;270:83–6.
- [3] Bach ME, Barad M, Son, H, Zhuo M, LuYF, Shih R, Mansuy I, Hawkins RD, Kandel ER. Age-related defects in spatial memory are correlated with defects in the late phase of hippocampal long-term potentiation in vitro, and are attenuated by drugs that enhance the cAMP signaling pathway. *Proc Natl Acad Sci USA* 1999;96:5280–5.
- [4] Barnes C. Electrophysiological changes in hippocampus of aged rodents: Predictions for functional change in aged primate. *Neurobiol Aging* 1993;14:645–6.
- [5] Barnes CA, McNaughton BL. An age comparison of the rates of acquisition, and forgetting. *Behav Neurosci* 1985;99:1040–8.
- [6] Bartus RT, Dean RL III, Beer B, Lippa AS. The cholinergic hypothesis of geriatric memory dysfunction. *Science* 1982;217:408–417.
- [7] Bergado Rosado JA, Almaguer Melian W. Mecanismos celulares de la neuroplasticidad. *Rev Neurol* 2000;31:1074–95.
- [8] Bergado JA, Fernández CI, Gómez-Soria A, González O. Chronic intraventricular infusion with NGF improves LTP in old cognitively-impaired rats. *Brain Res* 1997;770:1–9.
- [9] Bergado JA, Moreno H, Nuñez N. Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biol Res* 1996;29:197–202.
- [10] Buzsáki G, Gage FH. Absence of long-term potentiation in the subcortically deafferented dentate gyrus. *Brain Res* 1989;484:94–101.
- [11] De Lacalle S, Cooper JD, Svendsen CN, Dunnett SB, Sofroniew MV. Reduced retrograde labelling with fluorescent tracer accompanies neuronal atrophy of basal forebrain cholinergic neurons in aged rats. *Neuroscience* 1996;75:19–27.
- [12] deToledo-Morrell L, Geinisman Y, Morrell F. Age-dependent alterations in hippocampal synaptic plasticity: relation to memory disorders. *Neurobiol Aging* 1988;9:581–90.
- [13] Diana G, Domenici MR, De Carolis AS, Loizzo A, Sagratella S. Reduced hippocampal CA1 Ca^{2+} -induced long-term potentiation is associated with age-dependent impairment of spatial learning. *Brain Res* 1995;686:107–10.
- [14] Dringenberg HC, Vanderwolf CH. Cholinergic activation of the electrocorticogram: an amygdaloid activating system. *Exp Brain Res* 1996;108:285–96.
- [15] Fernández CI, Bergado J, De la Cuétara K, Nunez N, Castellanos O, González O, Torres A, Macías R, Alvarez L, Molina H. Brain aging, and neurotransplantation. II. Effects of septal cell suspension grafts to hippocampus of aged rodents on learning and memory impairments. *Arch Gerontol Geriatr* 1994;18(Suppl. 4):59–65.
- [16] Foster TC. Involvement of hippocampal synaptic plasticity in age-related memory decline. *Brain Res Rev* 1999;30:236–49.

- [17] Foster TC, Norris CM. Age-associated changes in Ca^{2+} -dependent processes: relation to hippocampal synaptic plasticity. *Hippocampus* 1997;7:602–12.
- [18] Frey S, Bergado JA, Seidenbecher T, Pape HC, Frey JU. Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP. *J Neurosci* 2001; 21:3697–703.
- [19] Frey U. Cellular mechanisms of long-term potentiation: late maintenance. In: Donahoe JW, Dorsel VP, editors. *Neural Network Models of Cognition: Biobehavioral Foundations*. Amsterdam: Elsevier Science Press; 1996: in press.
- [20] Frey U, Morris RGM. Synaptic tagging: implications for late maintenance of hippocampal long-term potentiation. *Trends Neurosci* 1997;21:181–8.
- [21] Frey U, Schroeder H, Matthies H. Dopaminergic antagonists prevent long-term maintenance of posttetanic LTP in the CA1 region of rat hippocampal slices. *Brain Res* 1990;522:69–75.
- [22] Gallagher M, Nicolle MM. Animal models of normal aging: Relationship between cognitive decline, and markers in hippocampal circuitry. *Behav Brain Res* 1993;57:155–62.
- [23] Gutierrez H, Gutierrez R, Ramirez-Trejo L, Silva-Gandarias R, Ormsby CE, Miranda MI, Bermudez-Rattoni F. Redundant basal forebrain modulation in taste aversion memory formation. *J Neurosci* 1999;19:7661–9.
- [24] Ikegaya Y, Saito H, Abe K. High-frequency stimulation of the basolateral amygdala facilitates the induction of long-term potentiation in the dentate gyrus in vivo. *Neurosci Res* 1995;22:203–7.
- [25] Jas J, Almaguer W, Frey JU, Bergado J. Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus. *Brain Res* 2000;861:186–9.
- [26] LeDoux JE. Emotional memory systems in the brain. *Behav Brain Res* 1993;58:69–79.
- [27] LeDoux JE. The amygdala: contribution to fear, and stress. *Semin Neurosci* 1994;6:237.
- [28] López Planes J, Almaguer Melian W, Jas García J, Bergado Rosado J. Influencia de la frecuencia de estimulación sobre procesos de plasticidad sináptica en el giro dentado de la rata. *Arch Neurocién (Mex)* 1999;4:9–20.
- [29] McGaugh JL, Cahill L. Interaction of neuromodulatory systems in modulating memory storage. *Behav Brain Res* 1997;83:31–8.
- [30] Miyagawa H, Hasegawa M, Fukuta T, Amano M, Yamada K, Nabeshima T. Dissociation of impairment between spatial memory, and motor function, and emotional behavior in aged rats. *Behav Brain Res* 1998;91:73–81.
- [31] Papatheodoropoulos C, Kostopoulos G. Age-related change in excitability, and recurrent inhibition in the rat CA1 hippocampal region. *Eur J Neurosci* 1996;8:510–20.
- [32] Seidenbecher T, Reymann KG, Balschun D. A post-tetanic time window for the reinforcement of long-term potentiation by appetitive, and aversive stimuli. *Proc Natl Acad Sci USA* 1997;94:1494–9.
- [33] Shen JM, Barnes CA. Age-related decrease in cholinergic synaptic transmission in three hippocampal subfields. *Neurobiol Aging* 1996; 17:439–51.
- [34] Taylor L, Griffith WH. Age-related decline in cholinergic synaptic transmission in hippocampus. *Neurobiol Aging* 1993;14:509–15.

Capítulo 11: La estimulación post-entrenamiento de la amígdala mejora en aprendizaje espacial en ratas con lesión de fimbria-fornix

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Los efectos descritos hasta ahora se han obtenido estudiando solo una forma de neuroplasticidad (plasticidad sináptica del tipo LTP) en una estructura cerebral (el giro dentado del hipocampo) y en animales sanos. Si los mecanismos propuestos para explicar estos efectos estuvieran restringidos solo a esas condiciones, su estudio sería de poco valor teórico y casi ninguno práctico.

En los dos trabajos que se presentan en el final de este documento hemos intentado extender la exploración de estos mecanismos, en un intento de obtener evidencias de su universalidad.

El primero de ellos explora los efectos de la estimulación de la amígdala en animales lesionados, y estudia una forma más general de expresión de la neuroplasticidad: la memoria espacial. La hipótesis que lo guía se puede expresar como:

Si el reforzamiento de procesos neuroplásticos es un fenómeno universal, entonces debe expresarse en otras formas de plasticidad e incluso en animales lesionados.

Utilizamos ratas con lesión de la fimbria fornix que fueron entrenadas en un modelo de memoria espacial utilizando el laberinto acuático diseñado por Morris. Las ratas fueron lesionadas una semana antes y en ese momento se les implantó un electrodo de estimulación en la región basolateral de la amígdala.

La lesión de fimbria fornix provoca un severo déficit en la adquisición de pruebas de este tipo, lo cual se comprueba en todos los grupos controles con lesión. La estimulación diaria de la amígdala, 15 minutos después de concluido el entrenamiento de cada animal, mejoró su aprendizaje, a pesar de la lesión que se les había provocado. Estos animales, a

partir del 3er día de entrenamiento se diferencian de los restantes grupos lesionados y se aproximan al comportamiento de los animales sanos empleados como control.

Este resultado muestra que el reforzamiento de la neuroplasticidad por estimulación de la amígdala no es exclusivo de la LTP y que puede darse también en portadores de lesiones cerebrales.

Post-training stimulation of the basolateral amygdala improves spatial learning in rats with lesion of the fimbria-fornix

William Almaguer^a, Vladimir Capdevila^b, Magaly Ramírez^a, Araceli Vallejo^a, Juan C. Rosillo^a and Jorge A. Bergado^{a,*}

^a*Centro Internacional de Restauración Neurológica (CIREN), Ciudad de La Habana, Cuba*

^b*Instituto de Ciencias Básicas y Preclínicas “Victoria de Girón”, Ciudad de La Habana, Cuba*

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Abstract. *Purpose:* To evaluate the capacity of amygdala stimulation to improve neural plasticity in animals bearing lesions of the fimbria-fornix (FF) system. *Methods:* The animals were lesioned under narcosis (chloral hydrate, 420 mg/kg ip.) using a bilateral transection of the FF procedure. During the same surgery some animals were implanted with an electrode in the right basolateral amygdala (BLA) to allow the electrical stimulation of this structure. Training was carried out one week after surgery using a Morris water maze. Animals were trained in four consecutive days (8 trials/day) in the non-visible platform condition except in the fourth day in which only 4 trials were performed followed by a probe trial in which the escape platform was removed. On day 5 of training 8 trials with visible platform were performed. After each of the first 3 training days one group of animals received trains of electrical stimulation to the BLA, while control groups were not stimulated. A group of non-lesioned animals served as control. The location of the electrode was confirmed histologically after the end of the experiments. *Results:* The learning capacity of the lesioned animals was improved by the electrical stimulation of the amygdala. The latency to find the submerged platform within this group approaches that of the non lesioned animals in the course of training (2-way ANOVA with repeated measures), while other lesioned animals continued to show severely impaired learning abilities. *Conclusions:* This is the first evidence that stimulating the BLA can positively influence the learning abilities of lesioned animals. Further experiments should contribute to improve the stimulation paradigms to make it more effective, if possible.

1. Introduction

Plasticity is a common property of the Nervous System that can be expressed in several forms, from subtle functional changes to massive growth of neurites, or even neurons (at least in certain regions of the adult brain) [13].

Long-term potentiation (LTP) is one form of plasticity characterized by a lasting change in synaptic efficacy

that is induced after tetanic stimulation of specific afferents [16]. According to its duration and the mechanisms involved two phases can be distinguished in LTP [41, 60]. An initial early-LTP (E-LTP) lasting less than 4 hours is followed by a protein synthesis-dependent late-LTP (L-LTP) lasting more than 4 hours [29,30].

We have recently shown that an E-LTP in the dentate gyrus, induced by a weak tetanus to the perforant pathway, can be reinforced and converted into a L-LTP by the stimulation of the basolateral amygdala within a 30 minutes time window [28]. The effect is protein synthesis dependent, suggesting a real conversion of the phenomenon into an L-LTP. This suggests that the

*Corresponding author: Dr. Jorge A. Bergado, Centro Internacional de Restauración Neurológica (CIREN), Ave. 25 # 15805, 11300 Playa, Ciudad de La Habana, Cuba. Tel.: +53 7 2716844; 2716841; Fax: +53 7 332420; E-mail: Bergado@neubas.sld.cu.

amygdala activity is able to stimulate the cellular cascades involved in L-LTP in other brain regions.

However, LTP as induced in the laboratory is a localized form of neural plasticity restricted mainly to the stimulated synaptic population. We were, therefore, interested in testing whether amygdala stimulation might also be able to support more widespread forms of neural plasticity.

The amygdala is able to modulate memory storage in other brain regions, like the hippocampus [11, 39, 47, 52, 54] providing thus a functional link between affective and mnemonic processes [2, 46], and suggesting that neural plasticity might be influenced by emotional/motivational factors. Memory formation is perhaps the most widespread and common form of expression of neural plasticity. Memory may depend on plastic mechanisms at systems and cellular level. LTP is considered a cellular mechanism of memory [49], and the fact that both, memory and LTP can be reinforced by amygdala-depending emotional factors seems to reflect a functional relationship and not merely a coincidence.

The amygdala is a key structure in emotional behavior, motivation and emotional memory [43, 44] in rodents and humans [3, 21, 71]. An impaired amygdala function is likely to be involved in several psychiatric/neurological conditions affecting mood and memory, like Alzheimer's disease, autism and depression [4, 12, 35, 38, 65]. However, all animal studies linking amygdala, memory and synaptic plasticity have been performed in healthy animals, but it is not known whether similar actions can be demonstrated in animals bearing brain lesions.

The fimbria-fornix (FF) fiber system is an important projection system to the hippocampus. The FF provides the hippocampal formation with aminergic terminals arising in brain stem nuclei, as well as a strong cholinergic projection from the basal forebrain [69]. It has been extensively documented that lesions of the FF provoke a severe and long lasting impairment in spatial learning [5, 17, 19, 22, 31, 68]. It has been previously shown that such impairment shows no spontaneous recovery over several months [10, 20].

Such a lesion model appears, therefore, suitable to study whether the stimulation of the amygdala is capable of improving memory in lesioned animals.

Based on these antecedents we have designed a first experiment to test the hypothesis that the stimulation of the basolateral nucleus of the amygdala can be beneficial for improving the learning abilities impaired by a specific brain lesion. The results that will be presented provide preliminary evidences that this is indeed possible.

2. Materials and methods

2.1. Animals

Male Sprague-Dawley rats weighing 250–300 g at the time of surgery were used. The animals were obtained from professional breeders (CENPALAB, Havana) and housed in translucent plastic cages (5 animals per cage) under controlled environmental conditions (temperature, humidity, light-dark cycle) with free access to water and food (Rat Chow, CENPALAB, Havana) throughout the experiment.

2.2. Surgery

For surgery, the animals were anaesthetized with chloral hydrate (420 mg/kg) and mounted in a stereotaxic frame. Hair and skin of the scalp were removed and the points for trepanation were marked on the skull. The procedure for FF lesion has been described elsewhere [37]. Briefly, a bilateral window was opened at coordinates: anterior-posterior (AP) = −1.4 mm; medial-lateral (ML) from 0.5 to 5.2 mm (all coordinates from bregma). A reduced no. 11 blade was then lowered at AP = −1.4 mm; ML = 0.8 mm and dorsoventral (DV) −5.0 mm tilted 15° from the vertical. The blade was slowly moved 3.1 mm laterally and, after 1 min, extracted from the skull.

A stainless steel electrode was positioned at the coordinates AP = −2.8 mm; ML = 5.0 mm; DV = 8.5 mm, aiming the basolateral part of the amygdaloid nuclear complex (BLA). The electrode was connected to a female socket and fixed to the skull with dental cement. A miniscrew attached to the frontal bone served as return electrode and to anchoring the implant to the skull.

2.3. Training

One week after surgery the animals were trained in the Morris water maze for 5 consecutive days. During the first 4 days the escape platform remained submerged in a definite place in the NE quadrant of the circular pool. In the 5th day, the platform was moved to the SW quadrant and made visible by lowering the water level and applying a dark blue ribbon to its border. Eight trials were performed each day starting randomly from 4 different points. The time spent by the animal to find the platform and escape from water (escape latency) was measured. The animals were allowed to search for the platform during 60 sec in each trial. After that

time, they were gently transferred to the platform. A resting period of 30 sec on the platform was allowed between trials. In the 4th day only 4 ordinary trials were performed. The results of 4 consecutive trials were averaged and presented as blocks, numbered for the day and block (e.g. Block 3.1 corresponds to the first 4 trials of day 3).

A fifth trial with removed platform was performed (probe trial) at the end of the day 4 training session, and the number of crossing over the place where the platform used to be located were counted.

2.4. Experimental groups

According to surgery and the treatment received after training several groups of animals were formed. One group of lesioned animals (FFBLA) received electrical stimulation of the amygdala 15 minutes after ending the behavioral training (3×15 impulses at 200 Hz, 400 μ A). A second group of lesioned animals bearing also the electrodes in the amygdala were handled as the former and connected to the stimulation device, but no stimulus was applied (FFCE). A different group of lesioned and implanted animals returned to their cages after training and were not handled neither stimulated (FFCI). A group of lesioned animals without electrodes (FF) and a non-lesioned control group, both without post-training handling were also included.

2.5. Statistics

A two way ANOVA (Group and Block) with repeated measures was used to evaluate results of the training, both in the non-visible and visible platform conditions. For the probe trial a one-way ANOVA (Group) was performed. A post hoc analysis using the Duncan's test was performed when the ANOVA resulted significant to assess between groups differences. A p value inferior to 0.05 was considered significant.

2.6. Histology

After the end of training the animals were transcardially perfused with formalin under narcosis. The brains were removed and processed (Cresyl violet) for microscopic confirmation of the electrode location, and with acetylcholine esterase histochemistry to confirm the efficacy of the FF-lesion. Only those animals showing a correct electrode placement were included in the final evaluation (see Fig. 4).

3. Results

Figure 1 shows the results of training in the water maze with non-visible platform for the different experimental groups. There is a significant time effect in the reduction of the latencies as a consequence of learning (ANOVA, $F_{6,51} = 22.93$, $p < 0.05$). This is, however, different among the experimental groups. The ANOVA for the group factor showed significant differences between groups ($F_{4,51} = 6.33$, $p < 0.05$). The *post hoc* Duncan's test showed that there were no significant differences between groups for the first block of 4 trials (B1.1). However, during the last four trials of the first day (B1.2) and the second training day (B2.1 and B2.2) all lesioned groups, including those with stimulation of the amygdala, differed significantly from the non-lesioned controls. This shows that lesioned animals are impaired in their spatial learning abilities. After two days of training and electrical stimulation of the amygdala a different pattern appears. Starting in the first block of trials from day 3 (B3.1) and through the remaining trials, the group of animals receiving post-trial stimulation of the amygdala does not differ any more from the non-lesioned controls. They do not differ either from the rest of the lesioned groups, but it should be stressed that all those groups of animals bearing lesion, with the only exception of the BLA stimulated animals, do significantly differ from control until the end of the training period.

The results of the probe trial (Fig. 2) showed that the number of crossings was significantly different between groups ($F_{4,51} = 14.48$, $p < 0.05$). The control animals crossed the area of the previous location of the platform significantly more times than any lesioned group. The group that received stimulation of the amygdala (FFBLA) showed a significantly higher number of crossings than the sham-stimulated group (FFCE).

The last two blocks of trials with visible platform performed at day 5 (Fig. 3) showed no differences between groups (Two way ANOVA with repeated measures, $F_{4,50} = 1.31$, $p > 0.05$).

4. Discussion

The absence of differences in the visible platform condition is an indicator that lesioned animals suffer no major visual or motor impairments that could negatively influence their escape latencies. This is relevant because in the non-visible platform condition the an-

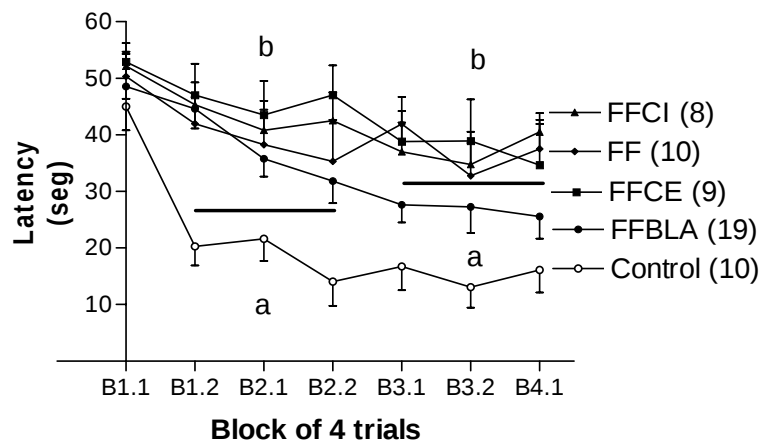


Fig. 1. Results of the training in the Morris water maze (non-visible platform). The mean values ($\pm$ standard error of the mean) of the escape latencies for blocks of 4 trials are given for each experimental group (see legend). In parenthesis are the final numbers of animals in each group. The horizontal lines separate different groups according to the *post hoc* comparison with the Duncan's test ($p < 0.05$). No difference between groups was found for block B1.1. For blocks B1.2 to B2.2 two different groups were discriminated by the Duncan's test. The control animals (a) differed significantly from all other groups (b). From block B3.1 to the end of training controls animals and animals with BLA stimulation do not differ between each other and were separated together (a) from the rest of the experimental groups, above the line (b).

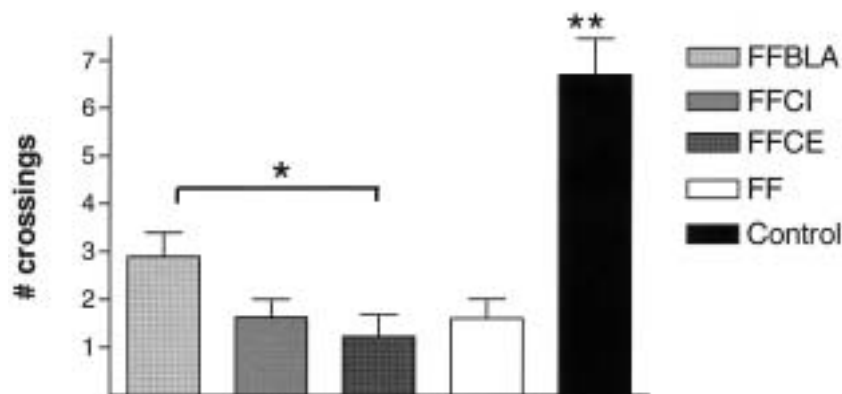


Fig. 2. Average number of crossings (mean $\pm$ SEM) of the experimental groups in the probe trial. The control animals showed significantly more crossings than any other group. The group with BLA stimulation (FFBLA) showed significantly more crossings over the area of former platform location than the sham-stimulated animals (FFCE). ($p < 0.05$ post hoc Duncan's test).

imals have to use visual information from extra maze cues to solve the problem and escape from water. The differences observed in the non-visible platform condition and in the probe trials are, therefore, attributable to an impaired spatial learning ability of the lesioned animals.

Several studies have proven that FF-lesioned animals showed learning impairments in similar tests [7, 20, 45]. These impairments showed no spontaneous recovery over long time periods [10] and have been attributed [58, 68] to the cholinergic denervation at the hippocampus provoked by the lesion. While the probable mediation of other modulatory influences provided to the hippocampus by the FF (i.e. dopaminergic, no-

radrenergic, serotonergic) can not be disclosed, the cholinergic hypothesis of memory dysfunction have inspired a great deal of experimental research showing that the restoration of cholinergic inputs via trophic factors or transplants can benefit learning abilities impaired by FF lesion [14, 18, 23, 24, 26, 42, 67].

Other approaches, however, using repeated training [36] or self stimulation [73] suggest that improvements on learning abilities in FF-lesioned animals can also be induced by non-cholinergic mechanisms.

Our results show that stimulating the BLA 15 minutes after ending each day training session have a positive influence on the learning abilities. This is expressed mainly, in the fact that the behavior of the ani-

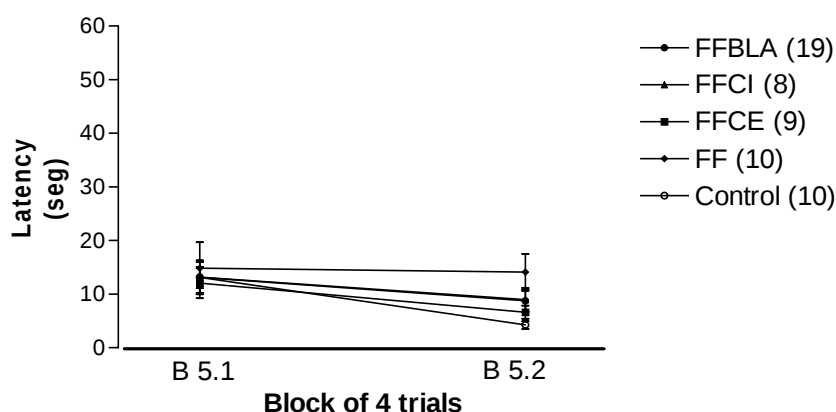


Fig. 3. Results of the training in the Morris water maze (visible platform). The mean values ($\pm$ standard error of the mean) of the escape latencies for blocks of 4 trials are given for each experimental group (see legend). In parenthesis are the final numbers of animals in each group. No significant differences among groups were found (two-way ANOVA).

mals in the FFBLA group progressively tend to be similar to that showed by control, non-lesioned animals. In the probe trial, these animals, although still showing fewer crossings than the control group, have a better retention of the platform location when compared to other of the lesioned groups (FFCE).

The inclusion in the experimental design of lesioned groups bearing electrodes in the amygdala (FFCI) and sham-stimulated (FFCE) disclose any effect of the presence of the electrode at the BLA or the post-training handling of the animals as causal factors for the improvement observed in the FFBLA group, and strongly suggests that the behavioral recovery showed by these animals is due to the electrical activation of the BLA region of the amygdala.

The ability of the amygdala to improve memory by interactions with other brain structures have been extensively documented [15,34,50,51,55]. The mechanisms for the memory-improving effect of amygdala stimulation might involve its reinforcing effect on LTP-like processes in the hippocampus [1,6,8]. LTP-reinforcement by amygdala stimulation is protein synthesis-dependent [28]. Based on pharmacological studies we have hypothesized that the synthesis of plasticity-related proteins might be activated by modulatory afferents (i.e. cholinergic and/or noradrenergic) [28].

We have previously shown that the lesion paradigm used in the present study leads to a severe cholinergic denervation of the dorsal hippocampus which showed no recovery over a period of 3 months [9]. It is therefore unlikely, that the improvement shown by the animals under BLA stimulation might be caused by a re-growth of cholinergic terminals to this structure. An alterna-

tive explanation may arise from the BLA projection to the entorhinal cortex which is the main glutamatergic-excitatory input to the hippocampal formation [40,70]. These projections are reciprocal and may mediate functional interactions between both structures leading to memory consolidation [62,64,66,72]. Our results show that also in animals bearing lesions of the FF, the activation of the amygdala is still able to exert a reinforcing action on memory consolidation.

As previously mentioned, the amygdala is considered a central structure for emotions and motivationally guided behavior [48,53]. There is strong evidence supporting the view that amygdala activation, in a learning context, might represent a functional link between the affective status and memory consolidation [2,61]. This does not mean, however, that the amygdala is the only structure whose activity is able to prolog LTP, enhance memory consolidation or promote memory recovery in lesioned animals. The amygdala is part of a widely distributed network involving other cortical (i.e. the prefrontal cortex) or subcortical structures, whose activation might also contribute to recovery. In line with this, we have previously shown that the stimulation of the septum [27] or the locus coeruleus (Frey S., unpublished results) are also able to reinforce LTP at the hippocampus in healthy animals. Of course, a role for the septum under our experimental conditions is difficult to conceive because the lesion interrupted its main projection to the hippocampal formation.

It remains to be established whether other patterns of amygdala stimulation (different from the ones we used here) can be also effective, or even more effective. This is particularly important regarding the timing of the stimuli (internal frequency within a train, number

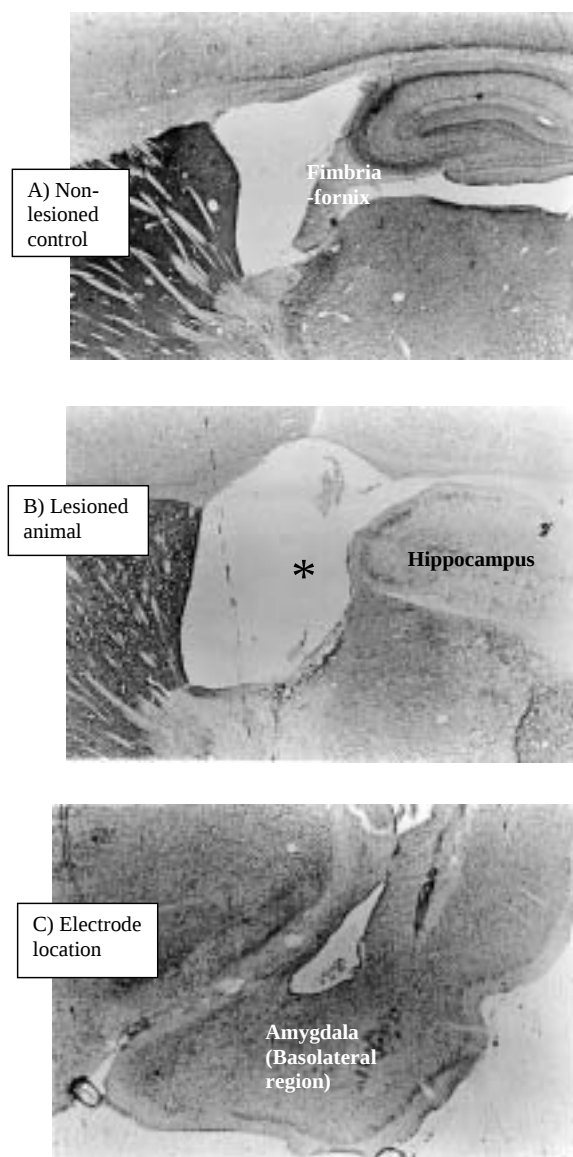


Fig. 4. Histology. A) Low magnification (5x) microphotograph showing the fimbria-fornix system (FF) and the hippocampal formation from a control non-lesioned rat. Notice the intense staining of acetylcholine esterase (ACE) in all hippocampal subfields. B) Low magnification (5x) microphotograph showing the absence of the FF (asterisk) and the severe reduction of ACE staining in the hippocampus. C) Low magnification (5x) microphotograph of the temporal region showing the basolateral amygdala and the electrode track.

of trains, intertrain interval) as well as whether there exist a restricted time window for the effect as is the case with LTP-reinforcement [28].

Further experiments should also contribute to clarify the mechanisms involved in the improving effect shown here. For instance, whether there is an activation of

growth factors. Considering the universality of the mechanisms of neural plasticity [13] it is likely that other forms of plasticity might also be modulated by amygdala-dependent emotional factors. This can also be true for plastic re-arrangements mediating recovery of brain function after trauma or degeneration.

Depression is a common sequel of brain lesion [33, 59] affecting the speed and completeness of recovery [25,57]. A new form for treating depression have emerged in recent years using transcranial magnetic stimulation (TMS) of the dorsolateral prefrontal cortex [32,63]. As there is a link from the prefrontal cortex to the amygdala [56] is likely that both, the improving effect of depressive symptoms and the reinforcement of neural plastic processes may share common mechanisms. Such would open perspectives for the use of TMS in neurological patients to treat their depression and simultaneously, enhance the plastic mechanisms stimulated by the rehabilitation therapy.

The results presented here constitute a first evidence that amygdala stimulation might be a useful tool to stimulate neural plasticity in the lesioned Central Nervous System, and could be useful for future developments in the treatment of chronic neurological diseases. New experiments are required to clarify the possible mechanisms of such effects and to establish the most effective stimulation paradigms to reach that goal.

References

- [1] K. Abe, Modulation of hippocampal long-term potentiation by the amygdala: a synaptic mechanism linking emotion and memory, *Jpn. J Pharmacol.* **86** (2001), 18–22.
- [2] K. Abe, Modulation of hippocampal long-term potentiation by the amygdala: a synaptic mechanism linking emotion and memory, *Jpn. J. Pharmacol.* **86** (2001), 18–22.
- [3] R. Adolphs, L. Cahill, R. Schul and R. Babinsky, Impaired declarative memory for emotional material following bilateral amygdala damage in humans, *Learn. Mem.* **4** (1997), 291–300.
- [4] J.P. Aggleton, The contribution of the amygdala to normal and abnormal emotional states, *Trends Neurosci.* **16** (1993), 328–333.
- [5] J.P. Aggleton, A.B. Keith, J.N. Rawlins, P.R. Hunt and A. Sahgal, Removal of the hippocampus and transection of the fornix produce comparable deficits on delayed non-matching to position by rats, *Behav. Brain Res.* **52** (1992), 61–71.
- [6] I. Akirav and G. Richter-Levin, Mechanisms of amygdala modulation of hippocampal plasticity, *J. Neurosci.* (2002), 9912–9921.
- [7] W. Almaguer Melian, J. Jas García, L. Francis Turner, I. Antúnez Potashkina and J. Bergado Rosado, Estudio comparativo de la lesión de fimbria-fornix por aspiración y transección, *Rev. Neurol.* **29** (1999), 704–709.
- [8] W. Almaguer-Melian and J.A. Bergado Rosado, Interactions between the hippocampus and the amygdala in synaptic plasticity processes. A key to understanding the relations between motivation and memory, *Rev. Neurol.* **35** (2002), 586–593.

- [9] W. Almaguer-Melian, J. Jas Garcia, L. Francis-Turner, I. Antunez Potashkina and J. Bergado-Rosado, Estudio comparativo de la lesión de fimbria-fornix por aspiración y transección, *Rev. Neurol.* **29** (1999), 704–709.
- [10] W. Almaguer-Melian, A. Vallejo, M. Ramírez, V. Capdevila, J.C. Rosillo and J.A. Bergado-Rosado, Estudio comparativo de la lesión bilateral de corteza entorrinal y de la fimbria-fornix, *Rev. Neurol.* **37** (2003), 619–622.
- [11] E.O. Alvarez and M.B. Ruarte, Glutamic acid and histamine-sensitive neurons in the ventral hippocampus and the basolateral amygdala of the rat: functional interaction on memory and learning processes, *Behav. Brain Res.* **152** (2004), 209–219.
- [12] S. Baron-Cohen, H.A. Ring, E.T. Bullmore, S. Wheelwright, C. Ashwin and S.C.R. Williams, The amygdala theory of autism, *Neurosci. Biobehav. Rev.* **24** (2000), 355–364.
- [13] J.A. Bergado Rosado and W. Almaguer Melian, Mecanismos celulares de la neuroplasticidad, *Rev. Neurol.* **31** (2000), 1074–1095.
- [14] J.A. Bergado, H. Moreno, J. Soto, O. Castellano and L. Castillo, Septal fetal tissue transplants restore long-term potentiation in the dentate gyrus of fimbria-fornix-lesioned rats, *J. Neural Transplant. Plast.* **6** (1997), 31–40.
- [15] M. Bianchin, Mello, J.H. Medina and I. Izquierdo, The amygdala is involved in the modulation of long-term memory, but not in working or short-term memory, *Neurobiol. Learn. Mem.* **71** (1999), 127–131.
- [16] T.V. Bliss and T. Lomo, Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path, *J. Physiol. (Lond)* **232** (1973), 331–356.
- [17] M.J. Buckley, D.P. Charles, P.G. Browning and D. Gaffan, Learning and retrieval of concurrently presented spatial discrimination tasks: role of the fornix, *Behav. Neurosci.* **118** (2004), 138–149.
- [18] G. Buzsáki, F.H. Gage, J. Czopf and A. Bjorklund, Restoration of rhythmic slow activity (theta) in the subcortically denervated hippocampus by fetal CNS transplants, *Brain Res.* **400** (1987), 334–347.
- [19] J.C. Cassel, S. Cassel, R. Galani, C. Kelche, B. Will and L. Jarrard, Fimbria-fornix vs selective hippocampal lesions in rats: effects on locomotor activity and spatial learning and memory, *Neurobiol. Learn. Mem.* **69** (1998), 22–45.
- [20] J.-C. Cassel, S. Cassel, R. Galani, C. Kelche, B. Will and L. Jarrard, Fimbria-fornix vs selective hippocampal lesions in rats: Effects on locomotor activity and spatial learning and memory, *Neurobiol. Learn. Mem.* **69** (1998), 22–45.
- [21] R.J. Davidson and W. Irwin, The functional neuroanatomy of emotion and affective style, *Trends Cognit. Sci.* **3** (1999), 11–21.
- [22] J.P. de Bruin, M.P. Moita, H.M. de Brabander and R.N. Joosten, Place and response learning of rats in a Morris water maze: differential effects of fimbria fornix and medial prefrontal cortex lesions, *Neurobiol. Learn. Mem.* **75** (2001), 1640–178.
- [23] E. Duconseille, A. Cressant, C. Kelche, S. Woerly, B. Will, B. Poucet and J.C. Cassel, Homotopic septal grafts combined with a hydrogel bridge promote functional recovery in rats with fimbria-fornix lesions: A unit recording study, *Restor. Neurol. Neurosci.* **15** (1999), 305–317.
- [24] S.B. Dunnett, W.C. Low, S.D. Iversen, U. Stenevi and A. Bjorklund, Septal transplants restore maze learning in rats with fornix-fimbria lesions, *Brain Res.* **251** (1982), 335–348.
- [25] A. Erfurth, M. Loew, G. Wendler and A. Floreanu, [Depressive disorders in neurologic rehabilitation: therapy with paroxetine], *Psychiatr. Prax.* **28** (2001), 43–44.
- [26] L. Francis-Turner and V. Valouskova, Nerve growth factor and nootropic drug Cerebrolysin but not fibroblast growth factor can reduce spatial memory impairment elicited by fimbria-fornix transection: short-term study, *Neurosci. Lett.* **202** (1996), 193–196.
- [27] S. Frey, J.A. Bergado and J.U. Frey, Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum, *Neuroscience* **118** (2003), 1055–1062.
- [28] S. Frey, J.A. Bergado, T. Seidenbecher, H.C. Pape and J.U. Frey, Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP, *J. Neurosci.* **21** (2001), 3697–3703.
- [29] U. Frey, Cellular mechanisms of long-term potentiation: late maintenance, in: *Neural Network Models of Cognition: Biobehavioral Foundations*, J.W. Donahoe, V.P. Dorsel, eds, Elsevier Science Press, Amsterdam, 1997, pp. 105–128.
- [30] U. Frey, M. Krug, K.G. Reymann and H. Matthies, Anisomycin, an inhibitor of protein synthesis, blocks late phases of LTP phenomena in the hippocampal CA1 region in vitro, *Brain Res.* **452** (1988), 57–65.
- [31] R. Galani, S. Obis, E. Coutureau, L. Jarrard and J.C. Cassel, A comparison of the effects of fimbria-fornix, hippocampal, or entorhinal cortex lesions on spatial reference and working memory in rats: short versus long postsurgical recovery period, *Neurobiol. Learn. Mem.* **77** (2002), 1–16.
- [32] M.S. George, Z. Nahas, M. Molloy, A.M. Speer, N.C. Oliver, X.B. Li, G.W. Arana, S.C. Risch and J.C. Ballenger, A controlled trial of daily left prefrontal cortex TMS for treating depression, *Biol Psychiatry* **48** (2000), 962–970.
- [33] H. Ghoge, S. Sharma, S. Sonawalla and R. Parikh, Cerebrovascular diseases and depression, *Curr. Psychiatry Rep.* **5** (2003), 231–238.
- [34] S.B. Hamann, T.D. Ely, S.T. Grafton and C.D. Kilts, Amygdala activity related to enhanced memory for pleasant and aversive stimuli, *Nat Neurosci.* **2** (1999), 289–293.
- [35] S.B. Hamann, E.S. Monarch and F.C. Goldstein, Memory enhancement for emotional stimuli is impaired in early Alzheimer's disease, *Neuropsychology* **14** (2000), 82–92.
- [36] D.K. Hannesson and R.W. Skelton, Recovery of spatial performance in the Morris water maze following bilateral transection of the fimbria/fornix in rats, *Behav. Brain Res.* **90** (1998), 35–56.
- [37] D.K. Hannesson and R.W. Skelton, Recovery of spatial performance in the Morris water maze following bilateral transection of the fimbria/fornix in rats, *Behav. Brain Res.* **90** (1998), 35–56.
- [38] R. Heun, M. Mazanek, K.R. Atztor, J. Tintera, J. Gawehn, M. Burkart, M. Gäsicke, P. Falkai and P. Stoeter, Amygdala-hippocampal atrophy and memory performance in dementia of Alzheimer type, *Dementia* **8** (1997), 329–336.
- [39] N.C. Huff and J.W. Rudy, The amygdala modulates hippocampus-dependent context memory formation and stores cue-shock associations, *Behav. Neurosci.* **118** (2004), 53–62.
- [40] R. Kajiwar, I. Takashima, Y. Mimura, M.P. Witter and T. Iijima, Amygdala input promotes spread of excitatory neural activity from perirhinal cortex to the entorhinal-hippocampal circuit, *J. Neurophysiol.* **89** (2003), 2176–2184.
- [41] M. Krug, B. Lössner and T. Ott, Anisomycin blocks the late phase of long-term potentiation in the dentate gyrus of freely moving rats, *Brain Res. Bull.* **13** (1984), 39–42.

- [42] G. Leanza, G. Nikkiah, O.G. Nilsson, R.G. Wiley and A. Bjorklund, Extensive reinnervation of the hippocampus by embryonic basal forebrain cholinergic neurons grafted into the septum of neonatal rats with selective cholinergic lesions, *J. Comp Neurol.* **373** (1996), 355–357.
- [43] J.E. LeDoux, Emotional memory systems in the brain, *Behav. Brain Res.* **58** (1993), 69–79.
- [44] J.E. LeDoux, The amygdala: contribution to fear and stress, *Semin. Neurosci.* **6** (1994), 237.
- [45] L. Liu, S. Ikonen, T. Heikkinen, M. Heikkilä, J. Puoliväli, T. Van Groen and H. Tanila, Effects of fimbria-fornix lesion and amyloid pathology on spatial learning and memory in transgenic APP+PS1 mice, *Behav. Brain Res.* **134** (2002), 433–445.
- [46] W. Almaguer-Melian and J.A. Bergado-Rosado, [Interactions between the hippocampus and the amygdala in synaptic plasticity processes. A key to understanding the relations between motivation and memory], *Rev. Neurol.* **35** (2002), 586–593.
- [47] K. Majak and A. Pitkanen, Projections from the periamygdaloid cortex to the amygdaloid complex, the hippocampal formation, and the parahippocampal region: a PHA-L study in the rat, *Hippocampus* **13** (2003), 922–942.
- [48] E.J. Maratos, R.J. Dolan, J.S. Morris, R.N. Henson and M.D. Rugg, Neural activity associated with episodic memory for emotional context, *Neuropsychologia* **39** (2001), 910–920.
- [49] H. Matthies, Neuronale Grundlagen der Gedächtnisbildung, in: *Hogrefe Verlag für Psychologie*, F. Klix, H. Spada, eds, Göttingen, 1998, pp. 15–42.
- [50] J.L. McGaugh, Memory consolidation and the amygdala: a systems perspective, *Trends Neurosci.* **25** (2002), 456–461.
- [51] J.L. McGaugh, L. Cahill and B. Roozendaal, Involvement of the amygdala in memory storage: Interaction with other brain systems, *Proc. Natl. Acad. Sci. USA* **93** (1996), 13508–13514.
- [52] J.L. McGaugh, C.K. McIntyre and A.E. Power, Amygdala modulation of memory consolidation: interaction with other brain systems, *Neurobiol. Learn. Mem.* **78** (2002), 539–552.
- [53] H.C. Pape and O. Stork, Genes and mechanisms in the amygdala involved in the formation of fear memory, *Ann. NY Acad. Sci.* **985** (2003), 92–105.
- [54] D. Pare, Role of the basolateral amygdala in memory consolidation, *Prog. Neurobiol.* **70** (2003), 409–420.
- [55] D. Pare, Role of the basolateral amygdala in memory consolidation, *Prog. Neurobiol.* **70** (2003), 409–420.
- [56] D. Paré, D.R. Collins and J.G. Pelletier, Amygdala oscillations and the consolidation of emotional memories, *Trends Cogn. Sci.* **6** (2002), 306–314.
- [57] T. Platz and P. Denzler, Do psychological variables modify motor recovery among patients with mild arm paresis after stroke or traumatic brain injury who receive the Arm Ability Training?, *Restor. Neurol. Neurosci.* **20** (2002), 37–49.
- [58] A.E. Power, A. Vazdarjanova and J.L. McGaugh, Muscarinic cholinergic influences in memory consolidation, *Neurobiol. Learn. Mem.* **80** (2003), 178–193.
- [59] L. Provinciali and M. Coccia, Post-stroke and vascular depression: a critical review, *Neurol Sci.* **22** (2002), 417–428.
- [60] K.G. Reymann, Postsynaptic mechanisms of hippocampal long-term potentiation The involvement of glutamate receptors and protein kinases in its late maintenance, in: *Excitatory Amino Acids*, B.S. Meldrum, F. Moroni, R.P. Simon and J.H. Woods, eds, Raven Press, Ltd., New York, 1991, pp. 475–485.
- [61] G. Richter-Levin, The amygdala, the hippocampus, and emotional modulation of memory, *Neuroscientist* **10** (2004), 31–39.
- [62] G. Richter-Levin and I. Akirav, Amygdala-hippocampus dynamic interaction in relation to memory, *Mol. Neurobiol.* **22** (2000), 11–20.
- [63] P.S. Sachdev, R. McBride, C. Loo, P.M. Mitchell, G.S. Malhi and V. Croker, Effects of different frequencies of transcranial magnetic stimulation (TMS) on the forced swim test model of depression in rats, *Biol. Psychiatry* **51** (2002), 474–479.
- [64] T. Seidenbecher, T.R. Laxmi, O. Stork and H.C. Pape, Amygdala and hippocampal theta rhythm synchronization during fear memory retrieval, *Science* **301** (2003), 846–850.
- [65] T.L. Sweeten, D.J. Posey, A. Shekhar and C.J. McDougale, The amygdala and related structures in the pathophysiology of autism, *Pharmacol. Biochem. Behav.* **71** (2002), 449–455.
- [66] J. Tang, S. Wagner, M. Schachner, A. Dityatev and C.T. Wotjak, Potentiation of amygdaloid and hippocampal auditory-evoked potentials in a discriminatory fear-conditioning task in mice as a function of tone pattern and context, *Eur. J. Neurosci.* **18** (2003), 639–650.
- [67] V. Valouskova and L. Francis-Turner, The short-term influence of b-FGF, NGF and Cerebrolysin on the memory impaired after fimbria-fornix lesion, *J. Neural Transm* **280**(Suppl 47) (1996), 280.
- [68] F.J. van der Staay, W.G. Raaijmakers, A.J. Lammers and J.A. Tonnaer, Selective fimbria lesions impair acquisition of working and reference memory of rats in a complex spatial discrimination task, *Behav. Brain Res.* **32** (1989), 151–161.
- [69] E.S. Vizi and J.P. Kiss, Neurochemistry and pharmacology of the major hippocampal transmitter systems: synaptic and nonsynaptic interactions, *Hippocampus* **8** (1998), 566–607.
- [70] H.O. von Bohlen und D. Albrecht, Reciprocal connections of the hippocampal area CA1, the lateral nucleus of the amygdala and cortical areas in a combined horizontal slice preparation, *Neurosci. Res.* **44** (2002), 91–100.
- [71] P.J. Whalen, S.L. Rauch, N.L. Etcoff, S.C. McInerney, M.B. Lee and M.L. Jenike, Masked presentations of emotional facial expressions modulate amygdala activity without explicit knowledge, *J. Neurosci.* **18** (1998), 411–418.
- [72] D. Yaniv, R.M. Vouimba, D.M. Diamond and G. Richter-Levin, Simultaneous induction of long-term potentiation in the hippocampus and the amygdala by entorhinal cortex activation: mechanistic and temporal profiles, *Neuroscience* **120** (2003), 1125–1135.
- [73] D. Yoganarasimha and B.L. Meti, Amelioration of fornix lesion induced learning deficits by self-stimulation rewarding experience, *Brain Res.* **845** (1999), 246–251.

Capítulo 12: La estimulación de la amígdala mejora la adquisición de una habilidad motora.

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Este trabajo continúa y expande la hipótesis de universalidad de los mecanismos de reforzamiento afectivo de procesos neuroplásticos que, para este, pudiéramos expresar de la siguiente forma: *Si el reforzamiento de procesos neuroplásticos es un fenómeno universal, entonces debe expresarse en otras formas de plasticidad e incluso en otras regiones del encéfalo.*

La forma de neuroplasticidad seleccionada fue la memoria motora que se acompaña de cambios en los mapas motores corticales correspondientes al hemicuerpo y grupo muscular involucrado.

Para su realización empleamos animales sanos a los que se implantó un electrodo de estimulación en la amígdala basolateral. Los animales fueron entrenados en días sucesivos para la adquisición de una habilidad motora hasta alcanzar un criterio previamente definido. El grupo experimental recibió, 15 minutos después de cada sesión de entrenamiento, estimulación eléctrica de la amígdala basolateral, mientras que grupos controles fueron implantados y manipulados de la misma forma pero sin estimulación. Los resultados mostraron que la estimulación de la amígdala redujo el número de sesiones de entrenamiento requeridas para alcanzar el criterio. El entrenamiento por su parte, produjo en todos los animales entrenados una expansión del área cortical de representación de la extremidad anterior. La estimulación de la amígdala no modificó la naturaleza, ni la extensión del cambio cortical; solo hizo que ocurriera más rápido.

Estos resultados están en concordancia con la hipótesis y refuerzan la convicción de que los mecanismos de reforzamiento de procesos neuroplásticos constituyen un mecanismo universal.

Stimulation of the basolateral amygdala improves the acquisition of a motor skill

Jorge A. Bergado^{a,*}, Yeneissy Rojas^b, Vladimir Capdevila^b, Odalys González^a and William Almaguer-Melian^a

^a*Centro Internacional de Restauración Neurológica (CIREN), La Habana, Cuba*

^b*ICBP “Victoria de Girón”, La Habana, Cuba*

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Abstract. *Purpose:* We have previously shown that the stimulation of limbic structures related to affective life such as the amygdala can improve and reinforce neural plastic processes related to hippocampus-dependent forms of explicit memory, as spatial memory and LTP. We now assessed whether this effect is restricted to the mentioned structure and memory type, or represents a more general form of modulatory influence.

Methods: Young, male Sprague Dawley rats were implanted stereotactically with one electrode in the basolateral amygdala (BLA) and trained to acquire a motor skill using their right anterior limb. A group of animals received 3 trains of 15 impulses at the BLA 15 minutes after each daily training session. A second group of implanted animals was handled in the same way, but not stimulated, while a third group was not implanted. After reaching the training criterion the left motor cortex was mapped by the observation of the movements induced by stimuli applied in discrete points of the cortex.

Results: Cortical representation of the anterior limb was increased in all trained animals, showing that the motor cortex is involved in the acquisition of the new skill. Animals receiving stimulation of the BLA showed similar cortical changes, but learned faster than non-stimulated controls.

Conclusions: Reinforcement of neural plasticity by the activation of the amygdala is not restricted to hippocampus-dependent explicit memory, but it might represent a universal mechanism to modulate plasticity.

1. Introduction

Plasticity is one of the most interesting and potentially useful properties of the nervous system and has attracted the interest of researchers and clinicians in the last decades. A large number of studies have shed light on the different forms, mechanisms, and expressions of neural plasticity, as well as their involvement in development, learning, and recovery after different types of trauma [4].

The role played by emotional and motivational factors on learning was recognized, and applied -mostly on empirical terms- since long ago by teachers and trainers. The mechanisms of affective influences on learning have been the subject of a series of studies which have identified the key role played by the amygdala in reinforcing different forms of explicit learning [25, 26]. However, little is known about the potential influences of this limbic structure on other forms of plastic mechanisms.

In collaboration with the group of J. Frey in Germany, we have carried out a series of experiments that demonstrated a positive reinforcing role of the amygdala – in particular of its basolateral region (BLA)- on Long-Term Potentiation (LTP) a widely used model of synaptic plasticity. We have shown that the BLA is

*Corresponding author: Prof. Jorge A. Bergado, Centro Internacional de Restauración Neurológica (CIREN), Ave. 24 # 15805, Cubanacan, Playa 11300, Ciudad de La Habana, Cuba. Fax: +53 7 2732420; E-mail: jorge.bergado@infomed.sld.cu.

involved in reinforcement of LTP by motivational stimuli [3], and that the stimulation of the BLA can effectively substitute the behavioral stimulus to reinforce LTP [11]. This effects seemed to depend on the activation of noradrenergic and cholinergic afferents, and demands the synthesis of proteins, suggesting the mediation of molecular cascades regulating gene expression and macromolecular metabolism [5,11].

In an effort to extend this observations, we have recently shown that post-training stimulation of the BLA can improve the acquisition of spatial memory in fimbria-fornix lesioned rats [2]. This type of memory is hippocampus dependent, as well as the effects on LTP mentioned above.

Therefore, we were now interested in studying whether plastic processes occurring in other structures, like the cerebral cortex, and related to other forms of memory (i.e. implicit memory) like the acquisition of a motor skill, can also be influenced by the stimulation of the amygdala.

2. Materials and methods

2.1. Animals

Young (8 weeks) male Sprague Dawley rats, obtained from the Cuban Center for the Production of Laboratory Animals (CENPALAB) were used. The animals were caged in translucent plastic cages (5 animals per cage) under controlled room conditions (temperature, humidity, light: dark cycle) and with free access to water throughout the experiment. In the groups of animals submitted to training for skilled grasping the diet was restricted to 85% the daily requirements during the training period. All animal handling and experimental procedures were conducted under strict observance of the Cuban Regulations for the Use of Laboratory Animals.

2.2. Surgery

Two groups of animals were operated to implant a stimulating electrode into their right basolateral amygdala. The right BLA was selected to be stimulated in order to avoid damaging the left cortex, in the vicinity of the motor region, that may induce unspecific changes in the motor mapping. Under deep chloral hydrate narcosis (420 mg/kg) the animals were mounted in a stereotactic frame (David Kopf, Saint Louis). The scalp was opened and the skull exposed to drill a hole for inserting

a stainless steel (0.125 mm, Teflon coated) electrode aimed to the basolateral region of the amygdala (coordinates AP: -3.5 mm, ML: 5.0 mm, DV: -7.6 mm from bregma). A miniscrew with a soldered copper wire was screwed on the right frontal bone, to function as returning electrode during stimulation. Both electrodes were connected to a bipolar female socket and attached to the skull with dental cement.

2.3. Motor training

The animals were trained for skilled grasping movements with their right anterior limb using the staircase model introduced by Montoya and coworkers [29] (see Fig. 1A). The food-restricted animal was placed in the cage, upon a central platform surrounded left and right by a 7 steps staircase. In a central depression in each step 2 small pieces of food were placed, only in the right stair. The animals were allowed to pick up and eat all the pieces they can grasp and carry to their mouth using their right anterior limb during a period of 15 minutes every day. The construction of the cage prevented the animals to use their left limb or turning their body to improve reaching and grasping. Once an animal was able to collect all pieces up to level 6 during 3 consecutive days training was stopped and the motor cortex was mapped. The number of days required by each animal to reach this criterion was counted and considered an indicator of its motor learning ability.

2.4. Amygdala stimulation and experimental groups

In one group of animals (BLA, $n = 10$) the basolateral region of the amygdala was stimulated 15 minutes after finishing each training session. The stimulus consisted of 3 trains of 15 impulses (0.1 ms pulse duration) at 200 Hz and 400 μ A intensity. A second group of implanted animals (sham, $n = 8$) was also connected to the stimulus apparatus at the same time, but no stimulus was applied. A third group of control animals (trained, $n = 8$) was simply returned to the home cage at the end of training. Finally, a group of naïve, non-trained and non-stimulated animals ($n = 19$) was used only for mapping the motor cortex. The number of animals given in parentheses is the final amount of rats included in the statistical analysis for each group.

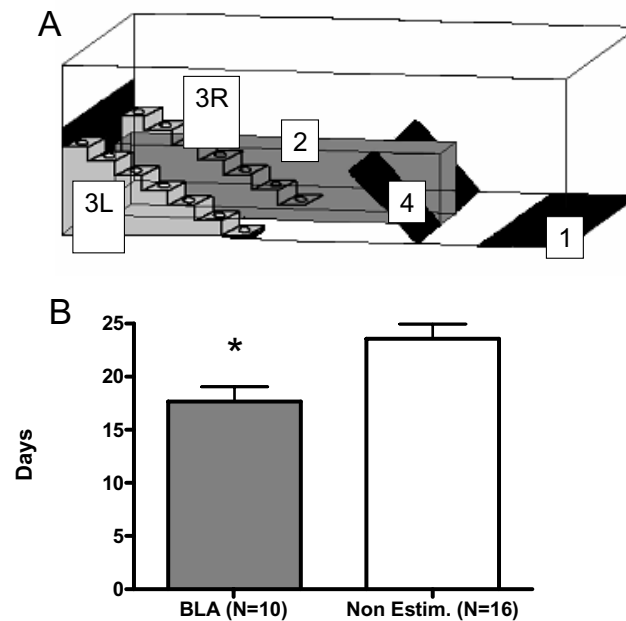


Fig. 1. A) Drawing of the staircase device used to train the animals. 1: opening to enter the rat; 2: central platform; 3: lateral staircases (Left and Right); 4: blocker of foot motion. Only the right staircase was baited. B) Results of training to develop a motor skill for the different groups (mean $\pm$ SEM). * $P < 0.05$, Student's *t* test.

2.5. Mapping the motor cortex

After reaching the learning criteria, the motor cortex of the animals was mapped and compared to a group of non-trained animals. The procedure was performed under ketamine narcosis (7 mg/kg) supplemented with diazepam and atropine. A window was opened on the left hemisphere (AP: 3.5 to -1 mm, ML: 0.0 to 5.0 mm, all coordinates from bregma) to expose the motor cortex. The dura was carefully removed and a 0.5 M Ω stainless steel monopolar microelectrode (World Precision Instruments, Sarasota, USA) was lowered to -1.5 mm from the cortical surface at different points with 0.5 mm interval in each direction (AP or ML) starting at AP: 3.5 mm, ML: 1.5 mm and finishing at AP: -0.5 mm, ML: 5.0 mm. In cases in which a motor response was obtained in any of the border points the stimulation area was increased in 0.5 mm in the required direction until a no-response border surrounded the whole area. The stimulus applied at each point consisted of 2 trains of 9 impulses (0.2 ms pulse duration, 3 ms interpulse interval and 400 μ A intensity) separated by a 1.2 sec intertrain interval. The response in any of the following contralateral body parts was determined by direct observation of one experienced observer, and annotated in the protocol using the following code numbers: 0- no response, 1- vibrissae, 2- anterior limb, 3- neck, 4-

shoulder, 5- trunk, and 6- posterior limb. The total area of the cortex controlling the movement of each body part was then calculated by multiplying the number of response points for each part by 0.25 mm². In case that the stimulation of one point was followed by a motor response in two body regions, an area of 0.125 mm² was added to each.

2.6. Histology

After finishing the motor mapping, and still under narcosis the animals were transcardially perfused with formaline, the brains removed and prepared for histological observation to ascertain the location of the implanted electrode. Only animals showing a correct location of the electrode at the BLA were considered for further analysis. One animal intended for the BLA-stimulated group, and two for the sham-stimulated group were rejected by this reason.

2.7. Statistics

One way and two ways ANOVAs were performed to ascertain significant differences among groups. A Student's *t*-test was performed, post hoc to compare between groups. In every case differences were considered significant only if $P < 0.05$.

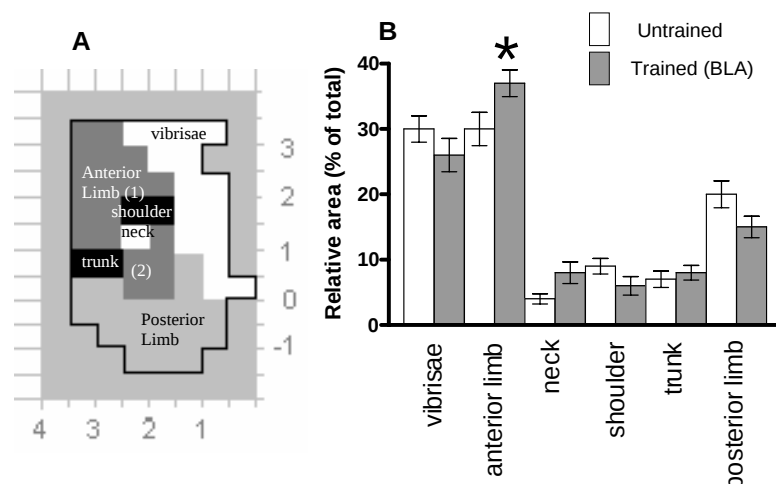


Fig. 2. A) A representative example of the motor map on the left cortical surface from non-trained animals. The areas representing the different body parts are named and identified using different tones. The values in the scale are in mm, the value 0 representing the position of the reference bregma. B) Changes in the relative area of the different body parts as a result of training. Shaded columns: trained animals (mean $\pm$ SEM). Non-shaded columns: non-trained animals (mean $\pm$ SEM). * Significant difference due to training (ANOVA 2-way).

3. Results

Analyzing the results of the motor training, the univariate ANOVA showed that the stimulation of the BLA significantly improved the acquisition of the motor task ($F_{2,23} = 6.377$, $p < 0.05$). The t-test showed no significant differences between the groups of animals in which the BLA was not stimulated, the trained and sham groups ($t = 1.630113$, $p = 0.125362$). When these two groups were pooled together and compared to the BLA stimulated animals, the t-test showed significant differences ($t = 2.918267$, $p = 0.007530$, see Fig. 1B).

The results of motor cortex mapping are shown on Fig. 2. At the left side (Fig. 2A) a representative motor map, obtained from naïve, non-trained animals, is shown. The total area of the motor cortex was about 10 mm². As can be seen, the area for the vibrissae, as well as the anterior and posterior limbs, occupies the greater portions of the motor cortex. The right part of the Fig. 2(B) shows the relative distribution of the mapped body regions in non-trained animals compared to that of trained animals receiving BLA stimulation. All the groups of trained animals showed similar changes, irrespective of post-training handling or stimulation. As can be noticed, the area representing the anterior limb was increased after training. The 2-way ANOVA showed that this increase is statistically significant ($F_{1,41} = 5.18$, $p < 0.05$) but caused only by training. The stimulation of the BLA did not influence

the final modification in the motor cortical map, except for the fact that it occurred faster.

Our results confirm that cortical maps can be modified by experience, as an expression of cortical plasticity, and show that the mechanisms involved in such changes can be modulated by the stimulation of limbic sub cortical structures like the BLA.

4. Discussion

The primary motor cortex is the main cortical output to the lower motor neurons in the spinal cord or the brain stem and it plays a pivotal role in the performance of skilled movements [30]. Within motor cortex there is a topographic representation of the different body parts, known as motor homunculus in humans, or motor map in the rat. Although such maps are hardwired, they maintain the ability to change [7,20] in response to different challenges like lesions [14] or training [32].

Previous reports have shown that the area representing the anterior limb within the rat primary motor cortex expands after learning specific skilled movement, but not after non-specific general exercise [21] or force training [32]. These results are in line with those reported in the present article. The training method applied to the animals lead to an expansion of the cortical area initiating movements of the anterior limb. We may, therefore, assume that the measured expansion is functionally related to the acquired motor ability.

The mechanisms involved in such changes might be several and may be different in the different stages of acquisition and consolidation of the learned skill [8]. Synaptic plasticity, in form of a LTP-like enhancement of connectivity between cortical neurons, has been demonstrated within the motor cortex [34]. This type of cortical LTP shows mechanistic similarities with hippocampal LTP, as its dependence on NMDA receptors activation [12], and has been related to the initial phases of skilled movement acquisition [28]. Interestingly, similar changes have been demonstrated in humans [39]. Continued motor training may lead to longer-lasting plastic changes within the motor cortex, like an increased synaptogenesis [22] during late phases of acquisition, which may finally be expressed in form of a modified motor map. All those mechanisms depend on protein synthesis [20] as it has been shown for neural plasticity in other brain regions [4].

Neural plasticity processes in the cortex may be influenced by the activity of modulatory subcortical systems. A relevant influence of basal forebrain cholinergic inputs on cortical plasticity, in motor [9], sensory [31], and association areas [38] have been experimentally documented. Our results show that the stimulation of the BLA can also exert a modulatory influence on motor cortical plasticity, facilitating the acquisition of a motor skill that correlates with changes in the motor cortex map.

We have previously shown that the stimulation of the BLA prolongs LTP in the dentate gyrus of the hippocampus [11]. This reinforcing effect is protein synthesis dependent and is likely mediated by cholinergic and noradrenergic inputs [11]. The modulating afferents seemed to reach the hippocampal formation via the basal forebrain-fornical projection [18] likely activated by the BLA stimulation [19].

A similar mechanism might mediate the facilitating effect of BLA stimulation on the acquisition of a motor skill. The motor cortex receives cholinergic and noradrenergic projections [23,27] from the basal forebrain [13] and the locus coeruleus [6,37]. These modulating systems can activate neuronal metabolic cascades [15,17] leading to an enhanced and/or modified synthesis of the proteins required to consolidate synaptic plasticity and to promote synaptogenesis.

Alternative pathways might include the prefrontal cortex which have been recently shown to influence the acquisition of motor skills [10], or the insular cortex [36].

As mentioned in the Methods section, we have stimulated the BLA contralateral to the hemisphere involved

in the motor skill acquisition to avoid damaging a near cortical region that might have caused plastic changes not related to training. The BLA projection to cortical regions is bilateral [1], but even if the contralateral projection is weaker, our results showing that stimulating the BLA improves the acquisition of a motor skill gain in value.

The amygdala is part of a complex brain system responsible for the evaluation of affective (emotion, motivation) status of the organism [24,35]. Accumulating evidence indicate that affective factors can influence neural plasticity, not restricted to memory, but also the cellular and subcellular mechanisms involved in plasticity [1,3,16,33,35]. Our results confirm and expand that observation showing that the activation of the BLA can influence also cortical plasticity.

Our results may be generalized to suggest that the modulatory influence of the amygdala on neural plasticity might represent a universal property of the brain, not just restricted to particular structures or processes. Understanding the interactions between affective factors and neural plasticity appears, therefore, to be not only of great theoretical interest, but potentially relevant to practical use in different fields, from educational fields to restorative neurology as well.

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References

- [1] I. Akirav and G. Richter-Levin, Mechanisms of amygdala modulation of hippocampal plasticity, *J. Neurosci.* (2002), 9912–9921.
- [2] W. Almaguer-Melian, V. Capdevila, M. Ramirez, A. Vallejo, J.C. Rosillo-Martí, and J. Bergado-Rosado, Post-training stimulation of the basolateral amygdala improves spatial learning in rats with lesion of the fimbria-fornix, *Restor. Neurol. Neurosci.* **23** (2005), 43–50.
- [3] W. Almaguer-Melian, L. Martínez-Martí, J.U. Frey and J.A. Bergado, The amygdala is part of the behavioral reinforcement system modulating long-term potentiation in rat hippocampus, *Neuroscience* **119** (2003), 319–322.

- [4] J.A. Bergado Rosado and W. Almaguer Melian, Mecanismos celulares de la neuroplasticidad, *Rev. Neurol.* **31** (2000), 1074–1095.
- [5] J.A. Bergado, W. Almaguer-Melian, S. Kostenko, S. Frey and J.U. Frey, Behavioral reinforcement of long-term potentiation in rat dentate gyrus *in vivo* is protein synthesis-dependent, *Neurosci Lett.* **351** (2003), 56–58.
- [6] C.W. Berridge and E.D. Abercrombie, Relationship between locus coeruleus discharge rates and rates of norepinephrine release within neocortex as assessed by *in vivo* microdialysis, *Neuroscience* **93** (1999), 1263–1270.
- [7] J. Biernaskie, A. Szymanska, V. Windle and D. Corbett, Bi-hemispheric contribution to functional motor recovery of the affected forelimb following focal ischemic brain injury in rats, *Eur. J. Neurosci.* **21** (2005), 989–999.
- [8] M.M. Buitrago, T. Ringer, J.B. Schulz, J. Dichgans and A.R. Luft, Characterization of motor skill and instrumental learning time scales in a skilled reaching task in rat, *Behav. Brain Res.* **155** (2004), 249–256.
- [9] J.M. Conner, A. Culbertson, C. Packowski, A. Chiba and M.H. Tuszynsky, Lesions of the basal forebrain cholinergic system impair task acquisition and abolish cortical plasticity associated with motor skill learning, *Neuron* **38** (2003), 819–829.
- [10] J.W. Dalley, R.N. Cardinal and T.W. Robbins, Prefrontal executive and cognitive functions in rodents: neural and neurochemical substrates, *Neurosci. Biobehav. Rev.* **28** (2004), 771–784.
- [11] S. Frey, J.A. Bergado, T. Seidenbecher, H.C. Pape and J.U. Frey, Reinforcement of early-LTP in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction of late-LTP, *J. Neurosci.* **21** (2001), 3697–3703.
- [12] D.J. Froc and R.J. Racine, N-methyl-D-aspartate receptor-independent long-term depression and depotentiation in the sensorimotor cortex of the freely moving rat, *Neuroscience* **129** (2004), 273–281.
- [13] R.P. Gaykema, G. Gaal, J. Traber, L.B. Hersh and P.G. Luiten, The basal forebrain cholinergic system: efferent and afferent connectivity and long-term effects of lesions, *Acta Psychiatr. Scand. Suppl* **366**(14–26) (1991), 14–26.
- [14] O.A. Gharbawie, C.L. Gonzalez, P.T. Williams, J.A. Kleim and I.Q. Whishaw, Middle cerebral artery (MCA) stroke produces dysfunction in adjacent motor cortex as detected by intracortical microstimulation in rats, *Neuroscience* **130** (2005), 601–610.
- [15] M. Grzegorzewska, M. Przybylo, A. Litynska and G. Hess, Chemically-induced long-term potentiation in rat motor cortex involves activation of extracellular signal-regulated kinase cascade, *Brain Res.* **1021** (2004), 192–199.
- [16] C. Helmeke, G. Poeggel and K. Braun, Differential emotional experience induces elevated spine densities on basal dendrites of pyramidal neurons in the anterior cingulate cortex of Octodon degus, *Neuroscience* **104** (2001), 927–931.
- [17] G. Hess, Synaptic plasticity of local connections in rat motor cortex, *Acta Neurobiol. Exp. (Wars.)* **64** (2004), 271–276.
- [18] J. Jas, W. Almaguer, J.U. Frey and J. Bergado, Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus., *Brain Res.* **861** (2000), 186–189.
- [19] E. Jolkkonen, R. Miettinen, M. Pikkarainen and A. Pitkanen, Projections from the amygdaloid complex to the magnocellular cholinergic basal forebrain in rat, *Neuroscience* **111** (2002), 133–149.
- [20] J.A. Kleim, R. Bruneau, K. Calder, D. Pocock, P.M. VandenBerg, E. MacDonald, M.H. Monfils, R.J. Sutherland and K. Nader, Functional organization of adult motor cortex is dependent upon continued protein synthesis, *Neuron* **40** (2003), 167–176.
- [21] J.A. Kleim, N.R. Cooper and P.m. VandenBerg, Exercise induces angiogenesis but does not alter movement representations within rat motor cortex, *Brain Res.* **934** (2002), 1–6.
- [22] J.A. Kleim, T.M. Hogg, P.m. VandenBerg, N.R. Cooper, R. Bruneau and M. Remple, Cortical synaptogenesis and motor map reorganization occur during late, but not early, phase of motor skill learning, *J. Neurosci.* **24** (2004), 628–633.
- [23] M. Latsari, I. Dori, J. Antonopoulos, M. Chiotelli and A. Dinopoulos, Noradrenergic innervation of the developing and mature visual and motor cortex of the rat brain: a light and electron microscopic immunocytochemical analysis, *J. Comp Neurol* **445** (2002), 145–158.
- [24] J.E. LeDoux, Emotional memory systems in the brain, *Behav. Brain Res.* **58** (1993), 69–79.
- [25] J.L. McGaugh, Memory consolidation and the amygdala: a systems perspective, *Trends Neurosci.* **25** (2002), 456–461.
- [26] J.L. McGaugh, C.K. McIntyre and A.E. Power, Amygdala modulation of memory consolidation: interaction with other brain systems, *Neurobiol. Learn. Mem.* **78** (2002), 539–552.
- [27] N. Mechawar, C. Cozzari and L. Descarries, Cholinergic innervation in adult rat cerebral cortex: a quantitative immunocytochemical description, *J. Comp Neurol* **428** (2000), 305–318.
- [28] M.H. Monfils and G.C. Teskey, Skilled-learning-induced potentiation in rat sensorimotor cortex: a transient form of behavioural long-term potentiation, *Neuroscience* **125** (2004), 329–336.
- [29] C.P. Montoya, L.J. Campbell-Hope and S.B. Dunnet, The “Staircase test”: A measure of independent forelimb reaching and grasping abilities in rats, *J. Neurosci. Meth.* **36** (1991), 219–228.
- [30] D.M. Piecharka, J.A. Kleim and I.Q. Whishaw, Limits on recovery in the corticospinal tract of the rat: Partial lesions impair skilled reaching and the topographic representation of the forelimb in motor cortex, *Brain Res. Bull.* **66** (2005), 203–211.
- [31] N. Prakash, S. Cohen-Cory, S. Penschuck and R. Frostig, The basal forebrain cholinergic system is involved in rapid nerve growth factor (NGF)-induced plasticity in the barrel cortex of adult rats, *J Neurophysiol.* (2003).
- [32] M.S. Remple, R.M. Bruneau, P.m. VandenBerg, C. Goertzen and J.A. Kleim, Sensitivity of cortical movement representations to motor experience: evidence that skill learning but not strength training induces cortical reorganization, *Behav. Brain Res.* **123** (2001), 133–141.
- [33] G. Richter-Levin, The amygdala, the hippocampus, and emotional modulation of memory, *Neuroscientist* **10** (2004), 31–39.
- [34] M.S. Rioult-Pedotti, D. Friedman, G. Hess and J.P. Donoghue, Strengthening of horizontal cortical connections following skill learning, *Nat Neurosci.* **1** (1998), 230–234.
- [35] M.T. Rogan and J.E. LeDoux, Emotion: Systems, cells, synaptic plasticity, *Cell* **85** (1996), 469–475.
- [36] C.J. Shi and M.D. Cassell, Cortical, thalamic, and amygdaloid connections of the anterior and posterior insular cortices, *J. Comp Neurol* **399** (1998), 440–468.
- [37] C.H. Vanderwolf and G.B. Baker, The role of brain noradrenaline in cortical activation and behavior: A study of lesions of the locus coeruleus, medial thalamus and hippocampus-neocortex and of muscarinic blockade in the rat, *Behav. Brain Res.* **78** (1996), 225–234.

- [38] E.C. Warburton, T. Koder, K. Cho, P.V. Massey, G. Duguid, G.R. Barker, J.P. Aggleton, Z.I. Bashir and M.W. Brown, Cholinergic neurotransmission is essential for perirhinal cortical plasticity and recognition memory, *Neuron* **38** (2003), 987–996.
- [39] U. Ziemann, T.V. Iliac, C. Pauli, F. Meintzschel and D. Ruge, Learning modifies subsequent induction of long-term potentiation-like and long-term depression-like plasticity in human motor cortex, *J Neurosci* **24** (2004), 1666–1672.

DISCUSIÓN GENERAL

Como elemento central de esta discusión reproducimos a continuación el texto del último trabajo realizado en esta serie.

Este experimento estuvo diseñado con el propósito de precisar el papel de los sistemas de neurotransmisión noradrenérgico y colinérgico, teniendo en cuenta las estructuras que les dan origen.

Para evitar la falta de especificidad que provoca la inyección intraventricular, se empleó la aplicación tópica de las sustancias en cada una de las estructuras seleccionadas.

Los resultados mostraron que el locus coeruleus es un mediador de los efectos reforzadores de la estimulación de la amígdala. La activación del locus se realiza mediante una proyección colinérgica. Así mismo, la proyección noradrenérgica del locus a la región septal y al giro dentado son parte imprescindible del sistema neural de reforzamiento. De esta estructura nace una proyección colinérgica que alcanza al giro dentado del hipocampo mediante la fimbria-fornix y que resulta necesaria para que el efecto reforzador se manifieste. También por la vía fimbria-fornix llegan al giro dentado fibras noradrenérgicas directas.

Basados en esos resultados se propone un mecanismo neural que se ilustra en la figura 7 del trabajo. En él se propone que el reforzamiento de la LTP en el giro dentado por factores afectivos requiere la activación secuencial y ordenada de la amígdala, la cuál por medio de una proyección colinérgica activa al locus coeruleus. La proyección noradrenérgica que nace de esta estructura tiene un efecto estabilizador sobre fases intermedias de la LTP inducida en el giro dentado y, por otra parte, activa la proyección

colinérgica del septo medial al giro dentado que se necesita para reforzar las fases tardías del proceso neuroplástico.



Cholinergic afferents to the locus coeruleus and noradrenergic afferents to the medial septum mediate LTP-reinforcement in the dentate gyrus by stimulation of the amygdala

Jorge A. Bergado ^a, Sabine Frey ^b, Jeffrey López ^c, William Almaguer-Melian ^a,
Julietta U. Frey ^{b,*}

^a International Center for Neurological Restoration, Havana, Cuba

^b Department of Neurophysiology, Leibniz Institute for Neurobiology, Brenneckestrasse 6, 39118 Magdeburg, Germany

^c National Institute for Neurology and Neurosurgery, Havana, Cuba

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Abstract

Transient long-term potentiation (E-LTP) can be transformed into a long-lasting LTP (L-LTP) in the dentate gyrus (DG) by behavioral stimuli with high motivational content. Previous research from our group has identified several brain structures, such as the basolateral amygdala (BLA), the locus coeruleus (LC), the medial septum (MS) and transmitters as noradrenaline (NA) and acetylcholine (ACh) that are involved in these processes. Here we have investigated the functional interplay among brain structures and systems which result in the conversion of a E-LTP into a L-LTP (reinforcement) by stimulation of the BLA (BLA-R). We used topical application of specific drugs into DG, and other targets, while following the time course of LTP induced by stimulation of the perforant pathway (PP) to study their specific contribution to BLA-R. One injection cannula, a recording electrode in the DG and stimulating electrodes in the PP and the BLA were stereotactically implanted one week before electrophysiological experiments. Topical application of atropine or propranolol into the DG blocked BLA-R in both cases, but the effect of propranolol occurred earlier, suggesting a role of NA within the DG during an intermediate stage of LTP maintenance. The injection of lidocaine into the LC abolished BLA-R indicating that the LC is part of the functional neural reinforcing system. The effect on the LC is mediated by cholinergic afferents because application of atropine into the LC produced the same effect. Injection of lidocaine inactivating the MS also abolished BLA-R. This effect was mediated by noradrenergic afferents (probably from the LC) because the application of propranolol into the MS prevented BLA-R. These findings suggest a functional loop for BLA-R involving cholinergic afferents to the LC, a noradrenergic projection from the LC to the DG and the MS, and finally, the cholinergic projection from the MS to the DG.

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Keywords: Early-LTP; Late-LTP; Hippocampus; Reinforcement; Memory formation; Learning; Memory system

1. Introduction

Long-term potentiation (LTP) is a form of neural plasticity that represents a common cellular mechanism for many forms of learning (Matthies, 1989), and is gaining

recognition as a putative mechanism for functional recovery after brain insult (Carmichael, 2003).

The importance of affective behavioral factors (motivation, emotions) for the consolidation of memory is well established on empirical basis, and supported by extensive animal research. Affective modulation of memory is mediated by limbic structures such as the amygdala (McIntyre, Power, Roozendaal, & McGaugh, 2003; Richter-Levin, 2004). Similar modulatory influences affect the acquisition of motor skills (Bergado, Rojas, Capdevila,

* Corresponding author. Fax: +49 391 6263 421.

E-mail address: frey@ifn-magdeburg.de (J.U. Frey).

Gonzalez, & Almaguer-Melian, 2006) and the recovery of mental functions in stroke patients (Dam, 2001; Narushima, Chan, Kosier, & Robinson, 2003) as well as memory formation in animal models created by the use of brain lesion (Almaguer-Melian et al., 2005). Thus, affective modulation can be described as an universal mechanism, not restricted to a particular form of plasticity process or brain locus.

LTP can also be modulated by affective factors. One example of this is the transformation of an early-LTP (E-LTP) into a late-LTP (L-LTP) lasting more than 4 hours, by the action of a behavioral act with a high motivational value (Seidenbecher, Reymann, & Balschun, 1997; Straube, Korz, & Frey, 2003; Uzakov, Frey, & Korz, 2005). This 'behavioral reinforcement' (BR) of LTP is mediated by protein synthesis (Bergado, Almaguer-Melian, Kostenko, Frey, & Frey, 2003) and depends on BLA function as demonstrated in experiments using animals with temporal or permanent inactivation of this limbic structure (Almaguer-Melian, Martínez-Martí, Frey, & Bergado, 2003). In addition, stimulating the BLA within a specific effective time window, before or after the induction of LTP using a weak tetanus (BLA-R, see Methods) mimics the effects of behavioral reinforcement (Frey, Bergado-Rosado, Seidenbecher, Pape, & Frey, 2001). In other words, a reinforcing effect can also be obtained by stimulation of the BLA (BLA-R). The reinforcing effect of the BLA on LTP in the dentate gyrus (DG) seems to involve noradrenergic and cholinergic afferent innervation (Frey et al., 2001). This suggests the participation of the locus coeruleus (LC) and the medial septum (MS) in these processes as the main sources of noradrenergic and cholinergic innervation to the DG (Vizi & Kiss, 1998). Indeed, we recently found that LTP in the DG can be reinforced by stimulation of the MS (Frey, Bergado, & Frey, 2003) and the LC (Frey, unpublished results).

However, in prior pharmacological studies the antagonists were applied intracerebroventricularly (icv), thus the question arises of where in the brain the mentioned pharmacological effects actually take place. Here we use an approach of topical antagonist application into the structures of interest: the DG, the BLA, the LC, and the MS. The results suggest the existence of a functional loop for BLA-R involving cholinergic afferents to the LC, a noradrenergic projection from the LC to the DG and the MS, and finally, the cholinergic projection from the MS to the DG.

2. Materials and methods

2.1. Animals

Male Wistar rats weighing 250–300 g (8 weeks old) at the beginning of the study were used. After surgery the animals were housed individually in plastic translucent boxes with free access to food and water and controlled room conditions (12 h light/dark cycle, temperature and humidity) during the entire experiments. For all experiments with animals ethical approval was sought prior to the studies according to the German requirements for the use of experimental animals.

2.2. Surgery and implants

Rats were anaesthetized using nembutal (40 mg/kg, i.p.) and mounted in a stereotactic frame (David Kopf, Saint Louis, USA). The standard preparation included the implantation of one stainless steel monopolar recording electrode in the DG (0.125 mm diameter, coordinates: antero-posterior AP = −4.0 mm, mediolateral ML = 2.3 mm from bregma and dorsoventral DV ≈ 2.5 mm from the dural surface, with lambda and bregma at the same high). A bipolar stainless steel electrode was placed in the medial perforant path (PP) (AP = −7.5 mm, ML = 4.1 mm from bregma, DV = −3.0 mm from the dural surface) and a monopolar electrode into the BLA (AP = −3.5 mm, ML = 5.0 mm from bregma, DV = −7.6 mm from the dural surface). The position in the DV axis of the electrodes in the DG and the PP was adjusted under observation of the evoked potentials to optimize its location.

Guide cannuli for the topical injection of substances were also placed in one of the following locations: LC (AP = −10.3 mm, ML = 1.1 mm, DV = −5.7 mm, bregma 3 mm lower than lambda); MS (AP = 0.7 mm, ML = 1.8 mm, DV = −6.6 mm, bregma and lambda at the same high, tilted 15° in lateral direction); the BLA or the DG (same coordinates as given for the electrodes). DV coordinates were selected so that the guide cannula was positioned with its tip approximately 1 mm above the target. In the animals with cannuli implanted into the BLA or the DG the electrode was attached to it, protruding 0.5 mm the bevel edge to form a so called chemitrode. The guide cannuli were made of 0.5 mm (external diameter) stainless steel tubes cut to the appropriate length according to the target. A 0.3 mm stainless steel "stiletto" was placed into the cannula to prevent the outflow of cerebrospinal fluid.

All implants were placed at the right hemisphere. Miniscrews were attached at the skull in the right frontal bone, left parietal bone and the left parieto-occipital junction, to serve as ground, indifferent electrode and mechanical anchorage. A silver wire with the tip thermally rounded was placed over the right parietal dura to serve as returning electrode for stimulating the BLA. Electrodes were then connected to flexible rubber female sockets and fixed to the skull with dental cement.

2.3. Electrophysiology

2.3.1. Habituation and I/O curves

A week after surgery the animals were tested and habituated to the recording chamber for no less than 4 h. An input–output was obtained stimulating the PP (100–800 μ A) to determine the stimulus intensity to be used in each animal (40% maximal population spike). There were no differences in the average stimulus intensity and baseline values between the experimental groups.

2.3.2. General design

The experiments were performed on the following day. The general experimental design included the recording of a baseline for 1 h (12 recordings with 5 min interval) followed by the induction of an early-LTP by a weak tetanus (see below) to the PP. Ten minutes after tetanus substances were applied topically into the LC, the MS, the BLA or the DG. The BLA was stimulated 15 min after LTP induction (i.e. 5 min after administration of substances) with an identical pattern of weak tetanus. The time course of LTP was followed up for 8 h with recordings every 15 min. A further recording was made 24 h after tetanization (see Fig. 1e).

2.3.3. Recording of evoked potentials at the dentate gyrus

Each record consisted of 5 averaged waveforms evoked by square pulses (0.1 ms) at 0.1 Hz (Isolated Pulse Stimulator, A-M System, USA) to the PP. Signals were filtered between 1 and 5000 Hz using an AB 621 G Bioelectric Amplifier (Nihon Kohden, Japan) and fed to a 1401 Plus A/D converter (CED, Cambridge, UK), and processed by a special software (Intracell, Institute for Neurobiology, Magdeburg, Germany). The amplitude of the population spike (PSA) was measured in mV (from the most positive point reached by the first deflection to the subsequent most negative point, see Fig. 1e—measurement between the two markers

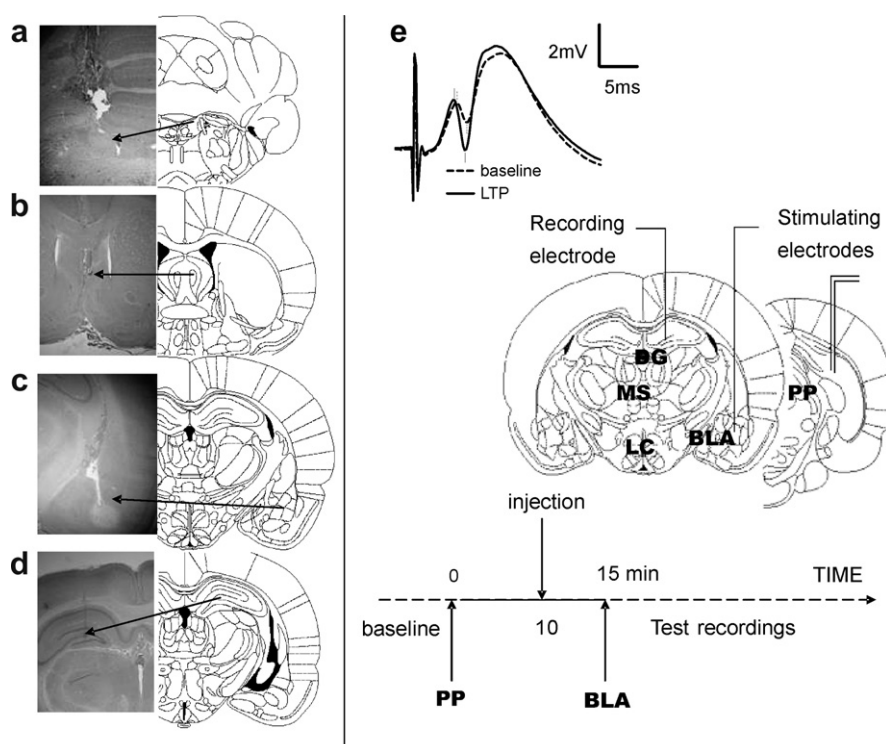


Fig. 1. (a–d) Schematic summary of the results of the histological study, showing the location of the cannuli at the LC (a) the MS (b) the BLA (c) and the DG (d). A representative microphotograph and a schematic representation of the electrode location, and the approximate cannula locations are shown. (e) The inset above, shows representative evoked potential before (broken line) and after the induction of LTP (filled line) recorded at the DG. The population spike amplitude (PSA) was measured between the markers shown. Below, a summarized view of the experimental design is presented as an example with recording electrodes positioned in the dentate gyrus and stimulating electrodes in the PP as well as in the BLA.

per analog trace). The PSA was averaged in the last 6 baseline recordings to determine pre-tetanus baseline value. The PSA of the post-tetanus recordings for each animal were compared to these values and expressed in percent changes. We used the PSA as measure of LTP instead of the EPSP for several reasons. Working in vivo, in freely moving animals, and using a single recording electrode it is difficult to assess simultaneously the EPSP and the PSA. On the other hand, the PSA represents the discharge of action potentials by the postsynaptic population, which is functionally the more relevant outcome of synaptic function.

2.3.4. LTP induction

A weak tetanus protocol was used to induce LTP at the PP-DG synapses and to stimulate the amygdala. It consisted of 3 trains of 15 impulses each at 200 Hz (square biphasic pulses 0.2 ms). Intertrain interval was 10 s. The same stimulus pattern was used for stimulation of the BLA using a constant current of 400 μ A.

2.3.5. Drugs and application

The substances used in this experiment were: lidocaine (Lidocainhydrochlorid 2%, Hexal, Holzkirchen, BRD) a local anesthetic, to produce the temporary inactivation of the target region. Propranolol ((S)-(–)-propranolol hydrochloride, Sigma–Aldrich, Saint Louis, USA) to block locally the β -adrenergic receptors, and atropine (atropine sulfate, salt, Sigma–Aldrich, Saint Louis, USA) to block locally the muscarinic acetylcholine receptors, all dissolved in saline solution. The injection cannuli were made of stainless steel tubes (0.3 mm external diameter) cut to an appropriate length to protrude 1 mm from the bottom of the guide cannula. The injector was carefully introduced in to the implanted guide cannula, and 1 μ l of each substance or vehicle was injected using a Hamilton microsyringe (CR-700-20, Hamilton Co., Reno, USA) in 2 min (≈ 0.05 μ l

every 5 s). The cannula was left in place for 1 min before slowly retracting it. The volumes injected contained 6.76 nmol propranolol or 1 nmol atropine, respectively.

In a small group of animals ($n = 3$) we have tested the injection procedure in the dentate gyrus. The injection of 1 μ l saline in the dentate gyrus does not change the form or amplitude of the evoked potentials recorded at the tip of the nearby located recording electrode (data not shown). The injection of 1 μ l lidocaine provoked a transient reduction of the evoked potentials of nearly 30% the PSA, which returned to normal values in about 20 min. Apparently, no mechanic disruption was caused by the injection despite the vicinity between the cannula and the electrode, however they were close enough to allow a direct influence of the injected substance on the recorded cell population.

The effectiveness of the used drug concentrations were documented in previous papers (Almaguer-Melian et al., 2003, 2005; Frey et al., 2001, 2003).

2.3.6. Experimental groups

Several experimental groups were randomly formed, according to the stimulation protocol, the substance applied and the place of injection.

Injections into the DG, 10 min after LTP induction (5 min before BLA stimulation) were done using atropine ($n = 9$), propranolol ($n = 10$), or saline ($n = 10$). To assess a possible direct effect of both substances on LTP or the evoked potentials, the following controls were also carried out: Injection of atropine ($n = 7$), propranolol ($n = 8$), or saline ($n = 10$) after LTP induction without stimulating the BLA; as well as injection of atropine ($n = 8$), propranolol ($n = 8$), or saline ($n = 8$) without LTP induction or BLA stimulation.

Similarly, 10 min after LTP induction one of these substances was applied into the LC: lidocaine ($n = 8$), atropine ($n = 8$), or propranolol ($n = 8$). For injection into the MS: lidocaine ($n = 7$), atropine ($n = 7$), or

propranolol ($n = 8$). For the injection into the BLA: atropine ($n = 9$), or propranolol ($n = 9$). A group of 9 animals injected in these structures with saline solution served as common control group, as there were no differences in their initial level and time course of potentiation (LC $n = 4$, MS $n = 3$ and BLA $n = 2$, Fig. 2a—inset). In all groups (with the exception of those assessing substance effects on baseline or simple LTP) the BLA was stimulated 5 min after injection, i.e. 15 min after LTP induction.

2.3.7. Behavioral control and time of the experiments

All electrophysiological (recordings or stimulation of the PP or the BLA) and injection procedures were performed in awake and freely moving animals. The behavior of the animals during, and in the period after each procedure was carefully observed by an experienced behavior analyst. No signs of behavioral modification were observed in any of these conditions. To minimize and standardize circadian influences all the experiments were carried out at approximately the same time of the day. Tetanization of the PP to induce LTP (time 0) was carried out always between 10 and 10:30 a.m.

2.3.8. Statistics

The percent values of the PSA were compared between treatments (groups) using a two-way ANOVA with TREATMENT and TIME (repeated measures) as factors in independent analysis for each injection site. Significant differences between groups at specific time points were ascertain using the post hoc test of Tukey (honest significant difference,

HSD) when the ANOVA showed significant differences between groups (TREATMENT) or the interaction. In every case a $p < .05$ was considered significant.

2.3.9. Histology

At the end of the experiment the animals were transcardially perfused with formaline under nembutal narcosis. The brain was extracted and post-fixed. Frozen brains were cut into 40 μm thick slices containing the regions of interest and stained with toluidin blue. The location of the cannula was estimated by direct inspection of the local damage in the surrounding tissue. Animals showing a misplaced cannula or electrode were rejected and not considered in the final group analysis.

3. Results

Histological analyses allowed us to select only those animals showing a correct placement of the cannuli. Fig. 1a–d shows a constructed summary for the cannula location in each of the targets. The LC is a small, deep region of the brain stem. For this reason the number of rejections for this structure was higher than for any other structure, where rejections were required only exceptionally.

The direct administration of the antagonists into the DG showed significant effects on the reinforcing effects of

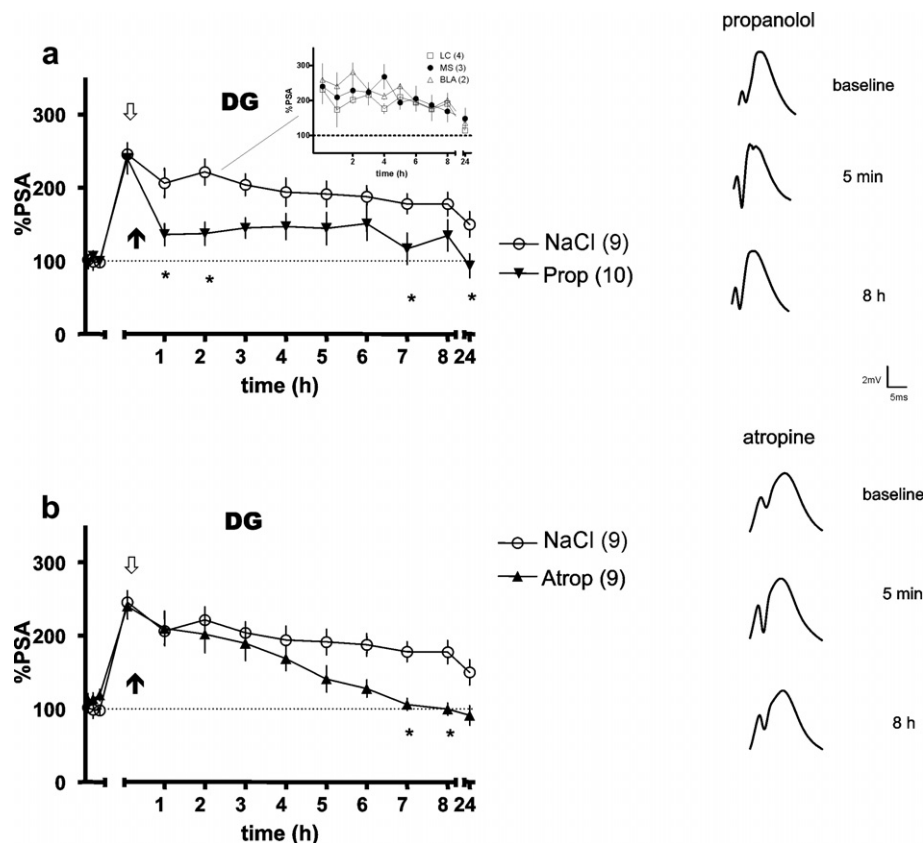


Fig. 2. Effects of topical application of drugs into the DG on LTP reinforcement by stimulation of the BLA—(a) after propranolol injection into the BLA (inset represents the time course of the averaged control group provided in the graphs which consists of 9 animals injected in structures of interest with saline solution ($n = 4$ LC, $n = 3$ MS and $n = 2$ BLA), and (b) after atropine. Mean values ($\pm$ SEM) of the PSA (% PSA, relative to baseline) for the experimental groups are shown (the numbers in parentheses indicates the number of animals in each groups). LTP was induced in the DG by stimulation of the PP at $t = 0$, and reinforced by stimulation of the BLA 15 min later (black arrow). Substances were topically applied into the DG between both stimuli ($t = 10$ min, open arrow). Asterisks indicate significant differences between groups in the post hoc test (Tukey HSD, $p < .05$). Representative traces from one animal in selected groups, before (baseline) and after E-LTP induction (5 min and 8 h) are shown in the right part of the figure.

BLA stimulation (Fig. 2). Two-way ANOVA showed TREATMENT as a significant factor ($F_{2,27} = 3.79097$). Atropine produced a delayed decay of LTP, which starts about 4–5 h and reached significance 7–8 h after LTP induction (Fig. 2b). Propranolol also showed a negative influence on BLA-R (Fig. 2a), but the effect was much earlier and apparently biphasic. The level of potentiation after BLA-R in the propranolol group was significantly reduced 1–2 h after induction, and at 7–24 h.

When these drugs were applied to non-tetanized animals, atropine showed no significant effect. Propranolol, however induced a mild, but significant depression of the PP-DG evoked potentials (Fig. 3a). When propranolol or atropine were applied to animals in which E-LTP was induced without BLA stimulation, none of them showed significant effects on LTP time course (Fig. 3b).

The administration of the same drugs at the BLA itself produced more moderate effects. No significant treatment effect was statistically confirmed ($F_{2,24} = 1.79602$). However the interaction time $\times$ treatment was significant ($F_{18,216} = 2.00483$). The post hoc Tukey test detected significant differences only 4 h after atropine application (see Fig. 4).

Fig. 5 shows the results of pharmacological manipulation in the LC after LTP induction and before BLA stimulation. The two-way ANOVA showed TREATMENT as a significant factor ($F_{3,30} = 7.01437$). Results of the post

hoc comparison of the drug/time interaction are shown in detail in graphs a, b, and c, all compared to the control group treated with saline (NaCl). The injection of lidocaine (Fig. 5a) abolished the reinforcing effect on DG-LTP of stimulating the amygdala. Significant differences between groups were seen starting at 2 h. The administration of atropine has a similar blocking effect as lidocaine (Fig. 5b), while propranolol produced no significant effect (Fig. 5c).

When substances were applied to the MS (Fig. 6) the factor TREATMENT also was significant (two-way ANOVA, $F_{3,28} = 15.17692$). Inactivation of this region by topical application of lidocaine caused a blockade of the reinforcing effect of BLA stimulation on DG-LTP (Fig. 5a) showing a decay of the potentiation that significantly differs from the saline controls (NaCl). The topical administration of propranolol in this structure caused a very fast and sustained decay of LTP (Fig. 5c). Atropine, on the other hand, produced no significant effect (Fig. 5b).

4. Discussion

Our results indicate that both, topically applied propranolol as well as atropine in the DG blocked the reinforcing effect of DG-LTP by stimulating the amygdala (BLA-R). These results are, in general terms, similar to those previously reported by our group showing that the

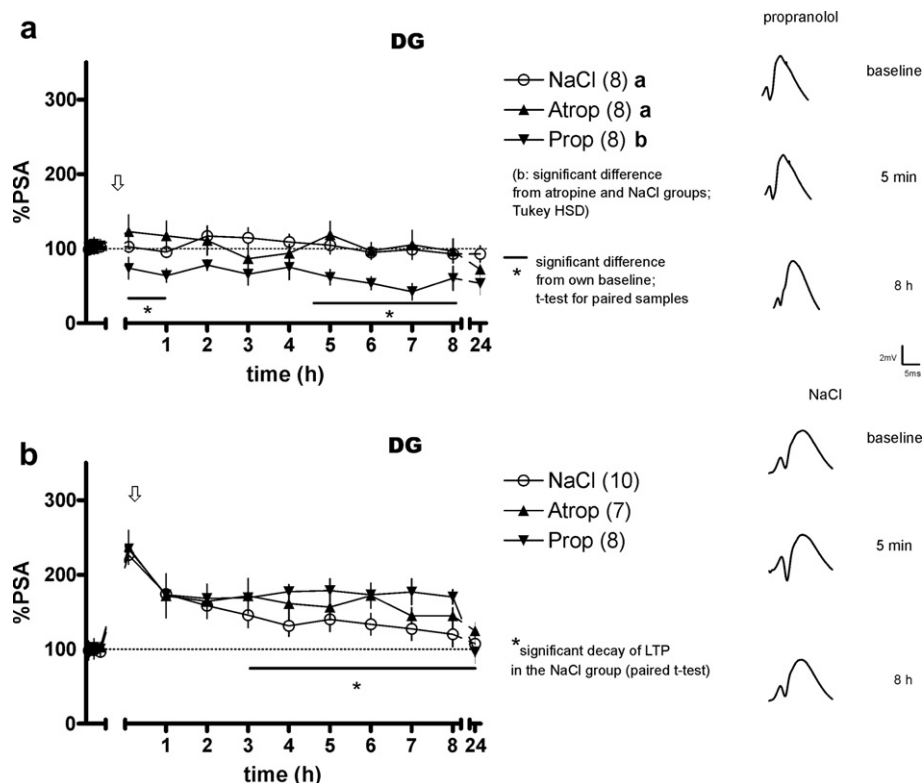


Fig. 3. Effects of topical application of drugs into the DG on baseline evoked potential (a) and on DG-LTP induced by PP stimulation (b). Mean values ($\pm$ SEM) of the population spike amplitude (% PSA, relative to baseline) for the experimental groups are shown (the numbers in parentheses indicate the number of animals in each groups). Asterisk indicates significant differences between groups in the post hoc test (Tukey HSD, $p < .05$). Representative traces from one animal in selected groups, before (baseline) and after E-LTP induction (5 min and 8 h) are shown in the right part of the figure.

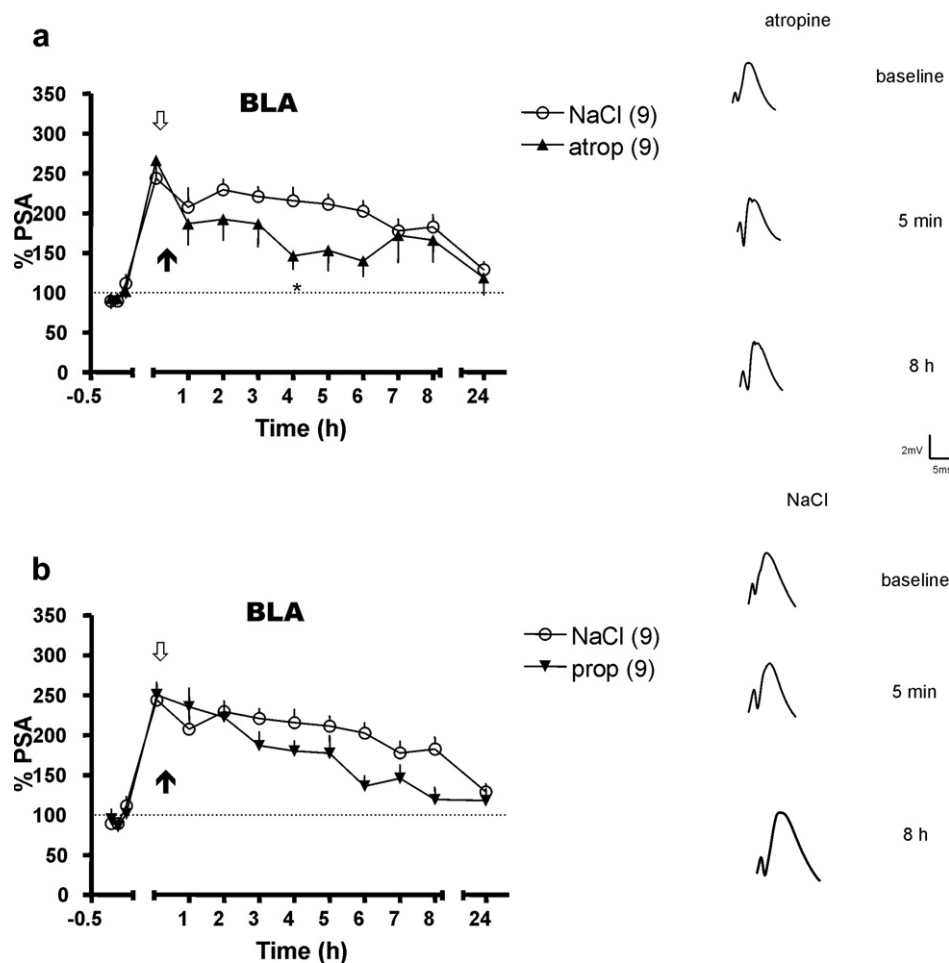


Fig. 4. Effects of topical application of drugs into the BLA on BLA-R. Mean values ($\pm$ SEM) of the PSA (% PSA, relative to baseline) for the experimental groups are shown in (a) after atropine, and (b) propranolol application (the numbers in parentheses indicates the number of animals in each groups). LTP was induced in the DG by stimulation of the PP at $t = 0$, and reinforced by stimulation of the BLA 15 min later (black arrow). Substances were topically applied into the BLA between both stimuli ($t = 10$ min, open arrow). The asterisk under the line indicates significant differences between groups in the post hoc test (Tukey HSD, $p < .05$). Representative traces from one animal in selected groups, before (baseline) and after E-LTP induction (5 min and 8 h) are shown in the right part of the figure.

intraventricular administration of propranolol or atropine blocked the reinforcing effect of BLA-stimulation on E-LTP in the DG (Frey et al., 2001) and suggest that the antagonisms of both substances are due, at least in part, to a direct action within the DG. However, in the case of propranolol a much earlier effect is seen when the substance is applied directly into the DG. The different time points at which noradrenergic or cholinergic innervation of the DG interfere with BLA-R suggest that NA is also required to sustain an intermediate stage of LTP—linking E-LTP and L-LTP—while acetylcholine apparently acts on mechanisms involved only at later stages. This might also explain our previous results on the effect of the direct application of NA and ACh agonists on LTP reinforcement, showing that epinephrine was able to mimic BLA-R, while a muscarinic agonist (oxotremorine) was unable to produce a similar effect (Almaguer-Melian et al., 2005). The lack of effect of the muscarinic agonist may depend on the need of a previous activation of the noradrenergic projection to the DG. These results suggest the

participation of the LC and the MS (the origin of NA and ACh innervation to the DG) in BLA-R which is supported by the results of directly applying substances in both targets.

A functional LC seems to be necessary for BLA-R, as the inactivation of the LC with lidocaine abolishes the reinforcement of DG-LTP. This effect is mediated by cholinergic afferents because atropine at the LC provokes a similar blockade. Noradrenergic afferents do not seem to be involved considering the lack of an effect of propranolol. However, the participation of other afferents to the LC cannot be ruled out. Similar caution is required regarding the specificity of the effects when substances are applied to the LC. Considering the small size of this structure and the volume injected, the possibility of partial diffusion outside the target can not be excluded. However, in this case the substance would reach first the periLC region which contains the peripheral dendrites of the LC neurons (Luppi, Aston-Jones, Akaoka, Chouvet, & Jouvett, 1995) which is functionally a receptive zone for the LC.

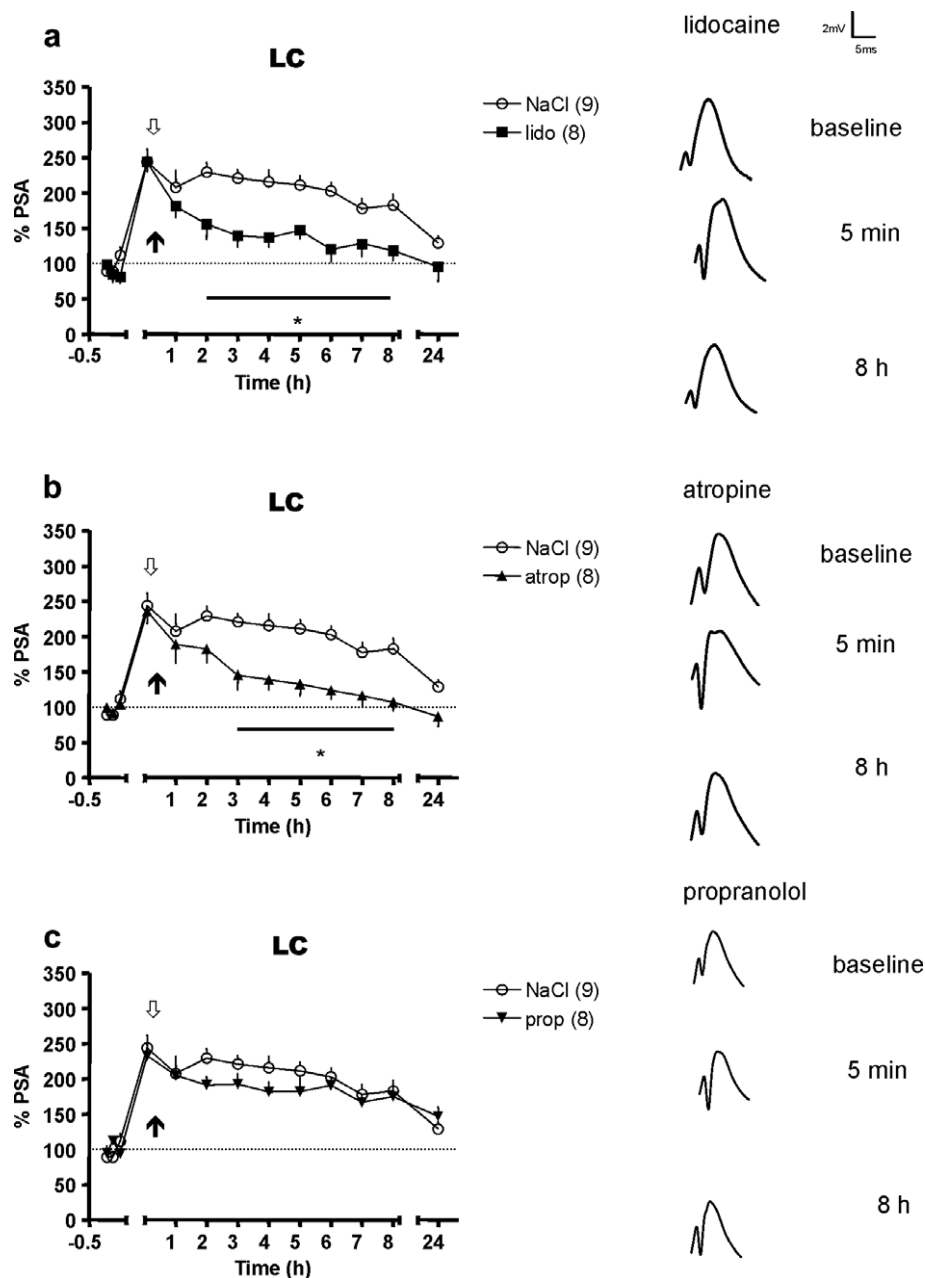


Fig. 5. Effects of topical application of drugs into the LC on BLA-R. Mean values ($\pm$ SEM) of the PSA (% PSA, relative to baseline) for the experimental groups are shown in (a) after lidocaine, (b) atropine, and (c) propranolol application (the numbers in parentheses indicate the number of animals in each group). LTP was induced in the DG by stimulation of the PP at $t = 0$, and reinforced by stimulation of the basolateral amygdala 15 min later (black arrow). Substances were topically applied into the LC between both stimuli ($t = 10$ min, open arrow). The asterisk under the line indicates significant differences between groups in the post hoc test (Tukey HSD, $p < .05$). Representative traces from one animal in selected groups, before (baseline) and after E-LTP induction (5 min and 8 h) are shown in the right part of the figure.

The LC is the main source of noradrenergic innervation to the forebrain and the hippocampal formation (Berridge & Waterhouse, 2003) but does not receive noradrenergic projections itself. This may explain the lack of an effect of propranolol applied to this structure on BLA-R. The LC receives afferents from the central amygdala mainly at the 'periLC'-region, but this projection does not seem to involve cholinergic fibers (Luppi et al., 1995). Cholinergic fibers to the LC have been described from neighbor brain stem nuclei, like the nucleus pedunculopontinus or the laterodorsal teg-

mental nucleus which form dendro-dendritic contacts within the LC and the 'periLC'-region (Luppi et al., 1995). Apparently, the strong blocking effect of disconnecting the LC with lidocaine, or the topical administration of cholinergic antagonists on BLA-induced DG-LTP reinforcement, is mediated by an indirect projection from the BLA to one of these putative regions from which cholinergic projection to the LC arises. The functional implications of the cholinergic projection on the activity of the LC are not well characterized yet, but our results indicate an activating, modulatory effect.

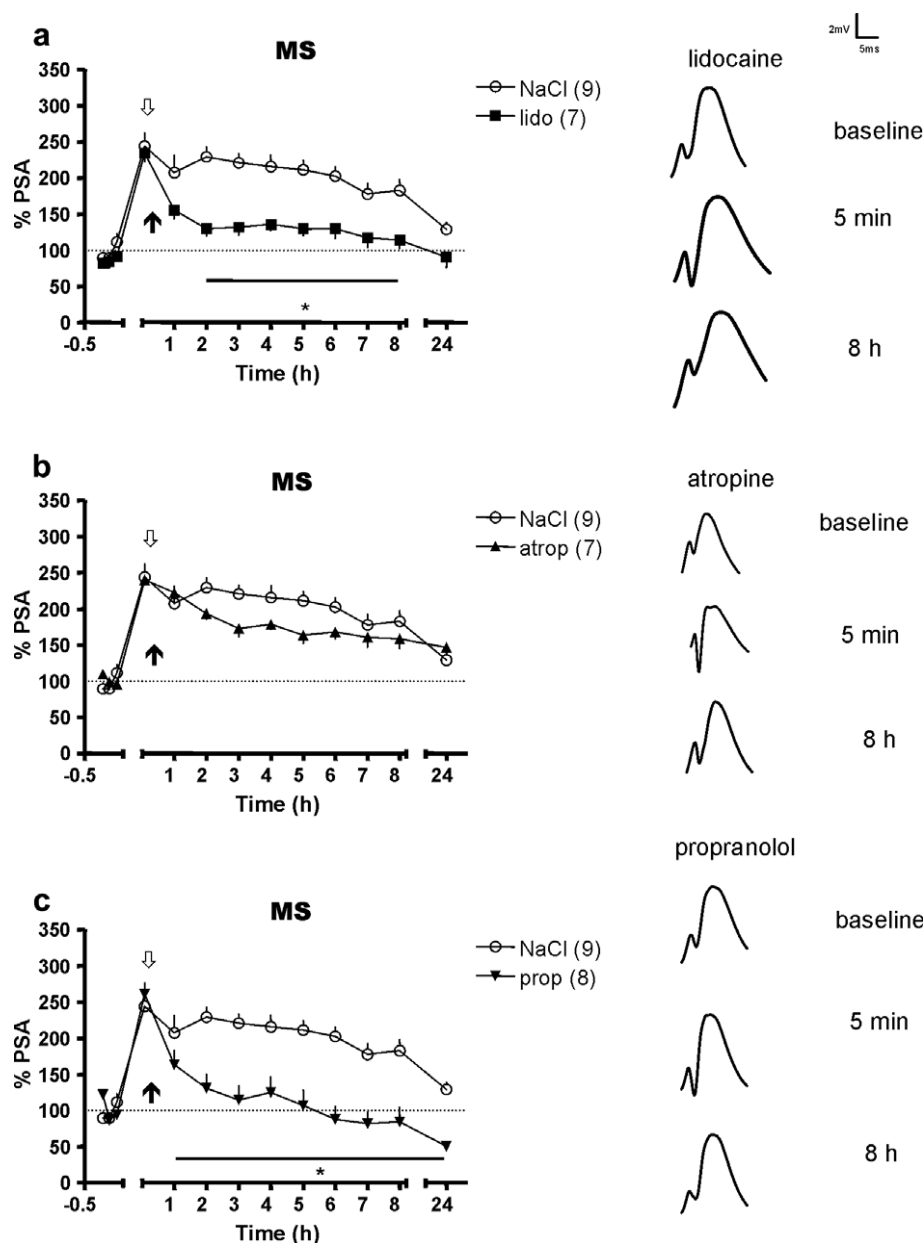


Fig. 6. Effects of topical application of drugs into the MS on BLA-R. Mean values ($\pm$ SEM) of the PSA (% PSA, relative to baseline) for the experimental groups are shown in (a) after lidocaine, (b) atropine, and (c) propranolol application (the numbers in parentheses indicates the number of animals in each groups). LTP was induced in the DG by stimulation of the PP at $t = 0$, and reinforced by stimulation of the BLA 15 min later (black arrow). Drugs were topically applied into the MS between both stimuli ($t = 10$ minutes, open arrow). The asterisk under the line indicates significant differences between groups in the post hoc test (Tukey HSD, $p < .05$). Representative traces from one animal in selected groups, before (baseline) and after E-LTP induction (5 min and 8 h) are shown in the right part of the figure.

Similarly, a functional MS at the time of BLA stimulation, appears to be necessary to obtain BLA-R; if the results after topical inactivation with lidocaine are considered. However, on this structure, the effect seems to be mediated by noradrenergic afferents, and not by cholinergic fibers, because propranolol at this site completely abolished BLA-R, while atropine was without effect. The LC sends a strong noradrenergic projection to the MS which forms synapses with cholinergic neurons in the basal forebrain (Zaborszky, Cullinan, & Luine, 1993) which project to the cortex and the hippocampal formation.

The topical application of drugs at the BLA showed only moderate effects. This might be a consequence of the direct electrical stimulation of that region which may overcome any postsynaptic blockade due to the one drug or the other. However, the late effect observed in the atropine treated group, suggests that some tonic BLA activity (probably initiated by cholinergic afferents) might be required during the poststimulus period.

Our results can be interpreted within a functional frame in which the activation of the BLA reinforces LTP in the DG by an indirect cholinergic activation of the LC, which

sends a direct noradrenergic projection to the DG, important to sustain the potentiated status in the transition between E-LTP and L-LTP. Additionally, the LC projection to the MS activates the cholinergic projection to the hippocampal formation whose activity seems responsible for L-LTP consolidation. There could be a direct activation of the MS via a putative excitatory projection from the BLA.

Regarding the cholinergic system, it has been extensively documented that the activation of the septal cholinergic projection to the hippocampal formation plays a critical role in memory consolidation (Balse et al., 1999; Blokland, 1995; Gasbarri, Introini-Collison, Packard, Pacitti, & McGaugh, 1993; Izquierdo et al., 1993; McGaugh & Cahill, 1997; Ramirez Lugo, Miranda, Escobar, & Bermudez-Rattoni, 2003). This cholinergic projection to the hippocampus and the DG modulates also LTP (Abe, Nakata, Mizutani, & Saito, 1994; Adams, Winterer, & Muller, 2004; Auerbach & Segal, 1994; Bergado, Moreno, & Nuñez, 1996; Blitzler, Gil, & Landau, 1990; Jerusalinsky, Kornisiuk, & Izquierdo, 1997; Leung, Shen, Rajakumar, & Ma, 2003; Markevich, Scorsa, Dawe, & Stephenson, 1997; Shinoo, Matsui, Taketo, & Manabe, 2005).

The MS, and its cholinergic projection play a pacemaker role in the induction of a rhythmic and synchronic neural activity characterized in EEG recordings as theta rhythm (Bland, Oddie, & Colom, 1999; Monmaur, Ayadi, & Breton, 1993; Vinogradova, Brazhnik, Stafekhina, & Kichigina, 1995; Vinogradova, Kitchigina, & Zenchenko, 1998). Hippocampal theta activity have been related to a variety of important functional states like REM sleep, exploration and attentive wakefulness (Bassant, Apartis, Jazat-Poinde-sous, Wiley, & Lamour, 1995; Bland & Bland, 1986; Bland, Seto, Sinclair, & Fraser, 1984; Vinogradova et al., 1995, 1998), as well as spatial memory (O'Keefe, 1993).

The aminergic-cholinergic synapses may be part of an extended neural system (summarized in Fig. 7) involved in the affective modulation of plasticity (alternative or complementary routes can also be implied: e.g. there is a direct—probably excitatory—input from the BLA to the basal forebrain (Zaborszky et al., 1993).

On the other hand, a noradrenergic projection from the LC reaches directly the DG, where it inhibits feed-forward inhibitory interneurons (Brown, Walling, Milway, & Harley, 2005) and may be responsible of some forms of potentiation described in the DG after LC activation (Kitchigina, Vankov, Harley, & Sara, 1997; Klukowski & Harley, 1994; Walling & Harley, 2004) or noradrenalin administration (Dahl & Sarvey, 1989; Izumi & Zorumski, 1999). This disinhibitory action of the noradrenergic projection may be related to the very rapid depotentiation that we have observed after propranolol injection in the DG in animals with a combined LTP/BLA-R protocol. Such an effect can also explain the depression in the evoked potentials at the DG after local propranolol injection.

Such a neural system would also make sense in physiological conditions. Sensory information is distributed from

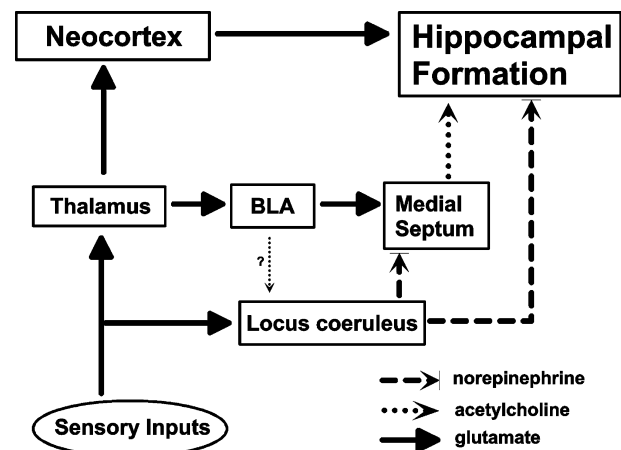


Fig. 7. Functional schemata of the putative neural system and neurotransmitters involved in the reinforcing effect of stimulating the BLA on LTP at the dentate gyrus. Explanation in the text.

the thalamus to the cortex and the amygdala. From higher order cortical fields the signal reaches the entorhinal cortex from where it is transferred to the DG and stored temporarily by mechanisms of short-term synaptic plasticity (i.e., E-LTP). The same information is processed in parallel by the limbic system (represented by the BLA) that via the neural system previously described, reinforces the storing mechanisms (modulation) by converting the E-LTP into a L-LTP. Note that parts of this system can be activated independently, under different circumstances. For instance, a strong sensory signal could activate the LC, as a part of an attentional system (Berridge & Waterhouse, 2003), which could have a similar effect on the plastic changes at synapses relevant for memory storage. We are aware that the circuit components presented here are likely to be incomplete. It has been described that the hippocampal theta-driving system includes other nuclei like the nucleus raphe in the brain stem, and the supramammillary nucleus of the hypothalamus (Borhegyi & Freund, 1998; Vertes, 2005; Vertes & Kocsis, 1997). Our own unpublished work suggests that the supramammillary nucleus is part of the reinforcing system in the DG, i.e. stimulation of this structure is also able to reinforce early- into late-LTP in the DG if it was stimulated 15 min after early-LTP-induction.

Finally, the reinforcing effects of affective factors and structures on neural plasticity processes may not be restricted to LTP in the dentate gyrus. There are reports that demonstrate an essential role for the cholinergic projection from the basal forebrain in cortical plasticity in sensory (Kilgard & Merzenich, 1998; Mercado, Bao, Orduna, Gluck, & Merzenich, 2001) and motor areas (Conner, Culberson, Packowski, Chiba, & Tuszynsky, 2003). We have recently demonstrated that stimulation of the BLA accelerates the plastic changes at the motor cortex, involved in the acquisition of a motor skill (Bergado et al., 2006). These might imply that the neural system proposed to explain the reinforcement of LTP at the dentate gyrus, could act on other forms of plasticity and

widespread cortical areas. Further studies are required to confirm the putative system advanced here; as well as to complete the list of structures and transmitters involved in the affective modulation of neural plasticity.

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References

- Abe, K., Nakata, A., Mizutani, A., & Saito, H. (1994). Facilitatory but nonessential role of the muscarinic cholinergic system in the generation of long-term potentiation of population spikes in the dentate gyrus *in vivo*. *Neuropharmacology*, 33, 847–852.
- Adams, S. V., Winterer, J., & Muller, W. (2004). Muscarinic signaling is required for spike-pairing induction of long-term potentiation at rat Schaffer collateral-CA1 synapses. *Hippocampus*, 14, 413–416.
- Almaguer-Melian, W., Capdevila, V., Ramirez, M., Vallejo, A., Rosillo-Marti, J. C., & Bergado-Rosado, J. (2005). Post-training stimulation of the basolateral amygdala improves spatial learning in rats with lesion of the fimbria-fornix. *Restorative Neurology and Neuroscience*, 23, 43–50.
- Almaguer-Melian, W., Martínez-Martí, L., Frey, J. U., & Bergado, J. A. (2003). The amygdala is part of the behavioral reinforcement system modulating long-term potentiation in rat hippocampus. *Neuroscience*, 119, 319–322.
- Almaguer-Melian, W., Rojas-Reyes, Y., Alvaré, A., Rosillo, J. C., Frey, J. U., & Bergado, J. A. (2005). Long-term potentiation in the dentate gyrus in freely moving rats is reinforced by intraventricular application of norepinephrine, but not oxotremorine. *Neurobiology of Learning and Memory*, 83, 72–78.
- Auerbach, J. M., & Segal, M. (1994). A novel cholinergic induction of long-term potentiation in rat hippocampus. *Journal of Neurophysiology*, 72, 2034–2040.
- Balse, E., Lazarus, C., Kelche, C., Jeltsch, H., Jackisch, R., & Cassel, J. C. (1999). Intrahippocampal grafts containing cholinergic and serotonergic fetal neurons ameliorate spatial reference but not working memory in rats with fimbria-fornix/cingular bundle lesions. *Brain Research Bulletin*, 49, 263–272.
- Bassant, M. H., Apartis, E., Jazat-Poindessous, F. R., Wiley, R. G., & Lamour, Y. A. (1995). Selective immunolesion of the basal forebrain cholinergic neurons: Effects on hippocampal activity during sleep and wakefulness in the rat. *Neurodegeneration*, 4, 61–70.
- Bergado, J. A., Almaguer-Melian, W., Kostenko, S., Frey, S., & Frey, J. U. (2003). Behavioral reinforcement of long-term potentiation in rat dentate gyrus *in vivo* is protein synthesis-dependent. *Neuroscience Letters*, 351, 56–58.
- Bergado, J. A., Moreno, H., & Nuñez, N. (1996). Fimbria-fornix lesion impairs long-term potentiation in the dentate gyrus of the rat. *Biological Research*, 29, 197–202.
- Bergado, J. A., Rojas, Y., Capdevila, V., Gonzalez, O., & Almaguer-Melian, W. (2006). The stimulation of the basolateral amygdala improves the acquisition of a motor skill. *Restorative Neurology and Neuroscience*, 24, 115–121.
- Berridge, C. W., & Waterhouse, B. D. (2003). The locus coeruleus-noradrenergic system: Modulation of behavioral state and state-dependent cognitive processes. *Brain Research Reviews*, 42, 33–84.
- Bland, B. H., Oddie, S. D., & Colom, L. V. (1999). Mechanisms of neural synchrony in the septohippocampal pathways underlying hippocampal theta generation. *Journal of Neuroscience*, 19, 3223–3237.
- Bland, B. H., Seto, M. G., Sinclair, B. R., & Fraser, S. M. (1984). The pharmacology of hippocampal theta cells: Evidence that the sensory processing correlate is cholinergic. *Brain Research*, 299, 121–131.
- Bland, S. K., & Bland, B. H. (1986). Medial septal modulation of hippocampal theta cell discharges. *Brain Research*, 375, 102–116.
- Blitzer, R. D., Gil, O., & Landau, E. M. (1990). Cholinergic stimulation enhances long-term potentiation in the CA1 region of rat hippocampus. *Neuroscience Letters*, 119, 207–210.
- Blokland, A. (1995). Acetylcholine A neurotransmitter for learning and memory? *Brain Research Reviews*, 21, 285–300.
- Borhegyi, Z., & Freund, T. F. (1998). Dual projection from the medial septum to the supramammillary nucleus in the rat. *Brain Research Bulletin*, 46, 453–459.
- Brown, R. A. M., Walling, S. G., Milway, J. S., & Harley, C. W. (2005). Locus coeruleus activation suppresses feedforward interneurons and reduces b-g electroencephalogram frequencies while it enhances q frequencies in rat dentate gyrus. *Journal of Neuroscience*, 25, 1985–1991.
- Carmichael, S. T. (2003). Plasticity of cortical projections after stroke. *Neuroscientist*, 9, 64–75.
- Conner, J. M., Culbertson, A., Packowski, C., Chiba, A., & Tuszyński, M. H. (2003). Lesions of the basal forebrain cholinergic system impair task acquisition and abolish cortical plasticity associated with motor skill learning. *Neuron*, 38, 819–829.
- Dahl, D., & Sarvey, J. (1989). Norepinephrine induces pathway-specific long-term potentiation and depression in the hippocampal dentate gyrus. *Proceedings of the National Academy of Sciences of the United States of America*, 86, 4776–4780.
- Dam, H. (2001). Depression in stroke patients 7 years following stroke. *Acta Psychiatrica Scandinavica*, 103, 287–293.
- Frey, S., Bergado, J. A., & Frey, J. U. (2003). Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum. *Neuroscience*, 118, 1055–1062.
- Frey, S., Bergado-Rosado, J., Seidenbecher, T., Pape, H. C., & Frey, J. U. (2001). Reinforcement of early long-term potentiation (early-LTP) in dentate gyrus by stimulation of the basolateral amygdala: Heterosynaptic induction mechanisms of late-LTP. *Journal of Neuroscience*, 21, 3697–3703.
- Gasbarri, A., Introini-Collison, I. B., Packard, M. G., Pacitti, C., & McGaugh, J. L. (1993). Interaction of cholinergic-dopaminergic systems in the regulation of memory storage in aversively motivated learning tasks. *Brain Research*, 627, 72–78.
- Izquierdo, I., Medina, J. H., Bianchin, M., Walz, R., Zanatta, M. S., Da Silva, R. C., et al. (1993). Memory processing by the limbic system: Role of specific neurotransmitter systems. *Behavioural Brain Research*, 58, 91–98.
- Izumi, Y., & Zorumski, C. F. (1999). Norepinephrine promotes long-term potentiation in the adult rat hippocampus *in vitro*. *Synapse*, 31, 196–202.
- Jerusalinsky, D., Kornisiuk, E., & Izquierdo, I. (1997). Cholinergic neurotransmission and synaptic plasticity concerning memory processing. *Neurochemical Research*, 22, 507–515.
- Kilgard, M. P., & Merzenich, M. M. (1998). Cortical map reorganization enabled by nucleus basalis activity. *Science*, 279, 1714–1718.
- Kitchigina, V., Vankov, A., Harley, C., & Sara, S. J. (1997). Novelty-elicited, noradrenaline-dependent enhancement of excitability in the dentate gyrus. *European Journal of Neuroscience*, 9, 41–47.
- Klukowski, G., & Harley, C. W. (1994). Locus coeruleus activation induces perforant path-evoked population spike potentiation in the dentate gyrus of awake rat. *Experimental Brain Research*, 102, 165–170.
- Leung, L. S., Shen, B., Rajakumar, N., & Ma, J. (2003). Cholinergic activity enhances hippocampal long-term potentiation in CA1 during walking in rats. *Journal of Neuroscience*, 23, 9297–9304.
- Luppi, P. H., Aston-Jones, G., Akaoka, H., Chouvet, G., & Jouvet, M. (1995). Afferent projections to the rat locus coeruleus demonstrated by retrograde and anterograde tracing with cholera-toxin B subunit and *Phaseolus vulgaris* leucoagglutinin. *Neuroscience*, 65, 119–160.

- Markevich, V., Scorsia, A. M., Dawe, G. S., & Stephenson, J. D. (1997). Cholinergic facilitation and inhibition of long-term potentiation of CA1 in the urethane-anaesthetized rats. *Brain Research*, 754, 95–102.
- Matthies, H. (1989). In search of cellular mechanisms of memory. *Progress in Neurobiology*, 32, 277–349.
- McGaugh, J. L., & Cahill, L. (1997). Interaction of neuromodulatory systems in modulating memory storage. *Behavioural Brain Research*, 83, 31–38.
- McIntyre, C. K., Power, A. E., Roozendaal, B., & McGaugh, J. L. (2003). Role of the basolateral amygdala in memory consolidation. *Annals of the New York Academy of Sciences*, 985, 273–293.
- Mercado, E., Bao, S., Orduna, I., Gluck, M. A., & Merzenich, M. M. (2001). Basal forebrain stimulation changes cortical sensitivities to complex sound. *Neuroreport*, 20(12), 2283–2287.
- Monmaur, P., Ayadi, K., & Breton, P. (1993). Hippocampal EEG responses induced by carbachol and atropine infusions into the septum and the hippocampus in the urethane-anaesthetized rat. *Brain Research*, 631, 317–324.
- Narushima, K., Chan, K. L., Kosier, J. T., & Robinson, R. G. (2003). Does cognitive recovery after treatment of poststroke depression last? A 2-year follow-up of cognitive function associated with poststroke depression. *American Journal of Psychiatry*, 160, 1157–1162.
- O'Keefe, J. (1993). Hippocampus, theta, and spatial memory. *Current Opinion in Neurobiology*, 3, 917–924.
- Ramirez Lugo, L., Miranda, I., Escobar, L., & Bermudez-Rattoni, F. (2003). The role of cortical cholinergic pre- and post-synaptic receptors in taste memory formation. *Neurobiology of Learning and Memory*, 79, 184–193.
- Richter-Levin, G. (2004). The amygdala, the hippocampus, and emotional modulation of memory. *Neuroscientist*, 10, 31–39.
- Seidenbecher, T., Reyman, K. G., & Balschun, D. (1997). A post-tetanic time window for the reinforcement of long-term potentiation by appetitive and aversive stimuli. *Proceedings of the National Academy of Sciences of the United States of America*, 94, 1494–1499.
- Shinoe, T., Matsui, M., Taketo, M. M., & Manabe, T. (2005). Modulation of synaptic plasticity by physiological activation of M1 muscarinic acetylcholine receptors in the mouse hippocampus. *Journal of Neuroscience*, 25, 11194–11200.
- Straube, T., Korz, V., & Frey, J. U. (2003). Bidirectional modulation of long-term potentiation by novelty-exploration in rat dentate gyrus. *Neuroscience Letters*, 344, 5–8.
- Uzakov, S., Frey, J. U., & Korz, V. (2005). Reinforcement of rat hippocampal LTP by holeboard training. *Learning and Memory*, 12, 165–171.
- Vertes, R. P. (2005). Hippocampal theta rhythm: A tag for short-term memory. *Hippocampus*, 15, 923–935.
- Vertes, R. P., & Kocsis, B. (1997). Brainstem–diencephalo–septohippocampal systems controlling the theta rhythm of the hippocampus. *Neuroscience*, 81, 893–926.
- Vinogradova, O. S., Brazhnik, E. S., Stafekhina, V. S., & Kichigina, V. F. (1995). Modulation of septal influences on hippocampal neurons by cholinergic substances. *Neuroscience and Behavioral Physiology*, 25, 453–461.
- Vinogradova, O. S., Kitchigina, V. F., & Zenchenko, C. I. (1998). Pacemaker neurons of the forebrain medial septal area and theta rhythm of the hippocampus. *Membrane Cell Biology*, 11, 715–725.
- Vizi, E. S., & Kiss, J. P. (1998). Neurochemistry and pharmacology of the major hippocampal transmitter systems: Synaptic and nonsynaptic interaction. *Hippocampus*, 8, 566–607.
- Walling, S. G., & Harley, C. W. (2004). Locus ceruleus activation initiates delayed synaptic potentiation of perforant path input to the dentate gyrus in awake rats: A novel beta-adrenergic- and protein synthesis-dependent mammalian plasticity mechanism. *Journal of Neuroscience*, 24, 598–604.
- Zaborszky, L., Cullinan, W. E., & Luine, V. N. (1993). Catecholaminergic–cholinergic interaction in the basal forebrain. *Progress in Brain Research*, 98, 31–49.

CONSIDERACIONES FINALES

El esquema que se propone al final del artículo sirve también para interpretar las deficiencias detectadas en los estudios con animales viejos. El deterioro funcional del sistema cerebral colinérgico y noradrenérgico han sido ampliamente documentados en la literatura científica. Estos sistemas se encuentran en el centro mismo de la interacción entre neuroplasticidad y afectividad y su deterioro con la edad, conduciría a un mayor deterioro de la memoria al dificultar el reforzamiento de los procesos neuroplásticos que le sirven de base.

El principio de universalidad que estamos proponiendo tiene un cierto carácter axiomático, en tanto que resulta imposible diseñar un experimento que lo niegue o confirme más allá de toda duda. Para confirmar que el reforzamiento de procesos neuroplásticos inducido por la estimulación de la amígdala, tiene verdaderamente un carácter universal habría que realizar experimentos veritativos explorando todos los tipos de procesos neuroplásticos, en todas las estructuras cerebrales, en todas las especies animales. Tal tarea está más allá de la capacidad de cualquier grupo y dista mucho del nivel de evidencia aquí presentado. Pero el hecho de que en los dos estudios realizados para comprobar la universalidad hayan ofrecido evidencias positivas es importante porque bastaría con solo una evidencia negativa para negarlo. En tanto la evidencia acumulada afiance la certeza, o que aparezcan resultados que la limiten, proponemos la tesis de que *los factores afectivos modulan los procesos de neuroplasticidad en todas sus formas de expresión, en todas las regiones del encéfalo y en todas las especies con un cerebro desarrollado. Los mecanismos se ajustan al mecanismo común de control celular mediante el cuál se regula el metabolismo macromolecular neuronal y los procesos de*

crecimiento o cambios funcionales que de ellos dependen. En los mamíferos los sistemas colinérgico de origen septal y noradrenérgico de origen en el locus coeruleus, activados por la amígdala forman parte fundamental del mecanismo fisiológico de modulación afectiva de procesos de neuroplasticidad.

De este principio se derivan algunos corolarios que pueden tener utilidad práctica como guía de futuras investigaciones:

- Los agentes que estimulan, modulan o refuerzan un proceso neuroplástico (p. ej. la memoria) tienen, con gran probabilidad, el mismo efecto sobre otras formas de neuroplasticidad (p.ej. el crecimiento neurítico).
- Los agentes que estimulan, modulan o refuerzan un proceso neuroplástico en una etapa del desarrollo tienen, con gran probabilidad, el mismo efecto en otras etapas del desarrollo.
- Los agentes que estimulan, modulan o refuerzan un proceso neuroplástico en una especie animal tienen, con gran probabilidad, el mismo efecto en otras especies.

Reiteramos que estos corolarios son solo una guía para la formulación de hipótesis experimentalmente verificables. Evidentemente, cualquier intento de aplicación concreta de algún presunto agente modulador que pueda inferirse basado en estos corolarios pasa, sin excepciones, por la inaplazable comprobación experimental previa.

En la actualidad las propiedades neuroplásticas del Sistema Nervioso constituyen la mejor herramienta de que dispone la Neurología Restaurativa para lograr detener o enlentecer, recuperar o compensar funciones nerviosas afectadas por enfermedad o trauma. Comprender sus relaciones con los factores afectivos y los mecanismos de que depende para utilizarlos sabiamente en el logro de mejores resultados y tratamientos es

una aspiración. Haber contribuido en algo a ese noble empeño es nuestra mejor y más valiosa recompensa.

CONCLUSIONES

1. El reforzamiento conductual actúa como evento modulador de la plasticidad sináptica mediante la activación de los mecanismos celulares de biosíntesis que aportan las proteínas requeridas por el proceso neuroplástico para su consolidación. Esta acción moduladora depende de la actividad de sistemas de neurotransmisión noradrenérgica y colinérgica.
2. La amígdala es parte del sistema neural implicado en el reforzamiento de procesos neuroplásticos por factores afectivos.
3. La proyección septo-hipocampal y la vía fimbria-fornix son mediadores necesarios en el reforzamiento de procesos neuroplásticos por factores afectivos.
4. Es posible mimetizar el efecto de reforzamiento afectivo mediante la administración sistémica de agonistas de la neurotransmisión noradrenérgica en dosis adecuadas.
5. El envejecimiento afecta la capacidad de reforzamiento de procesos neuroplásticos por factores afectivos que pudiera estar causada por la reducción en la capacidad de movilizar los neurotransmisores requeridos.
6. Los mecanismos de reforzamiento de procesos de neuroplasticidad por factores afectivos no son exclusivos de una forma particular de expresión de la neuroplasticidad, ni de una región específica del cerebro, ni del sujeto sano y parecen representar un mecanismo y universal mediante el cuál el valor afectivo de una experiencia modula el desarrollo y persistencia de fenómenos neuroplásticos inducidos por esa misma experiencia.

RECOMENDACIONES

1. El sistema neural propuesto para explicar la acción moduladora de los factores afectivos sobre los procesos de neuroplasticidad se basa, fundamentalmente en las evidencias obtenidas por nuestro grupo de trabajo en los experimentos que se recogen en este documento. Sin embargo, evidencias de otros autores sugieren que otras estructuras pudieran participar también en estos mecanismos. Entre ellas se destacan dos: el núcleo *accumbens septi* y el núcleo del rafe medial que están siendo estudiadas en experimentos en curso o concluidos recientemente.
2. El carácter universal que atribuimos a los mecanismos propuestos justifican la búsqueda de equivalentes funcionales en seres humanos y la posible extensión de estos resultados al tratamiento neurorestaurativo en uso en nuestra institución. Un trabajo, concluido recientemente, muestra que es posible lograr activación de la amígdala mediante técnicas de estimulación acupuntural. Eso posibilita el diseño de experimentos en humanos sanos para obtener evidencias del efecto modulador de la estimulación de la amígdala sobre procesos de neuroplasticidad y, de comprobarse, el diseño de ensayos terapéuticos para verificar su efecto en pacientes con afecciones neurológicas crónicas.

DIVULGACIÓN DE LOS RESULTADOS

RECOGIDOS EN LA TESIS

Eventos científicos

Los resultados recogidos en el presente documento de tesis han sido presentados en **eventos científicos** nacionales e internacionales celebrados en Cuba y en el extranjero. Entre ellos mencionamos los siguientes:

Sexto Congreso Latinoamericano Neuropsicología. Varadero, Octubre 17-20 de 1999.

- Plasticidad sináptica y envejecimiento.

- Sistema Límbico y memoria. Nuevas evidencias a la luz de la plasticidad sináptica.

USA-CUBA Neurometing. La Habana, Octubre 20-23 de 1999.

- Amygdala-hippocampal interaction in synaptic plasticity.

Xth Magdeburg International Neurobiological Symposium: Mechanisms of Learning and Memory. Magdeburgo 16-19 Septiembre 2000

- Aging impairs amygdala-hippocampus interactions in LTP.

2nd Experimental Neuroscience Meeting. La Habana Marzo 2001

- Heterosynaptic interactions on late phases of long-term potentiation in the hippocampus: Mechanisms and impairments by aging. Presentación oral

Joint Meeting 18th Biennial Meeting of the International Society for Neurochemistry
and 32nd Annual meeting of the American Society for Neurochemistry, Buenos Aires
26-31 de Agosto, 2001

-Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP.

Poster

Annual Meeting of the Society for Neuroscience (USA). San Diego. Noviembre 2001

- Heterosynaptic modulation of late phases of long-term potentiation in the
hippocampus by limbic structures and their impairment by aging. Bergado, J.;

Almaguer, W.; Frey, S*.; Frey, J.U*. Poster

Curso de Conferencias de la IBRO, (Organizado por la Visiting Lecture Team de
IBRO), La Habana, Junio 20-28 de 2001

- Synaptic plasticity, aging and memory.

6to Congreso Mundial de la IBRO, Praga, Julio 10-15, 2003

- Effects of aging on limbic plasticity and affective modulation of LTP. Almaguer,

W., Jas, J., Ravelo, A., Alvaré, J.A., Bergado, J., Frey, S., Frey, J.U.

I Simposium Internacional de Rehabilitación Neurológica: Neurorehabilitación 2003

- Influencia de los factores emocionales y motivacionales sobre los procesos

neuroplásticos. Bergado, J.A. Conferencia

Cuba US Workshop on Biological Psychiatry. La Habana 15-20 de enero 2004

- Motivation cognition and synaptic plasticity. Conferencia

IV Foro Iberoamericano sobre Demencias y Enfermedad de Alzheimer, La Habana 25 de marzo 2004

- Plasticidad sináptica y envejecimiento. Nuevas bases para entender la interacción entre afectividad y memoria

XXXIX Congreso Nacional de la Asociación Colombiana de Ciencias Biológicas.

Ibagué 11-15 octubre de 2004

- Memoria, motivación y plasticidad sináptica. Bergado, J.A; Almaguer-Melán, W.; Frey, S.; Frey, J.U. (Conferencia)

XI Magdeburg International Neurobiological Symposium, Magdeburgo 28 Mayo al 1ro de Junio, 2005

- Modulation of cellular memory consolidation in young and aged animals.

Conferencia Invitada

9th International Conference on Cognitive Neurosciences (ICON). La Habana, Sept. 6-9, 2005

- Affective modulation of synaptic plasticity. Implications for cognitive functions. Conferencia.

Publicaciones

Aunque aparecen en el cuerpo de la tesis, relaciono a continuación y en el orden de su aparición los **artículos científicos** que recogen los resultados que aquí se presentan:

- Jas, J., Almaguer, W., Bergado, J.: **Lesioning the fimbria-fornix impairs basolateral amygdala induced reinforcement of LTP in the dentate gyrus** Brain Res., 861: 186-189, 2000. (Factor de impacto FI: 2.296, Citado:
- Bergado, J., Almaguer, W.: **Mecanismos celulares de neuroplasticidad** Rev. Neurol. 31 (11) 1074-1095, 2000. (FI: 0.391, citado 9 veces)
- Frey, S., Bergado Rosado, J.; Seidenbecher, T., Pape, H-C, and Frey, J.U.: **Reinforcement of early LTP in dentate gyrus by specific stimulation of the basolateral amygdala: Heterosynaptic induction mechanisms of late LTP** J. Neurosci., 21, 3697-3703, 2001.(FI: 7.706, citado 83 veces)
- Bergado, J.A., Almaguer, W., Ravelo, J., Rosillo, J.C., Frey, J.U. **Behavioral reinforcement of long-term potentiation is impaired in aged rats with cognitive deficiencies** Neuroscience, 108(1):1-5,2001 (FI: 3.41, citado 6 veces)
- Almaguer, W., Estupiñán, B., Frey, J.U, Bergado, J.A. **Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP** Neurobiol. Aging, 23(2):319-324, 2002. (FI: 5.312, citado 11 veces)
- Bergado, J.A.; Almaguer-Melián, W.: **Aging and synaptic plasticity: a review** Neural Plast. 9 (4): 217-232, 2002. (FI: , citado 4 veces)
- Frey, S.; Bergado, J.A.; Frey, J.U. **Modulation of late phases of long-term potentiation in rat dentate gyrus by stimulation of the medial septum** Neuroscience, 118: 1055-1062, 2003 (FI: 3.41, citado 10 veces)

- Almaguer-Melian, W.; Martínez-Martí, L.; Frey, J.U.; Bergado, J.A. **The amygdala is part of the behavioral reinforcement system of long-term potentiation in rat hippocampus.** Neuroscience, 119 (2): 319-322, 2003 (FI: 3.41, citado 14 veces)
- Bergado, J.A.; Almaguer-Melian, W.; Kostenko, S.; Frey, S.; Frey, J.U: **Behavioral reinforcement of long-term potentiation in rat dentate gyrus in vivo is protein synthesis-dependent [rapid communication].** Neurosci. Lett., 351 (1) 56-58, 2003 (FI: 1.891, citado 8 veces)
- William Almaguer-Melian, Yeneissy Rojas-Reyes, Armando Alvare, Juan C. Rosillo, Julietta U. Frey and Jorge A. Bergado: **Long-term potentiation in the dentate gyrus in freely moving rats is reinforced by intraventricular application of norepinephrine, but not oxotremorine.** Neurobiol. Learn. Mem., 83: 72-78, 2005. (FI: 4.091, citado 9 veces)
- William Almaguer, Vladimir Capdevila, Magaly Ramírez, Araceli Vallejo, Juan C. Rosillo and Jorge A. Bergado: **Post-training stimulation of the basolateral amygdala improves spatial learning in rats with lesion of the fimbria-fornix.** Restor. Neurol. Neurosci., 23: 43-50, 2005. (FI: 1.825, citado 1 vez)
- W. Almaguer-Melian, R. Cruz-Aguado, C. de la Riva, K.M. Kendrick, J.U. Frey, J. Bergado: **Effect of LTP-reinforcing paradigms on neurotransmitter release in the dentate gyrus of young and aged rats.** Biochem. Biophys. Res. Comm., 327: 877-883, 2005. (FI: 3.0, citado 5 veces)

- Almaguer-Melian, W.; Rosillo, J.C.; Frey, J.U.; Bergado, J.A.: **Subcortical deafferentation impairs behavioral reinforcement of LTP in the dentate gyrus of freely moving rats.** Neuroscience, 2006, 138: 1083-1088 (FI: 3.41, citado 1 vez)
- Jorge A. Bergado, Yeneissy Rojas, Vladimir Capdevila, Odalys Gonzalez, William Almaguer-Melian: **The stimulation of the basolateral amygdala improves the acquisition of a motor skill.** Restor. Neurol. Neurosci. 2006, 24(2):115-121 (FI: 1.825)
- Jorge A. Bergado, Sabine Frey, Jeffrey López, William Almaguer and Julietta U. Frey: **Cholinergic afferents to the locus coeruleus and noradrenergic afferents to the medial septum mediate LTP-reinforcement in the dentate gyrus by stimulation of the amygdala** 2007 Neurobiol. Learn. Mem., 88: 331-341. (FI: 4.091)

Premios

Los trabajos que recoge la presente tesis han recibido las distinciones que a continuación se relacionan:

- Premio Anual de Salud 2000, MINSAP. Premio a nivel Nacional. Autor Principal del Trabajo: **Efecto de la desaferentación subcortical hipocampal sobre el reforzamiento de la LTP en el giro dentado por estimulación de la amígdala basolateral**
- Premio Anual de Salud 2003, MINSAP. Premio a nivel Nacional. Autor Principal del Trabajo: **El envejecimiento afecta la interacción amígdala-hipocampo implicada en la LTP hipocampal.**
- Premio Anual de Salud 2004, MINSAP. Premio al Nivel Nacional. Autor Principal del Trabajo: **Estudio sobre la participación de la amígdala en los procesos de reforzamiento conductual de la potenciación a largo plazo**
- Premio Anual de Salud 2005. La Tesis de Doctorado, de la cuál fui TUTOR: **Mecanismos de consolidación motivacional de la plasticidad sináptica duradera. Efectos del envejecimiento cerebral** de William Almaguer Melian recibió una de las tres MENCIONES que otorga el jurado del como Mejor Tesis Doctoral.
- Premio Anual de la Academia de Ciencias de Cuba 2005 por el trabajo: **Estudio de los mecanismos de reforzamiento de procesos neuroplásticos por factores afectivos.** Autor Principal
- Premio Anual de Salud 2006, MINSAP. Mención al Nivel Central al trabajo **Efecto de la administración de norepinefrina y oxotremorina sobre el mantenimiento de la potenciación sináptica de larga duración.** Coautor

- Premio Anual de Salud 2007, MINSAP: Premio al Nivel Central al trabajo

Universalidad de los efectos de la estimulación de la amígdala basolateral sobre los procesos de neuroplasticidad. Autor